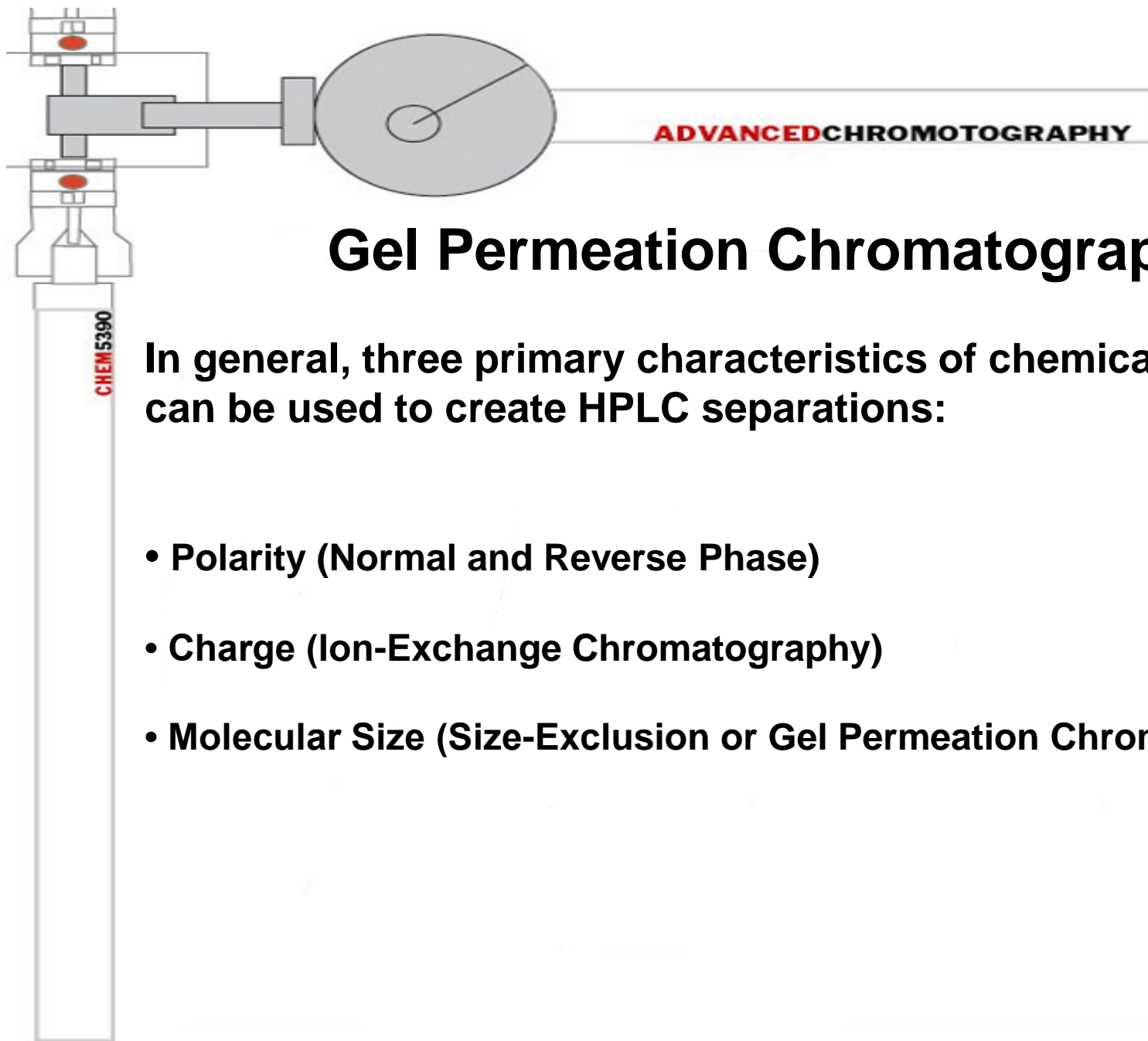


# Lecture 19

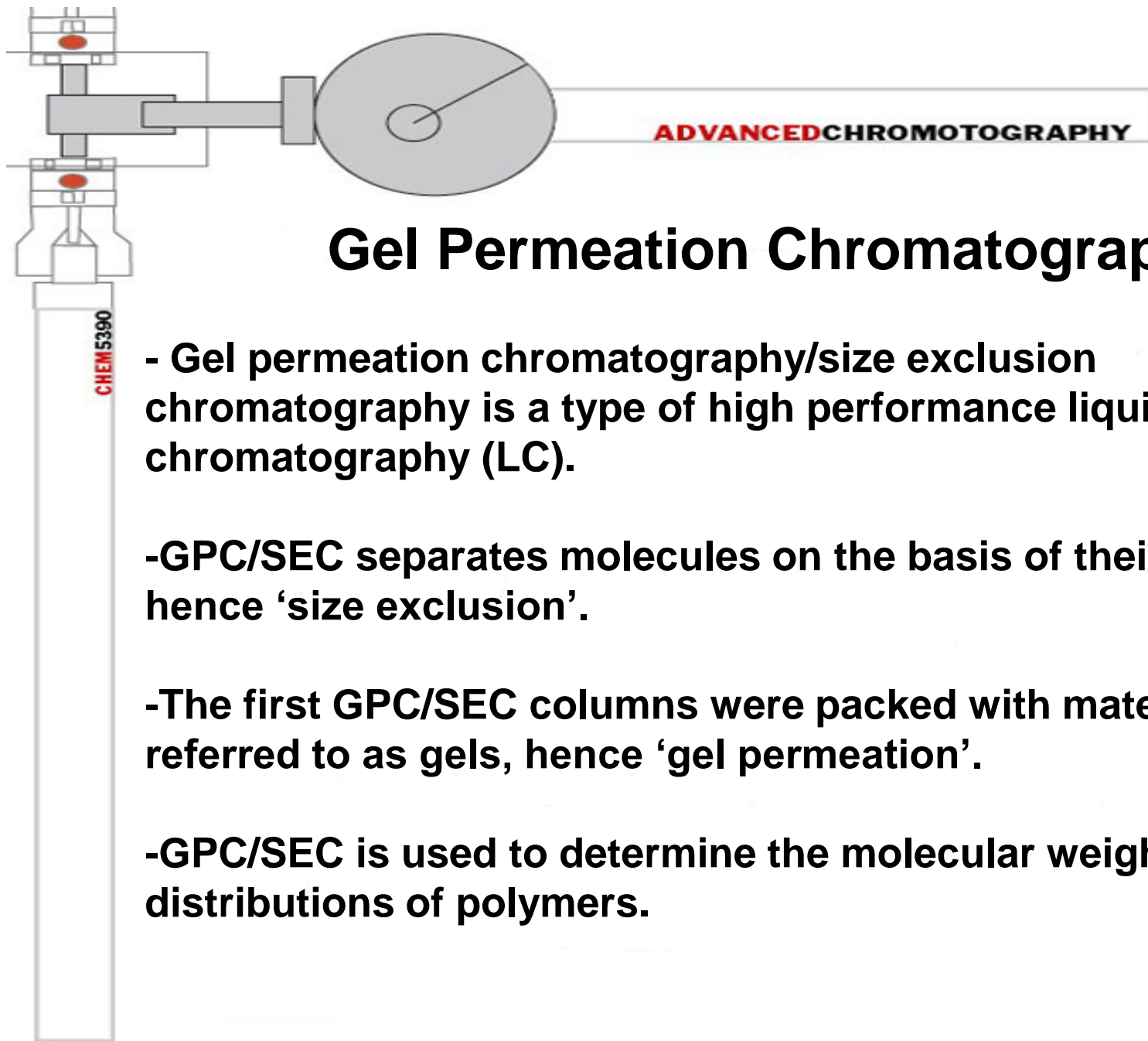
## Gel Permeation or Size Exclusion Chromatography



## Gel Permeation Chromatography

In general, three primary characteristics of chemical compounds can be used to create HPLC separations:

- Polarity (Normal and Reverse Phase)
- Charge (Ion-Exchange Chromatography)
- Molecular Size (Size-Exclusion or Gel Permeation Chromatography)



## Gel Permeation Chromatography

- Gel permeation chromatography/size exclusion chromatography is a type of high performance liquid chromatography (LC).
- GPC/SEC separates molecules on the basis of their size, hence 'size exclusion'.
- The first GPC/SEC columns were packed with materials referred to as gels, hence 'gel permeation'.
- GPC/SEC is used to determine the molecular weight distributions of polymers.



