

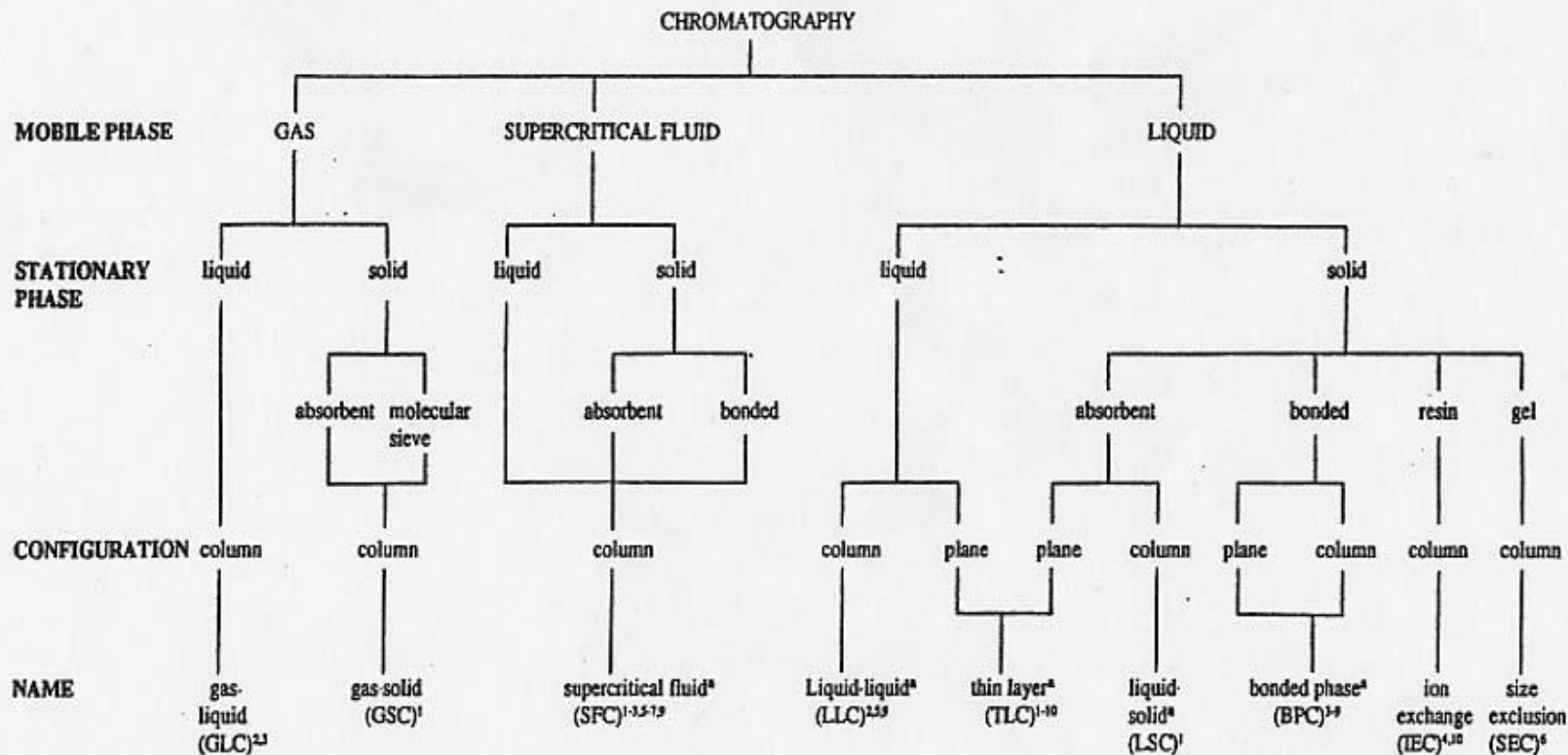


# CHEMISTRY 5570

## Advanced Analytical Chemistry Lecture 14



# Chromatography

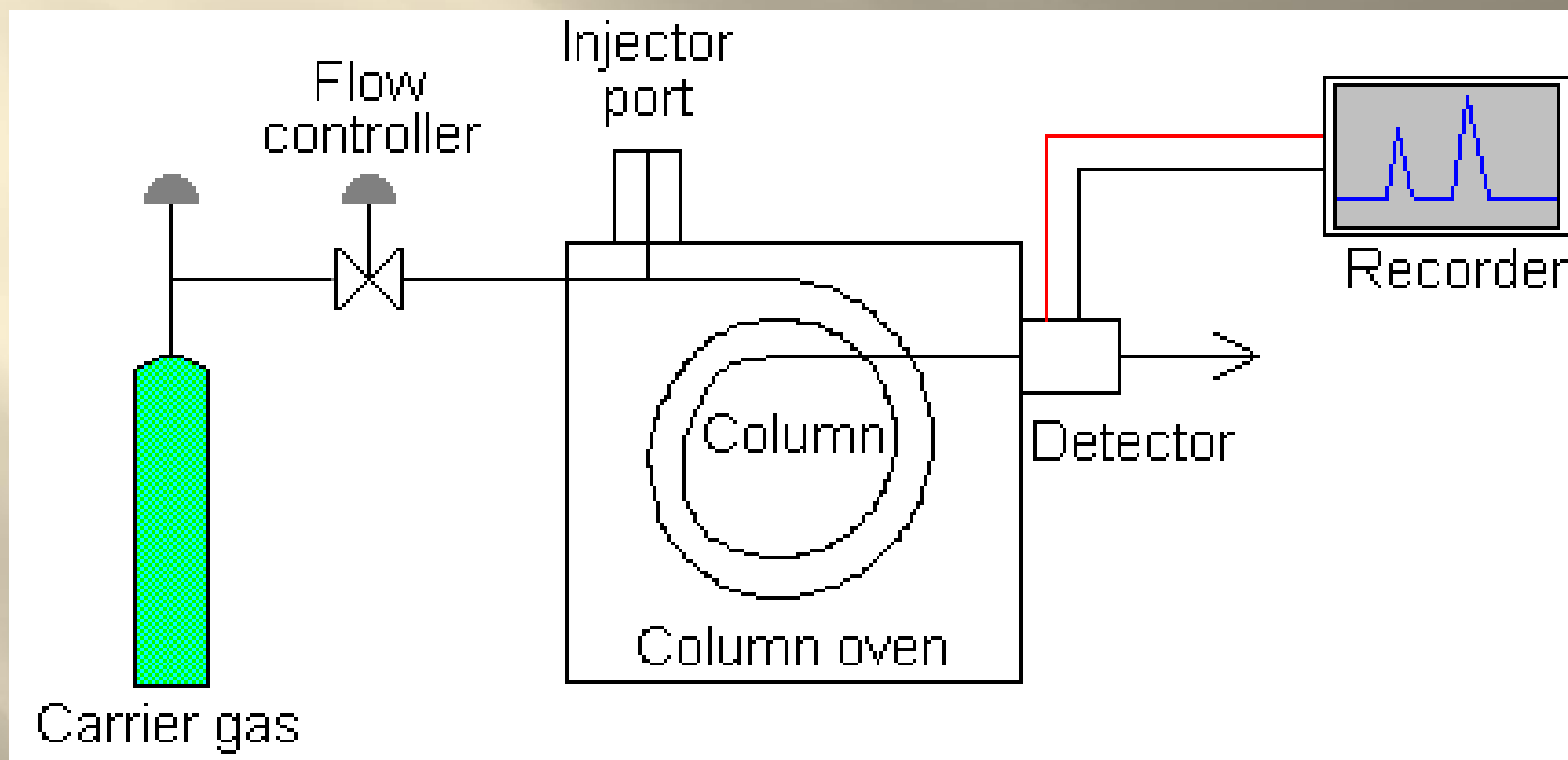


<sup>a</sup>For these techniques the combination of mobile and stationary phase can be varied to generate either a normal phase or reversed phase system. Mechanisms which have been exploited in the various techniques are identified as: <sup>1</sup>adsorption, <sup>2</sup>partition, <sup>3</sup>bonded phase, <sup>4</sup>ion exchange, <sup>5</sup>ion interaction, <sup>6</sup>size exclusion, <sup>7</sup>affinity, <sup>8</sup>micellar, <sup>9</sup>chelation, <sup>10</sup>ion exclusion.

Fig. 1.3. Classification of chromatographic systems.

# Chromatography

## Gas Chromatography – Columns



# Chromatography

## Gas Chromatography

Types of Columns

Packed Columns

Older type of column

Both solid and liquid stationary phase

Best column for preparatory GC and for use with thermal conductivity detectors

Sometimes used for very specific applications (low production volume less of an issue)

# Chromatography

## Gas Chromatography

Types of Columns

Open Tubular Columns

More modern columns

Much better analytical performance (large N values)

Most common in wall coated format (WCOT)

Variety of diameters (0.25 to 0.53 mm most common) allow high resolution vs. easier injection