# Gel Permeation Chromatography

Small  
molecules

Macromolecules

HPGPC

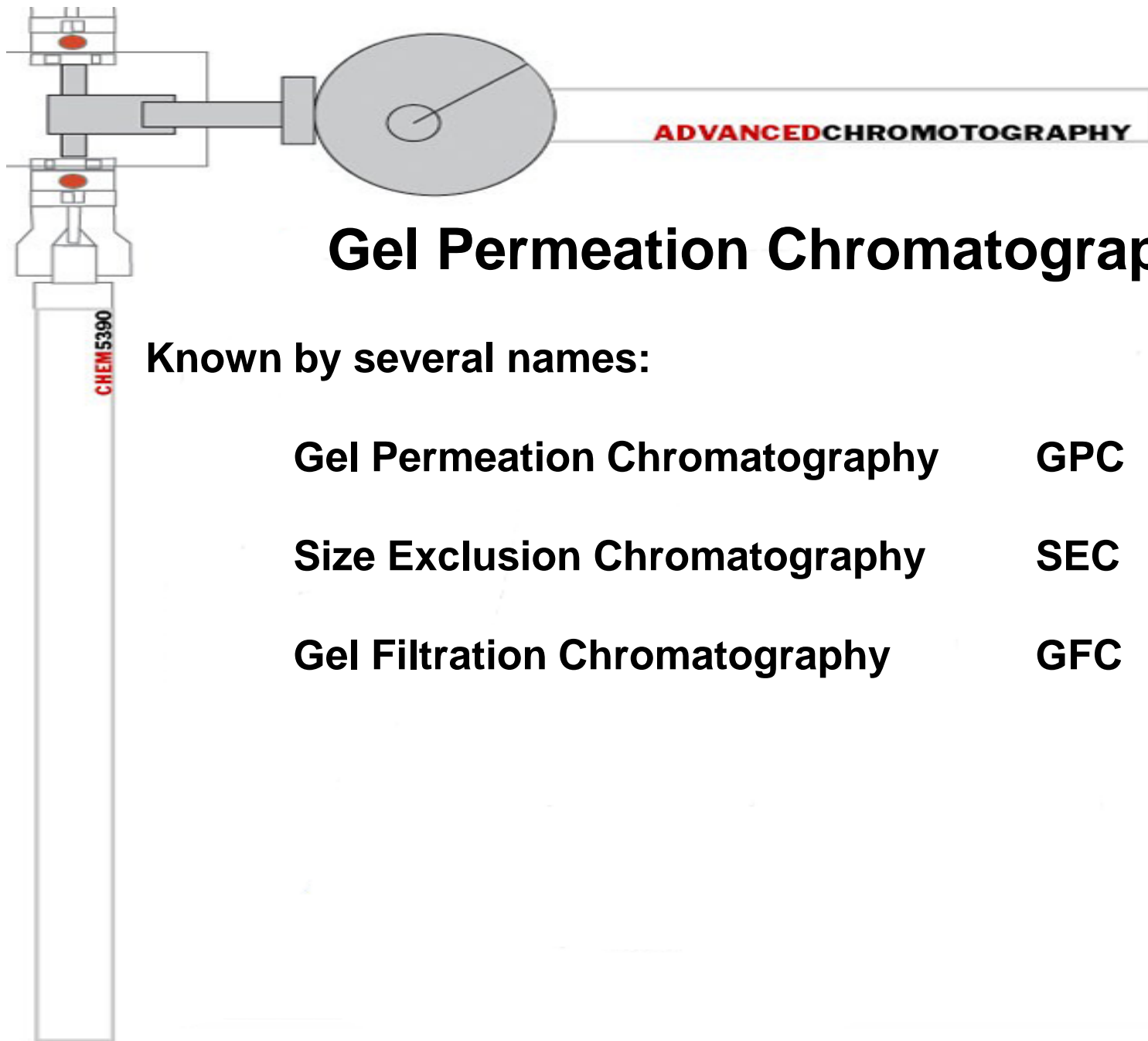
Traditional GPC

HPLC

GC

$10^1$   $10^2$   $10^3$   $10^4$   $10^5$   $10^6$   $10^7$