Stationary phases are mainly bonded of varying amounts of polarity

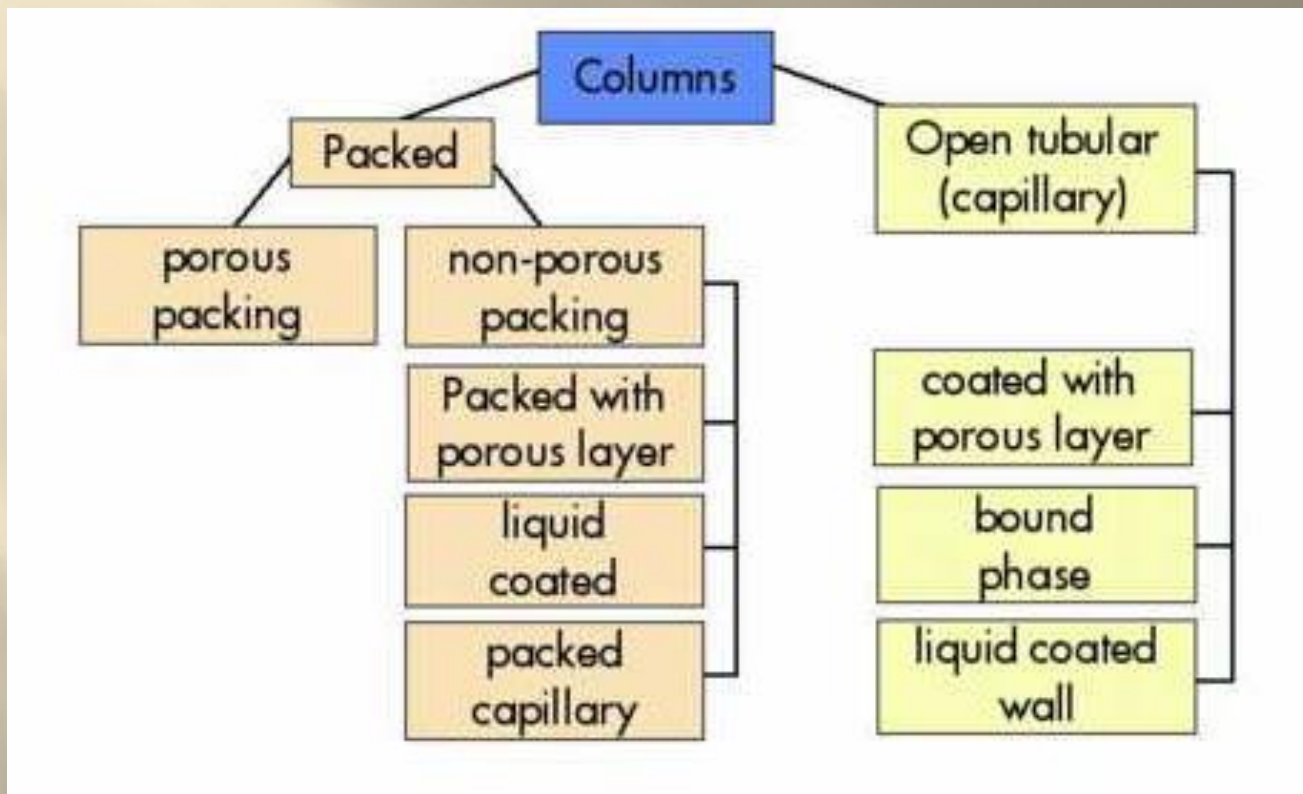
Good reliability

Disadvantages: harder to make and less capacity



# Chromatography

## Gas Chromatography



# Chromatography

## Gas Chromatography

### Columns

**Packed columns contain a finely divided, inert, solid support material (commonly based on diatomaceous earth) coated with liquid stationary phase. Most packed columns are 1.5 – 10 m in length and have an internal diameter of 2 – 4 mm.**

**Capillary columns have an internal diameter of a few tenths of a millimeter.**

**They can be one of two types; wall-coated open tubular (WCOT) or support-coated open tubular (SCOT).**

# Chromatography

## Gas Chromatography

### Columns

**Wall-coated columns consist of a capillary tube whose walls are coated with liquid stationary phase.**

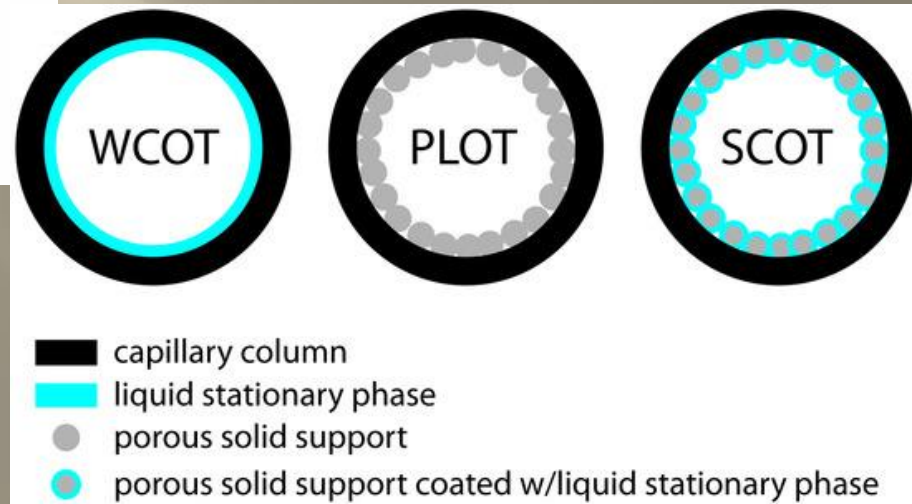
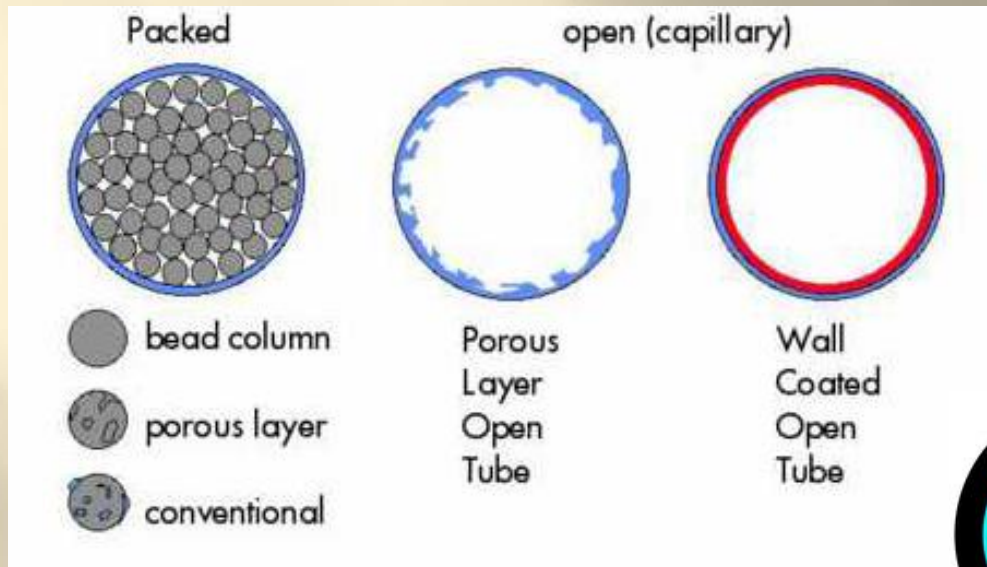
**In support-coated columns, the inner wall of the capillary is lined with a thin layer of support material such as diatomaceous earth, onto which the stationary phase has been adsorbed. SCOT columns are generally less efficient than WCOT columns.**

**Both types of capillary column are more efficient than packed columns.**



# Chromatography

## Columns



# Chromatography

## Gas Chromatography

### Column Materials

- Column tubing supports the stationary phase.
- Needs to be:
  - chemically inert (to prevent sample decomposition)
  - thermally stable
  - robust
  - flexible – form a coil to fit into oven

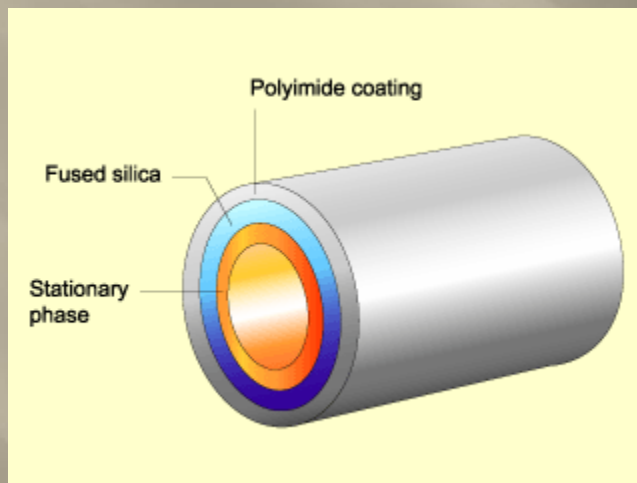
# Chromatography

## Column Materials

Early columns were metal  
(stainless steel, Cu, Al, Ni) but these were too reactive.

Glass is popular – but fragile

Fused silica columns – weak and subject to atmospheric corrosion.  
Use a protective outer Al or polymer sheath. These columns are  
used for most applications.



# Chromatography

Parameter	Capillary	Packed
Efficiency (plates/m)	2 000 – 4 000	500 - 1 000
Sample size (ng)	10 - 75	10 – 1 000 000
Relative pressure	Low	High
Relative speed	Fast	Slow
Chemical inertness	Best	Poorest
Flexible column	Yes	No



# Chromatography

## Gas Chromatography

### Columns

**Column Activity leads to:**

- tailing or skewed peaks**
- partial or total disappearance of a solute**
  - reversible peak adsorption**
  - non-reversible peak adsorption**
  - breakdown**

**The surface properties of the column – catalytic or sorptive are important to consider to eliminate column activity.**

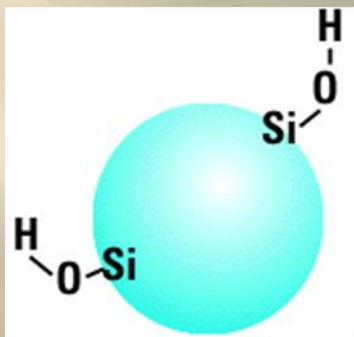


# Chromatography

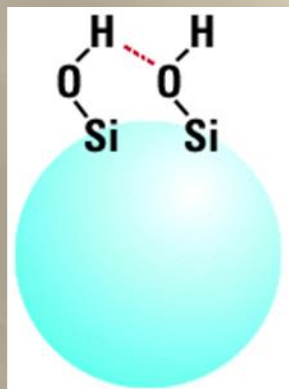
## Gas Chromatography

### Columns

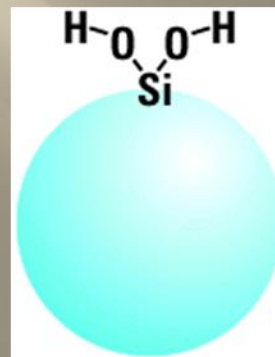
Various surface groups attributed to the silica surface have been identified:



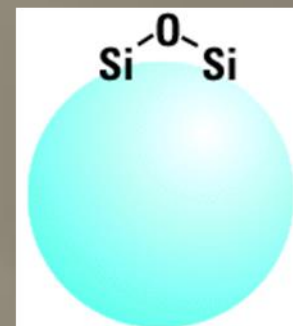
**Isolated  
Silanol**



**Vicinal  
Silanol**



**Geminal  
Silanol**



**Siloxane**

# Chromatography

## Gas Chromatography

### Columns

Surface hydroxyl groups can act as proton donors in hydrogen bonding interactions

- very strong sorptive sites for molecules with localized high electron density.

Surface siloxane bridge can act as

- proton acceptors functioning as sorptive sites for molecules such as alcohols

# Chromatography

## Columns

### **Metallic contamination**

**Metal ions present on the surface of soda lime or borosilicate glasses can function as Lewis acid sites absorbing molecules that have regions of localized high electron density such olefins, aromatic compounds, alcohols, ketones and amines. (Lewis acids – a species that accepts an electron pair)**

# Chromatography

## Gas Chromatography

### Columns

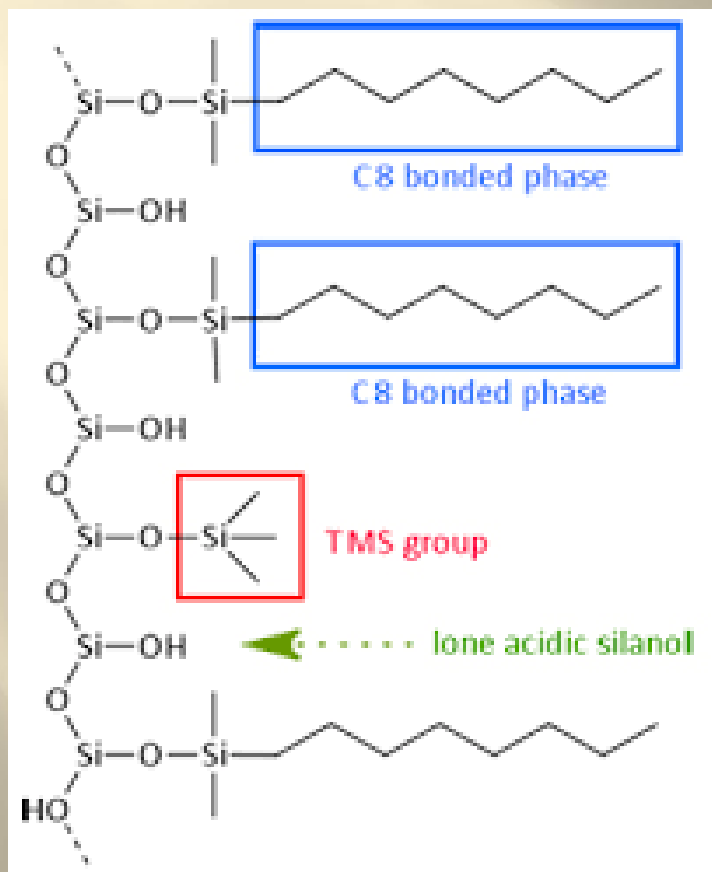
Columns are deactivated before use. One method deactivates reactive hydroxyl groups by silanizing or silylating the inner surface of the column.

Typical derivatization reactions are:

- Silylation
- Acylation
- Alkylation
- Esterification

# Chromatography

## Gas Chromatography



Columns are deactivated before use. Remember that these reactions are used to take care of a “problem” – but can just as well be used to “create” new stationary phases... we will cover this more later.

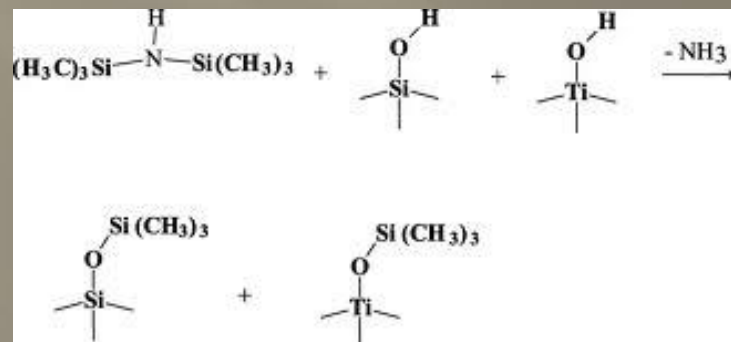
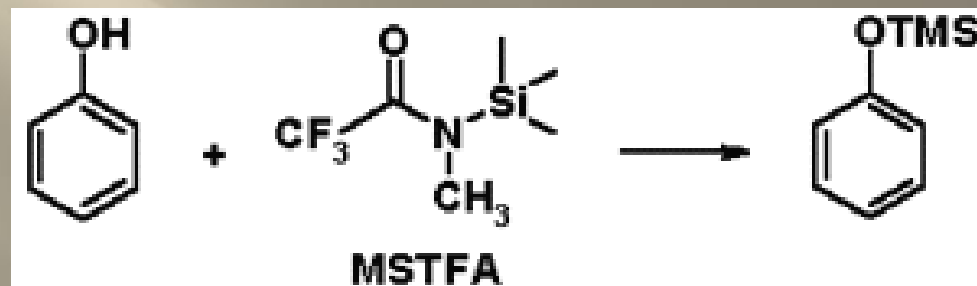
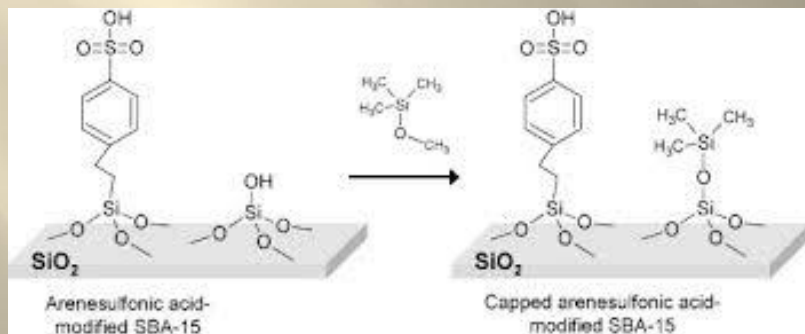


# Chromatography

## Gas Chromatography

### Columns

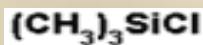
**Silylation** - It involves the replacement of an acidic hydrogen on the compound with an alkylsilyl group, for example,  $-\text{SiMe}_3$ . The derivatives are generally less polar, more volatile and more thermally stable.



# Chromatography

## Columns

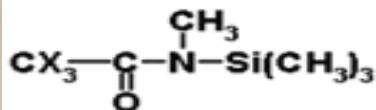
Various silanizing reagents can be used. The most common reagents for silylation are the trimethylsilyl (TMS) reagents.



Trimethylchlorosilane  
(TMCS)

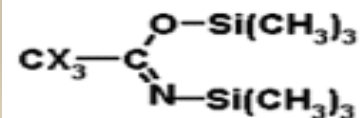


Hexamethyldisilazane  
(HMDS)



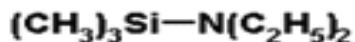
X=H, N-methyl-N-(trimethylsilyl)acetamide (MSTA)

X=F, N-methyl-N-(trimethylsilyl)trifluoroacetamide (MSTFA)

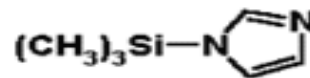


X=H, N,O-bis-(trimethylsilyl)acetamide (BSA)

X=F, N,O-bis-(trimethylsilyl)trifluoroacetamide (BSTFA)



N-trimethylsilyldiethylamine  
(TMSDEA)



N-trimethylsilylimidazole  
(TMSIM)

# Chromatography

## Gas Chromatography

### Columns

**Acylation** - Acylation is one of the most widely used derivatization procedures for gas chromatography. Acylation reduces the polarity of compounds that contain active hydrogens (-NH, -OH, -SH amino, hydroxyl and thiol groups) into amides, esters, or thioesters by using carboxylic derivatives.



# Chromatography

## Columns

The most widely used acylating reagents is acetic anhydride,  $(\text{CH}_3\text{CO})_2\text{O}$ .

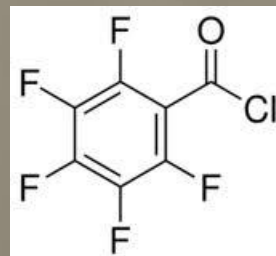
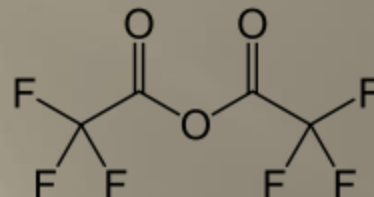
## Acylating reagents

Trifluoroacetic anhydride (TFAA)

Pentafluoropropionic anhydride (PFPA)

Heptafluorobutyryl anhydride (HFBA)

Pentafluorobenzoyl chloride



# Chromatography

## Columns

**Alkylation-** Alkylation is the replacement of a active hydrogen in  $R\text{-COOH}$ ,  $R\text{-OH}$ ,  $R\text{-SH}$ , and  $R\text{-NH}_2$  with an alkyl group or, sometimes aryl group.





# Chromatography

## Columns

### Alkylating reagents

- Alkyl halide (eg. pentafluorobenzyl bromide, PFBBBr, 3,5-ditrifluoromethylbenzyl bromide)
- Diazoalkane (eg. diazomethane)
- N,N-Dimethylformamide dialkyl acetal
- 3-alkyl-1-p-tolyltriazene
- Fluorosulfonate
- Trimethylanilinium hydroxide (TMAH)
- Trialkyloxonium fluoroborate
- Alkyl trichloroacetimidate

# Chromatography

## Columns

### Functional group

-OH (hydroxyl group) in primary, secondary and tertiary alcohols; phenols; carbohydrates)

-COOH (carboxylic acids)

S

-C=O (carbonyl group) in aldehydes and ketones

### Derivatization

Silylation  
Acylation  
Benzoylation  
Alkylation  
Dansylation  
Reaction with Dis-Cl  
Reaction with FDNB  
Reaction with NBD-Cl  
Ion-pair formation

Esterification  
silylation  
Ion-pair formation

Oxime formation  
Oxime formation and silylation  
Ketal/acetal formation  
Hydrazone formation  
Schiff's base formation  
Silylation

# Chromatography

## Functional group

-NH<sub>2</sub> (amino group) in primary amines, amino acids, amino sugars

-NH-R (amino group) in secondary amines, imino acids, substituted amino sugars

-NH<sub>2</sub> and -COOH in amino acids

-NO<sub>2</sub> (nitro compounds)

## Derivatization

Acylation  
Benzoylation  
Silylation  
Treatment with CS<sub>2</sub>  
Thiourea formation  
Schiff's base formation  
2,4-Dinitrophenylation  
Sulphonamide formation  
Carbamate formation  
Treatment with pyridoxal  
Treatment with NBD-Cl  
Alkylation  
Ion-pair formation

Acylation  
Benzoylation  
Silylation  
2,4-Dinitrophenylation  
Sulphonamide formation  
Treatment with NBD-Cl  
Ion-pair formation

Silylation  
Esterification + Acylation

Chromatograph without derivatization

# Chromatography

## Columns

### Stationary Phases

Selection of stationary phase affects  $k$  and  $\alpha$  values.

Main concerns of stationary phase are:

- polarity,
- functional groups,
- maximum operating temperature, and
- column bleed (loss of stationary phase due to decomposition)

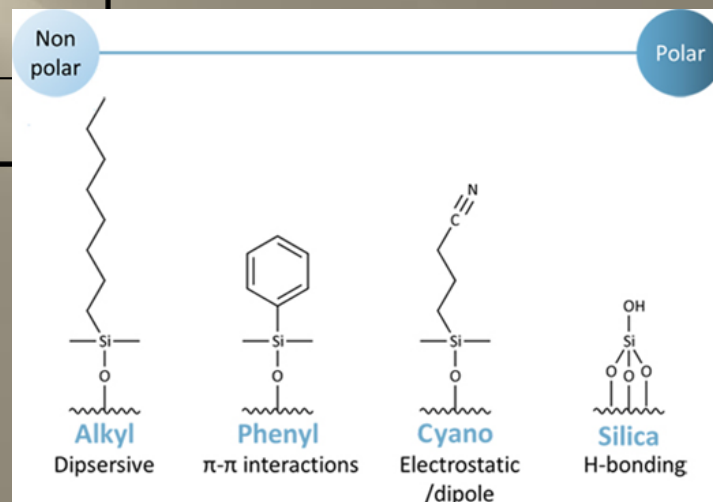
More polar columns suffer from lower maximum temperatures and greater column bleed

# Chromatography

## Columns

### Stationary Phases (examples)

Type	Functional Groups	Polarity
OV-1	methyl	Non-polar
OV-17	50% methyl/50% phenyl	Somewhat polar
OV-225	Cyanopropyl, methyl, and phenyl	More polar
carbowax	Ether groups	polar





# Chromatography

## Columns

### Stationary Phases (GSC)

GSC of inorganic gases is the one area where packed columns are still used almost exclusively. Main adsorbents for packed columns are based on:

- silica
- charcoal
- alumina
- molecular sieves

# Chromatography

## Columns

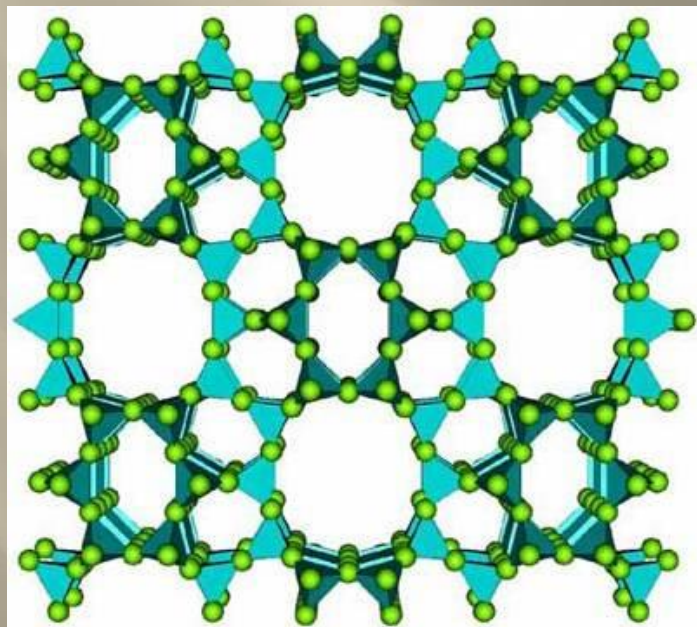
### Stationary Phases (GSC)

- Porous silica is used with a variety of surface areas and pore diameters (separate low molecular mass hydrocarbons).
- Alumina has a variety of surface areas – but different selectivities due to Lewis acid sites associated with aluminum ions on the surface. To modify selectivity and improve efficiency, both silica and alumina can be coated with a non-volatile liquid.

# Chromatography

## Stationary Phases (GSC)

- Molecular sieves are artificially prepared zeolites (i.e. Na, K, Ca – aluminosilicates) – small pore diameter used to separate low molecular weight molecules and hydrocarbons.

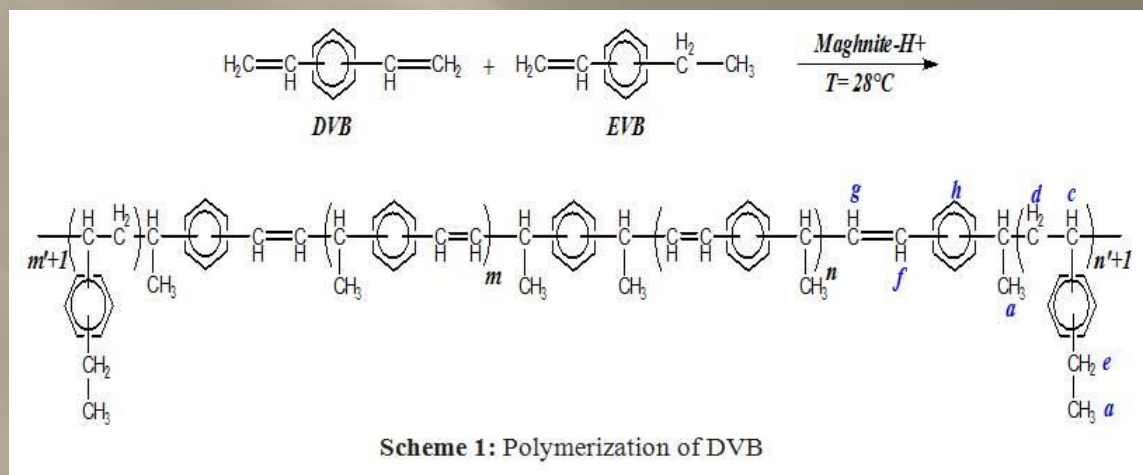
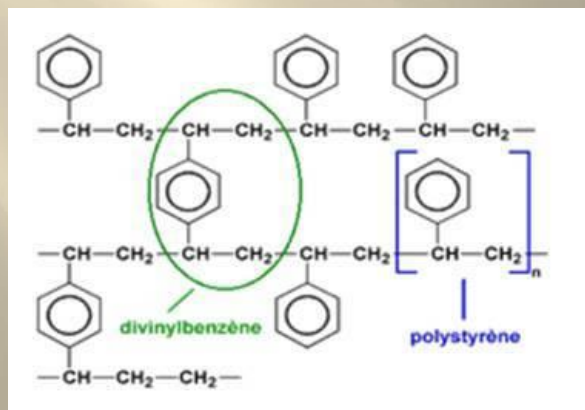


# Chromatography

## Stationary Phases (GSC)

Porous polymers – good for separating polar analytes. Usually consist of copolymers of divinylbenzene (DVB),  $\text{CH}_2=\text{CH}$ -(benzene ring)- $\text{CH}=\text{CH}_2$ , with another aromatic olefin (ethylene  $\text{CH}_2=\text{CH}_2$ ). Easy to use and has superior performance and unique separating abilities.

The nature of the separation is adsorption, but the mechanism is unclear.



# Chromatography

## Columns

### Stationary Phases (GSC)

#### Adsorbents for GSC

<u>Chemical Type</u>	<u>Specific Surface Area (m<sup>2</sup>g<sup>-1</sup>)</u>	<u>Pore Diameter (nm)</u>
Silica	~ 100 – 185	15 – 30
Alumina	-	-
Carbon Black	10 – 1000	1 – 2
Molecular Sieves	700 – 1000	0.5 – 2.0
Porous Polymers	~ 100 – 800	5 – 500



# Chromatography

## Columns - GLC

**Wall-coated columns consist of a capillary tube whose walls are coated with liquid stationary phase.**

**In support-coated columns, the inner wall of the capillary is lined with a thin layer of support material such as diatomaceous earth, onto which the stationary phase has been adsorbed.**

### **Liquid Phase – required properties**

- **low vapor pressure**
- **thermal and chemical stability**
- **high viscosity**
- **nonreactive toward sample components**
- **wide operating temperatures (-80 to 450°C)**
- **reasonable solvent properties**



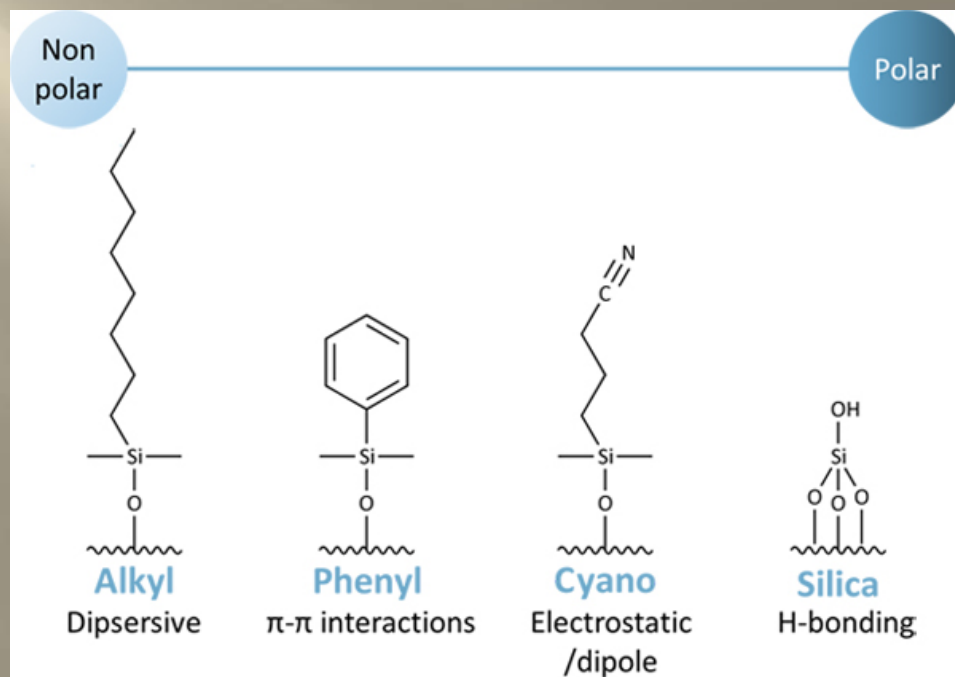
# Chromatography

## Columns

## Stationary Phases (GLC)

## Categories of stationary phases

- **non-polar**
- **polar**
- **specialty phases**



# Chromatography

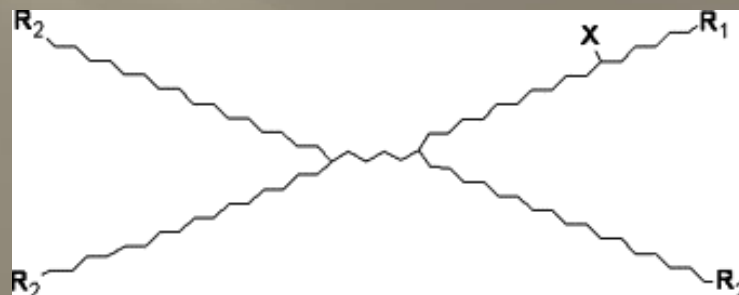
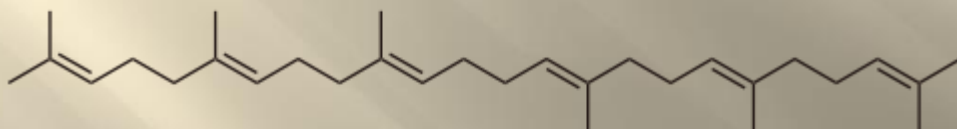
## Columns

### Stationary Phases (GLC)

Hydrocarbons – non-polar – have been used as non-polar stationary phase.

Most have a high molecular weight for low volatility.

Examples: Squalane ( $C_{30}H_{62}$ ), Apolane C87, long-chain n-alkanes, Apiezon L-not used much.



# Chromatography

## Columns

### Stationary Phases (GLC)

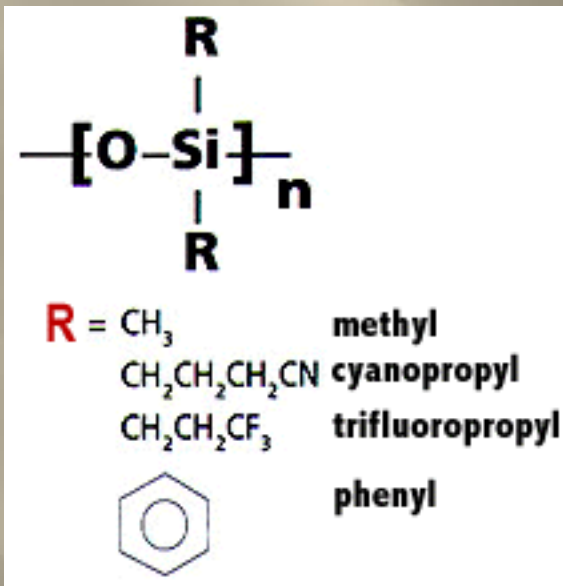
**Alkylsilicone Phases – can be non-polar or polar. Polymers based on silicon-oxygen-silicone backbone – most widely used group of stationary phases. Differences in these stationary phases differ mainly in the degree of substitution on the silicon backbone.**

# Chromatography

## Columns

### Stationary Phases (GLC)

Standard polysiloxanes are characterized by the repeating siloxane backbone. Each silicon atom contains two functional groups. The type and amount of the groups distinguish each stationary phase and its properties.



# Chromatography

## Columns

### Stationary Phases (GLC)

The most basic polysiloxane is the 100% methyl substituted. When other groups are present, the amount is indicated as the percent of the total number of groups.

For example, a 5% diphenyl-95% dimethyl polysiloxane contains 5% phenyl groups and 95% methyl groups.

The "di-" prefix indicates that each silicon atom contains two of that particular group.

Sometimes this prefix is omitted even though two identical groups are present.

If the methyl percentage is not stated, it is understood to be present in the amount necessary to make 100% (e.g., 50% phenyl-methyl polysiloxane contains 50% methyl substitution).

# Chromatography

## Columns

### Stationary Phases (GLC)

Cyanopropylphenyl percent values can be misleading.

A 14% cyanopropylphenyl-dimethyl polysiloxane contains 7% cyanopropyl and 7% phenyl (along with 86% methyl).

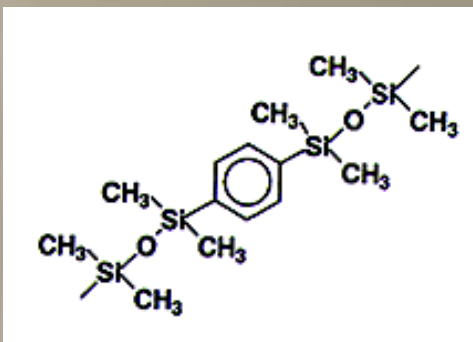
The cyanopropyl and phenyl groups are on the same silicon atom, thus their amounts are summed.



# Chromatography

## Columns

### Stationary Phases (GLC)



For select stationary phases, a low bleed or "ms" version is available.

These stationary phases incorporate phenyl or phenyl type groups into the backbone of the siloxane polymer.

These types of stationary phases are commonly called arylenes.

The phenyl group strengthens and stiffens the polymer backbone which inhibits stationary phase degradation at higher temperatures.

This results in lower column bleed and, in most cases, higher temperature limits.

# Chromatography

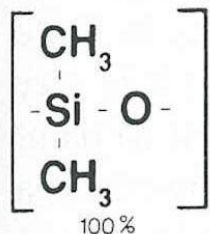
## Stationary Phases (GLC)

Chemical structure

Classification

Uses

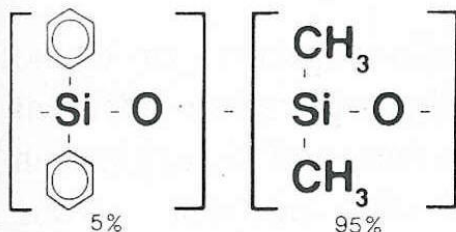
100 % dimethyl silicone



Non-polar

Boiling point  
separations (solvents,  
petroleum products,  
pharmaceuticals)

95 % dimethyl }  
5 % phenyl } silicone



Non-polar

Boiling point  
separations (aromatics,  
flavours, aromatic  
hydrocarbons)

86 % dimethyl }  
7 % phenyl }  
7 % cyanopropyl } silicone

Intermediate  
polarity

Pesticides, alcohols

# Chromatography

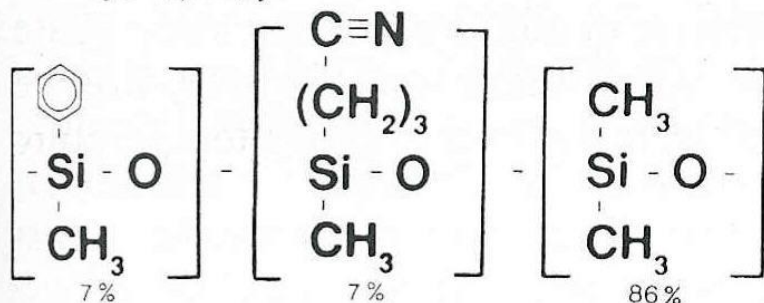
## Stationary Phases (GLC)

86% dimethyl  
7% phenyl  
7% cyanopropyl

} silicone

Intermediate  
polarity

Pesticides, alcohols

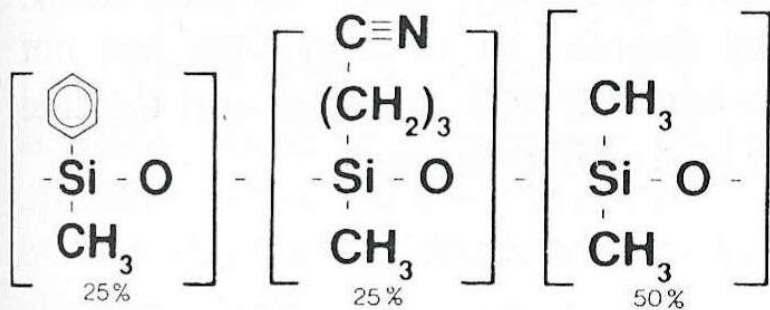


50% dimethyl  
25% phenyl  
25% cyanopropyl

} silicone

Polar

Triglycerides, phthalate  
esters



# Chromatography

## Columns

### Stationary Phases (GLC)

**Polyether Phases – polar phases – based on polyethylene glycol,**



**Marketed under the trade name, carbowax or superox.**

**Polyethylene glycols stationary phases are not substituted, thus the polymer is 100% of the stated material. They are less stable, less robust and have lower temperature limits than most polysiloxanes.**

**With typical use, they exhibit shorter lifetimes and are more susceptible to damage upon over heating or exposure to oxygen.**

# Chromatography

## Columns

### Stationary Phases (GLC)

Polyether Phases – polar phases – based on polyethylene glycol,



Advantage - Average molecular weight of 20,000, so can be used at high temperatures.

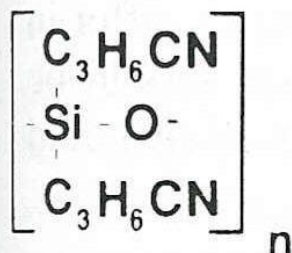
Disadvantage is that it reacts with even trace amounts of oxygen at high temperatures, as well as decomposing to acetaldehyde and acetic acid.



# Chromatography

## Stationary Phases (GLC)

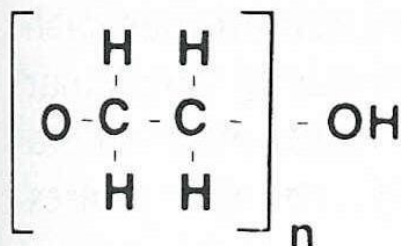
100 % cyano propyl silicone



Polar

Fatty acid methyl  
esters, carbohydrates

polyethylene glycol 20M



Polar

Flavours, fatty acid  
methyl esters, acids,  
amines

Fig. 3.17. Structure of polysiloxane and polyethylene glycol stationary phases.



# Chromatography

## Columns

### Stationary Phases (GLC)

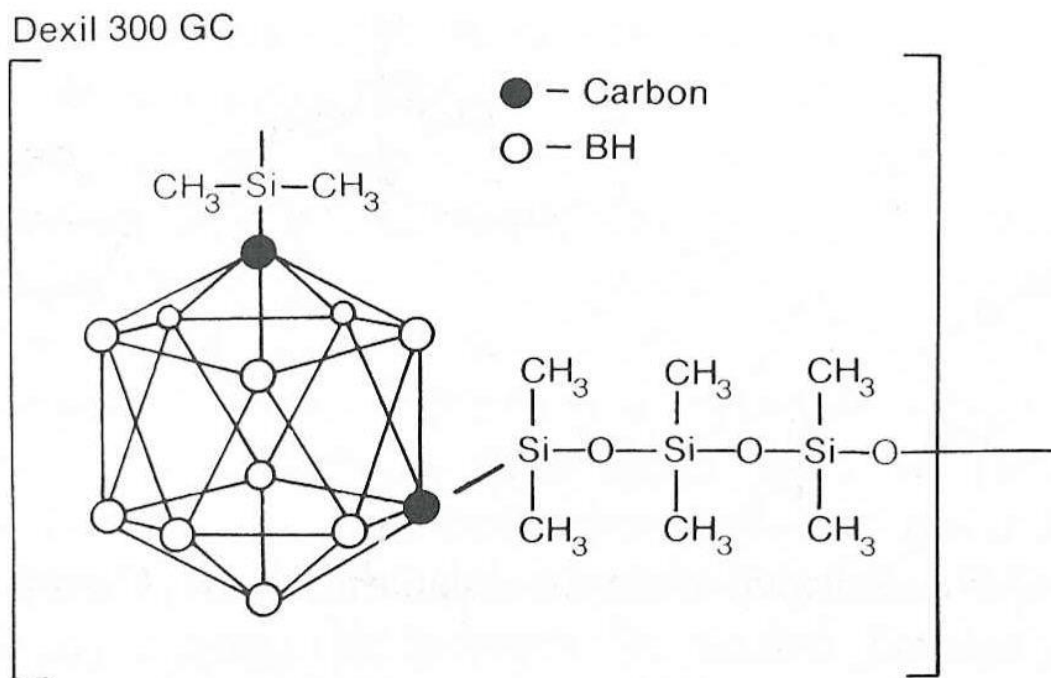
High-temperature phases – substituted silicone-carborane copolymers.

Stable up to 450°C - 500°C.

Their advantage is in their low stationary bleed – for use with highly sensitive detectors.

# Chromatography

## Stationary Phases (GLC) High-temperature phases



**Fig. 3.18.** A silicone-carborane copolymer used as a high temperature stationary phase.

# Chromatography

## Columns

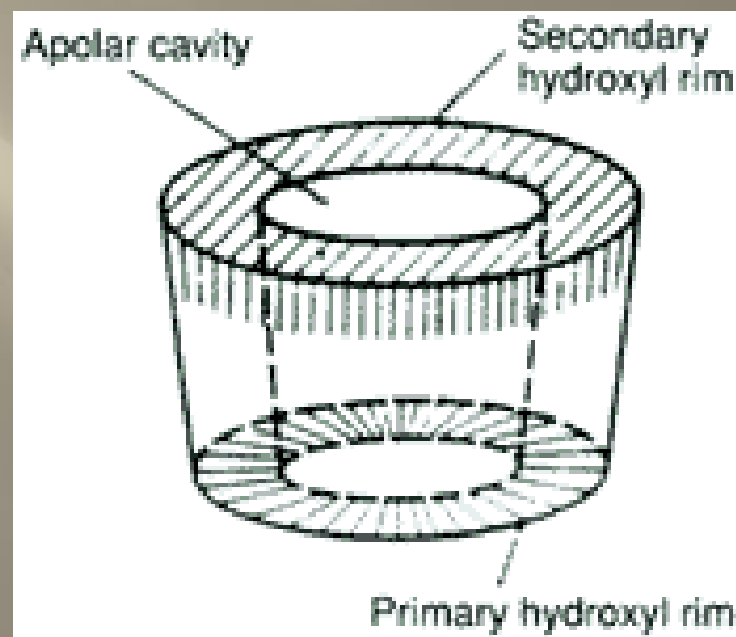
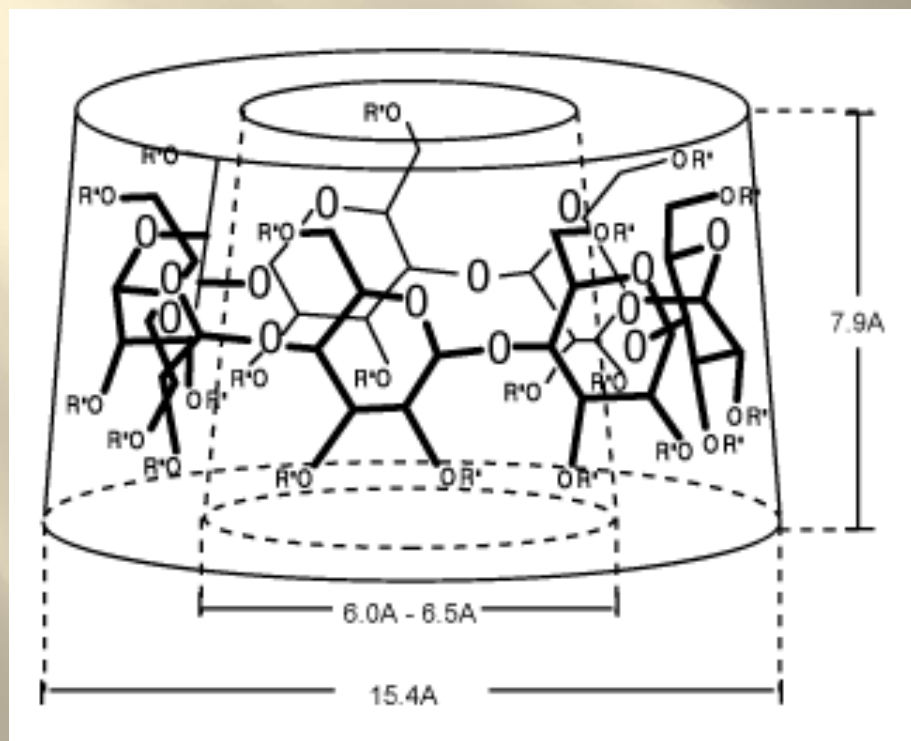
### Stationary Phases (GLC)

**Chiral Phases - used to distinguish between enantiomers of optically active compounds.**  
 **$\alpha, \beta, \gamma$  – cyclodextrins – composed of six or more D(+)-glucose units bonded through a  $\alpha$ -(1,4) – glycosidic linkage.**

# Chromatography

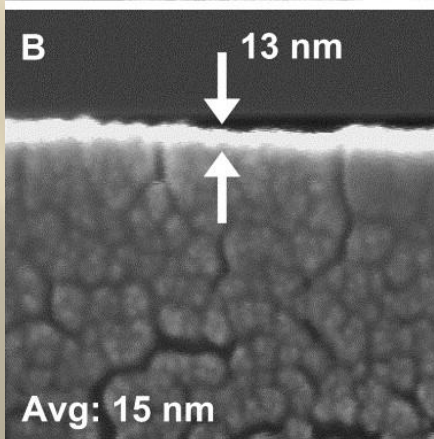
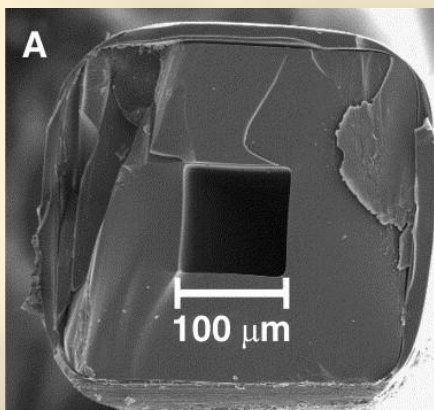
## Columns

### Stationary Phases (GLC)

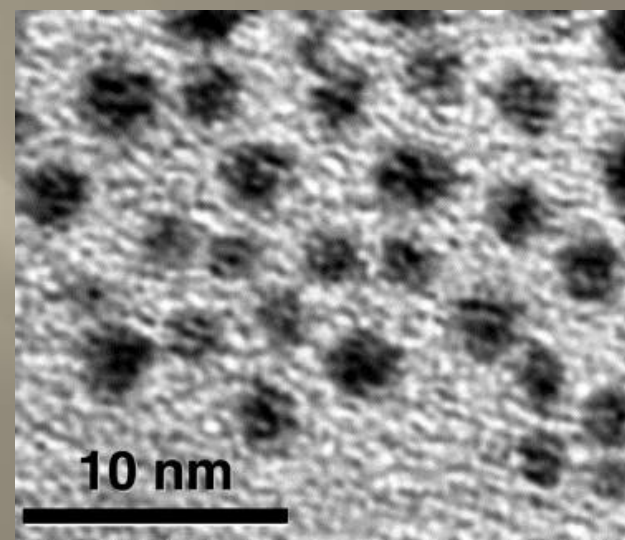
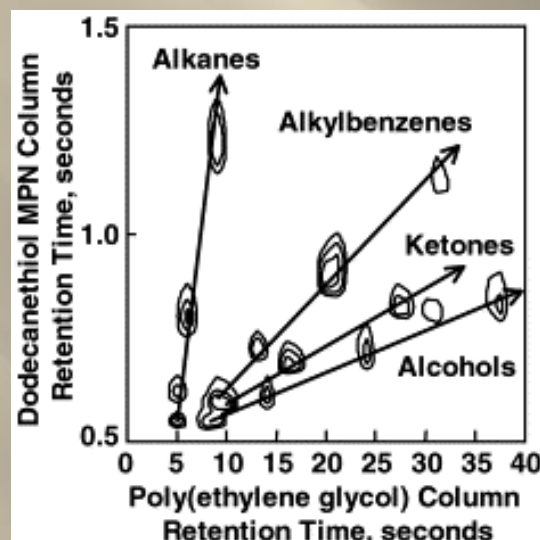


# Chromatography

## Stationary Phases



Dodecanethiol MPNs, in which the monolayer is dodecanethiol linked to the gold nanoparticle



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# Chromatography

## Columns

### Temperature Limits

Columns have lower and upper temperature limits.

If a column is used below its lower temperature limit, rounded and wide peaks are obtained (i.e., loss of efficiency).

No column damage has occurred; however, the column does not function properly.

Using the column at or above its lower limit maintains good peak shapes.



# Chromatography

## Columns

### Temperature Limits

Columns have lower and upper temperature limits. Upper temperature limits are often stated as two numbers.

The lower one is the isothermal temperature limit. The column can be used indefinitely at this temperature and reasonable column bleed and lifetime are realized.

The upper number is the temperature program limit. A column can be maintained at this temperature for 10-15 minutes without severely shortening column lifetime or experiencing excessively high column bleed.

Exceeding the upper temperature limits may damage the stationary phase.

# Chromatography

## Columns

### Temperature Programming

The column sits in an oven. If the temperature is held constant during the entire analysis it is isothermal.

If you vary the temperature during the analysis, you typically use a temperature program.

With homologues, the retention time increases exponentially with the number of carbon.

As  $t_R$  increases, width increases and the height decreases, making detection impossible after a few peaks have eluted.

# Chromatography

## Columns

## Temperature Programming

General steps to create a program assuming that the separation is possible.

- Determine initial temperature and time based on best possible separation of first few peaks.
- Repeat 1 for the last few peaks to find the best final temperature and time.
- Experiment with various ramps to account for the rest of the components.

# Chromatography

## Columns

### Capacity

Column capacity is the maximum amount of a solute that can be introduced into a column before significant peak distortion occurs. No damage occurs if a column is overloaded.

Column capacity is related to stationary phase polarity, film thickness, column diameter and solute retention. Higher column capacities are obtained for solutes that are similar in polarity to the stationary phase.

For example, a polar column has a higher capacity for a polar solute than a non-polar solute.

Thicker film and wider diameter columns have higher capacities.

# Chromatography

## Columns

### Stationary Phase Film Thickness

Stationary phase loadings were 20-30% w/w prior to 1970's.

Now loadings are in the order of 1-3%.

Film thickness,  $d_f$ , has a direct effect on the retention, sample capacity and elution temperature.

For best column efficiency, the film thickness is kept as thin as possible in order to reduce resistance to mass transfer in the stationary phase,  $C_s$ .



# Chromatography

## Columns

### Column Capacity in ng

Film Thickness ( $\mu\text{m}$ )	Column Diameter (mm)			
	0.18-0.20	0.25	0.32	0.53
0.10	20-35	25-50	35-75	50-100
0.25	35-75	50-100	75-125	100-250
0.50	75-150	100-200	125-250	250-500
1.00	150-250	200-300	250-500	500-1000
3.00		400-600	500-800	1000-2000
5.00		1000-1500	1200-2000	2000-3000

# Chromatography

## Columns

### Film Thickness Selection

1. For 0.18-0.32 mm I.D. columns, a film thickness of 0.18-0.25  $\mu\text{m}$  is average or standard (i.e., not thin or thick) and used for most analyses.
2. For 0.45-0.53 mm I.D. columns, a film thickness of 0.8-1.5  $\mu\text{m}$  is average or standard (i.e., not thin or thick) and used for most analyses.
3. Thick film columns are used to retain and resolve volatile solutes (e.g., light solvents, gases). Thick columns are more inert and have higher capacities. Thick film columns exhibit higher column bleed and decreased upper temperature limits.
4. Thin film columns are used to minimize the retention of high boiling, high molecular weight solutes (e.g., steroids, triglycerides). Thin columns are less inert and have lower capacities. Thin film columns exhibit lower column bleed.

# Chromatography

## Columns

### Column Length

Column length influences three parameters:

- efficiency,
- retention (analysis time) and
- pressure.

# Chromatography

## Columns

### Column Length

Column efficiency ( $N$ ) is proportional to column length.

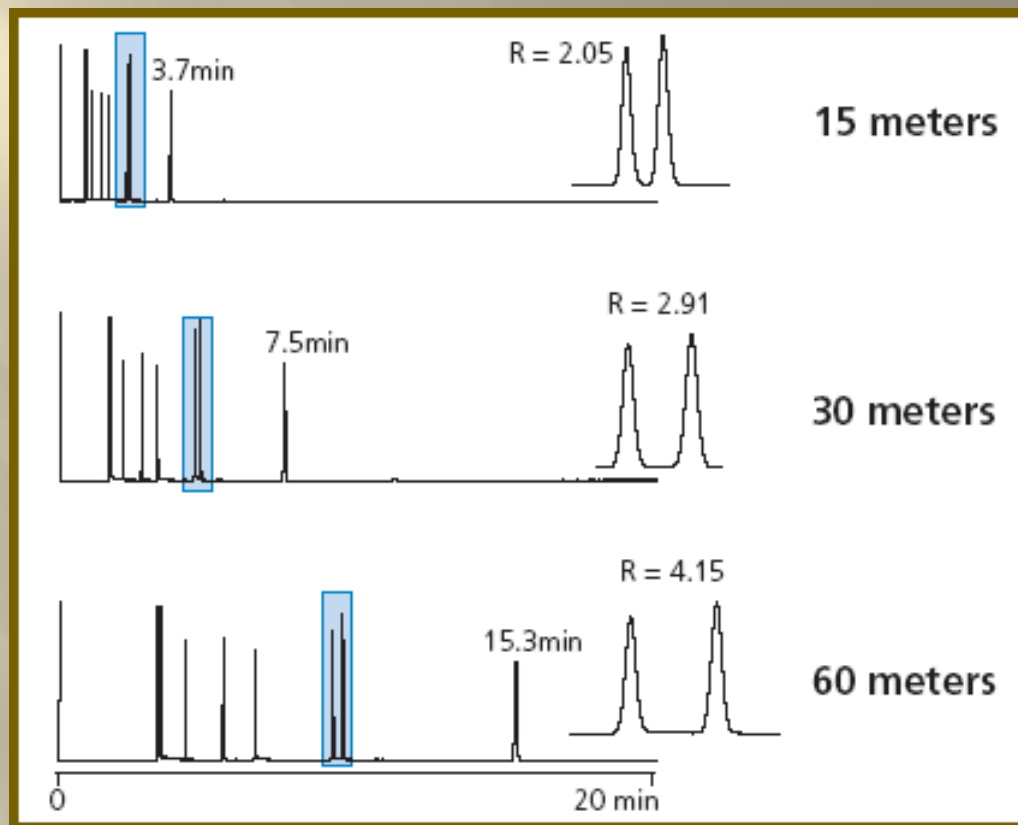
Resolution is a square root function of the theoretical plate number. For example, doubling column length (thus efficiency) theoretically increases resolution by 1.41 times (closer to 1.2-1.3 times in real practice).

Longer columns are used when peak separation is small and high column efficiency (i.e., narrow peaks) is needed.

# Chromatography

## Columns

### Column Length



Longer columns are used when peak separation is small and high column efficiency (i.e., narrow peaks) is needed.



# Chromatography

## Columns

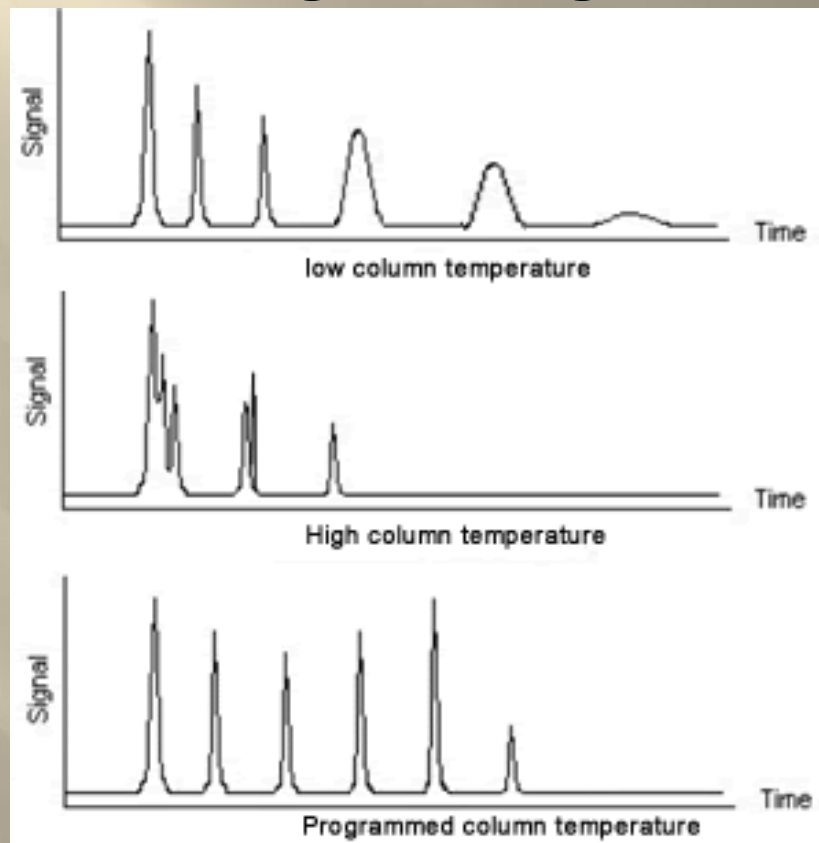
### Column Length Selection

1. Start with 25-30 meter columns when the best length is unknown.
2. 10-15 meter columns are well suited for samples containing very well separated solutes or very few solutes. Shorter lengths are used for very small diameter columns to reduce head pressures.
3. 50-60 meters should be used when resolution is not possible by other means (smaller diameter, different stationary phase, change in column temperature). Best suited for complex samples containing a large number of solutes. Long columns have long analysis times and higher cost.

# Chromatography

## Columns

### Column Temperature Programming



# Chromatography

## Columns

### Stationary Phase Selection

Usually historically based – do what someone else did.

### Selecting the Stationary Phase

Consider:

- polarity
- temperature limits
- column efficiency
- detector compatibility.

# Chromatography

## Columns

### Column Selection

**Non-polar phases usually exhibit superior lifetimes – so best to select least polar phase needed for a particular separation.**

**Most difficult factor to access is the ability of a phase to produce a good separation. In theory selection is based on knowing the types of interactions.**

# Chromatography

## Columns

### Column Selection

#### Types of interactions:

- London or dispersive forces (weak, non-specific). Temporarily induced dipole. As molecular weight increases dispersive forces increase (electrons further from nucleus)
- Dipole-dipole interactions (polar molecules)
- Acid – Base interactions (proton sharing or electron transfer) (hydrogen bonding)

-Dispersion is the dominant interaction for all polysiloxane and polyethylene glycol stationary phases.

-Dispersion can be simplified into the concept of volatility. The more volatile a solute, the faster it elutes from the column (i.e., shorter retention time).



# Chromatography

## Columns

### Types of interactions

- Empirical results have shown that dipole interaction stationary phases are well suited for samples containing compounds that have base or central structures to which different groups are attached in various positions.
- Examples include substituted aromatics, halocarbons, pesticides and drugs.

# Chromatography

## Columns

### Types of interactions

- The hydrogen bonding interaction occurs if there is hydrogen bonding between the solute molecules and the stationary phase.
- It is the difference in the strength of the hydrogen bonding that is critical.

### Relative Hydrogen Bonding Strengths

Strength	Compounds
Strong	Alcohols, carboxylic acids, amines
Moderate	Aldehydes, esters, ketones
Weak to None	Hydrocarbons, halocarbons, ethers

# Chromatography

## Columns

### Stationary phase interaction

Functional Group	Dispersion	Dipole	Hydrogen Bonding
Methyl	Strong	None	None
Phenyl	Very Strong	None	Weak
Cyanopropyl	Strong	Very Strong	Moderate
Trifluoropropyl	Strong	Moderate	Weak
PEG	Strong	Strong	Moderate

# Chromatography

## Columns

**Each chromatographic setup will vary to some degree.**

**Retention times for a known set of species can be hard to reproduce from one lab to another or even one instrument to another.**

**Retention indexing helps to standardize your results.**

# Chromatography

## Columns

### Column Selection

Methods to characterize stationary phases  
(Chromatographic systems):

Kovats Index/Rohrschneider Constants/McReynolds Constants

Use the relative retention of a solute – plot of the logarithm of the adjusted retention time versus Kovats retention index,  $I$  (linear).

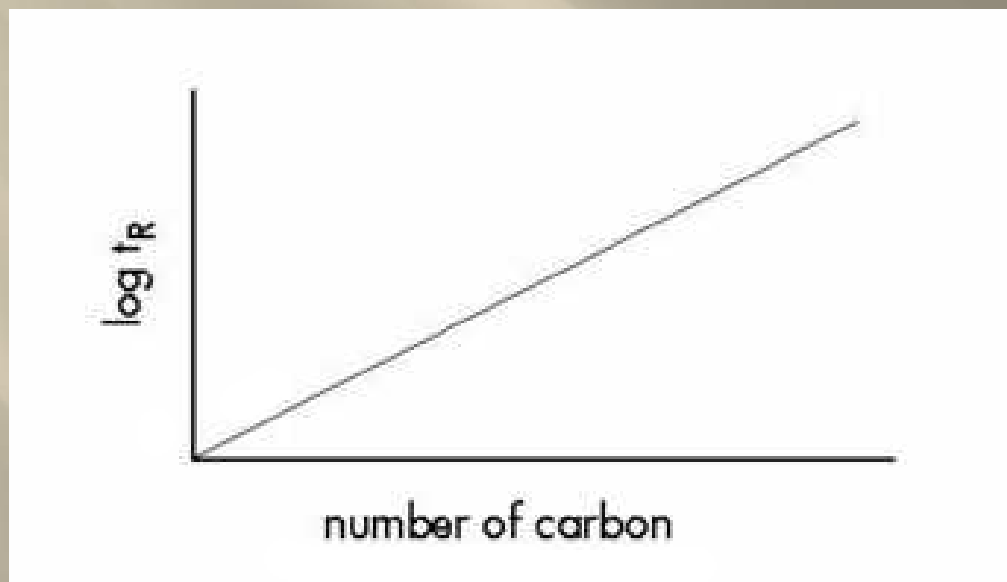


# Chromatography

## Columns

Method is based on results of a homologous series.

Plot of the logarithm of the adjusted retention time versus Kovats retention index,  $I$  (linear).



# Chromatography

## Columns

The Kovat index value for any material can be found by assigning the retention index (I) of an n-alkane to equal a value 100 times its carbon number.

n-octane = 800

n-decane = 1000

n-dodecane = 12000, etc.

An I value of something different than n-alkane can be determined by “spiking” the mixture with n-alkanes. Plot  $\log t'_r$  vs. I (or use equation) and I of solute is determined.

# Chromatography

## Columns

Equation:

$$I_x = 100Z + \frac{100(\log t'_{r,x} - \log t'_{r,z})}{\log t'_{r,z+1} - \log t'_{r,z}}$$

where,

**Z** - carbon # of n-alkane.

**t'\_{r,x}** - adjusted retention time of component under consideration.

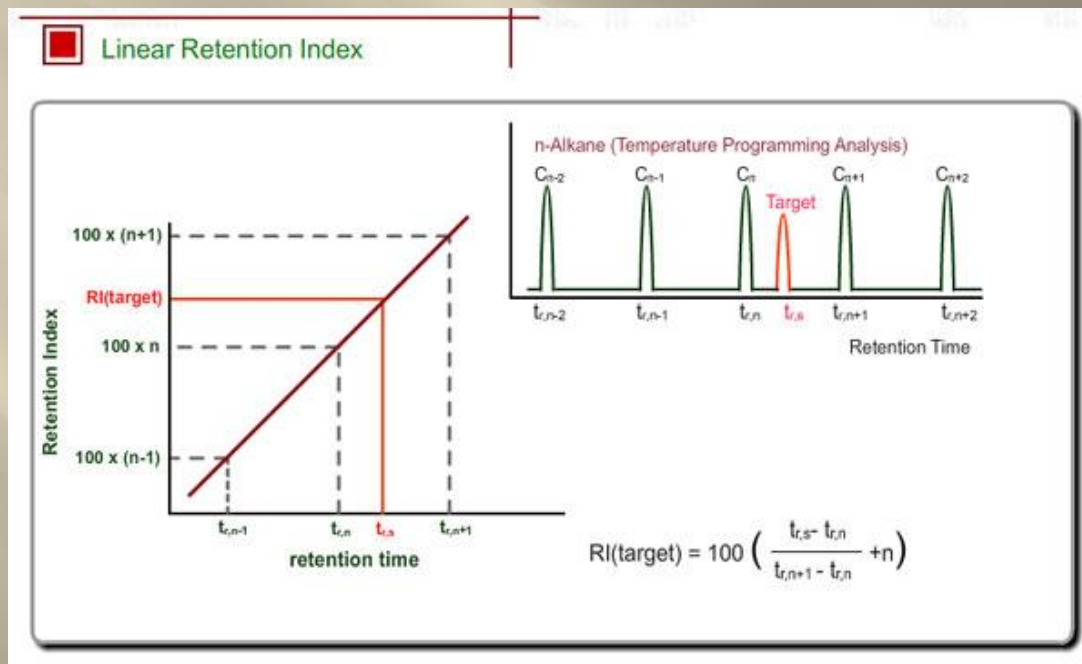
**t'\_{r,z}** - adjusted retention time of alkane eluting before x

**t'\_{r,z+1}** - adjusted retention time of alkane eluting after x.

# Chromatography

## Columns

All that is really being done is to normalize each component compared to n-alkanes. It assumes that you are dealing with either identical or at least very similar columns or packings.



# Chromatography

## Columns

Methods to evaluate a wide range of phases are based on measuring performance for a set of representative substances.

Can then tell if two phases should give comparable performance or if a phase is better for specific functional groups.

McReynolds Constants or Rohrschneider Constants



# Chromatography

## Columns

Use a series of reference compounds.

McReynolds used 10 probe solutes – each with a different functionality to measure a specific interaction with a liquid phase. He measured  $I$  for the 10 compounds on over 200 phases, including squalene (serves as a reference).

Calculated  $\Delta I$  value for each probe =  $I_{\text{liquid phase}} - I_{\text{squalene}}$

# Chromatography

## Columns

**As  $\Delta I$  increases for a probe on a given liquid phase, the degree of a specific interaction associated with the probe increases.**

**The probes are assigned symbols of X', Y', Z', U', S' (Rohrschneider Constants) and McReynolds expanding it adding H', J', K', L', M'.**

**For squalene all = 0**

# Chromatography

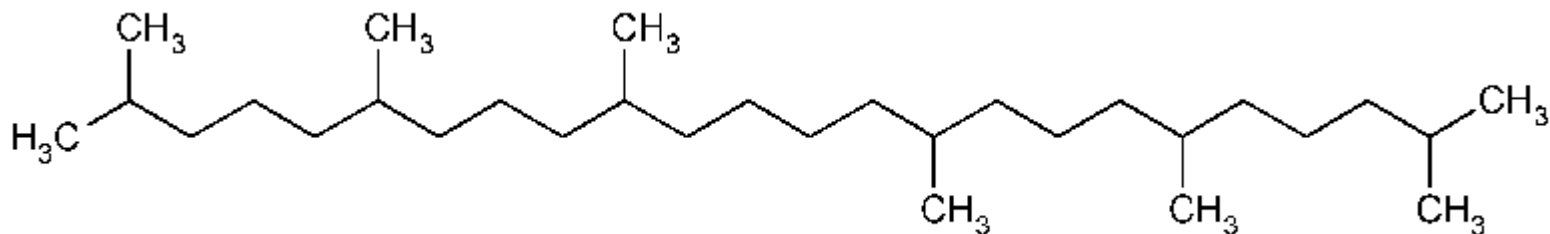
## Columns

Group	Substance	Symbol
aromatic, olefinic	benzene	X'
alcohols, phenols, acids	1-butanol	Y'
ketones, ethers, esters, aldehydes	methyl-n-propyl ketone	Z'
nitro, nitriles	nitropropane	U'
bases, aromatic heterocyclics	pyridine	S'

# Chromatography

## Columns

**Squalane is used as the reference material and all other packings are normalized to it.**



Packing	TMAX	X'	Y'	Z'	U'	S'
squalane	150	0	0	0	0	0
SE-30	350	15	53	44	64	41
OV-7	350	69	113	111	171	128
Carbowax 20M	250	322	536	368	572	59

# Chromatography

## Columns

TABLE 3.4 Probes Used in the McReynolds and Rohrschneider Classifications of Liquid Phases

Symbol	McReynolds Probe	Rohrschneider Probe	Measured Interaction
X'	Benzene	Benzene	Electron density for aromatic and olefinic hydrocarbons
Y'	<i>n</i> -Butanol	Ethanol	Proton donor and proton acceptor capabilities (alcohols, nitriles)
Z'	2-Pentanone	2-Butanone	Proton acceptor interaction (ketones, ethers, aldehydes, esters)
U'	Nitropropane	Nitromethane	Dipole interactions
S'	Pyridine	Pyridine	Strong proton acceptor interaction
H'	2-Methyl-2-pentanol	—	Substituted alcohol interaction similar to <i>n</i> -butanol
J'	Iodobutane	—	Polar alkane interactions
K'	2-Octyne	—	Unsaturated hydrocarbon interaction similar to benzene
L'	1,4-Dioxane	—	Proton acceptor interaction
M'	<i>cis</i> -Hydrindane	—	Dispersion–interaction

# Assignment

- ▣ Test II