Molecular weight /  $\text{gmol}^{-1}$



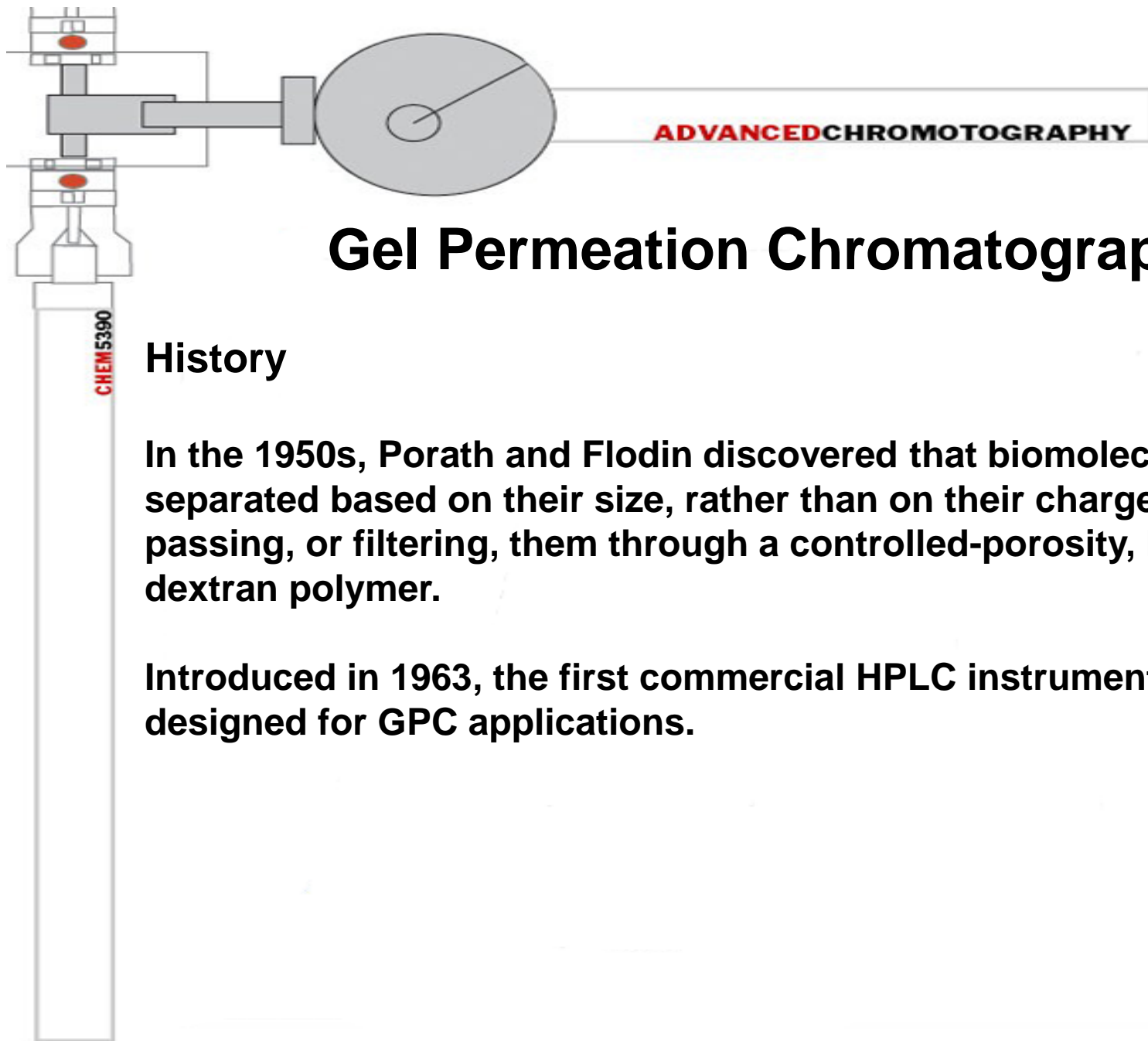
# Gel Permeation Chromatography

Known by several names:

Gel Permeation Chromatography	GPC
-------------------------------	-----

Size Exclusion Chromatography	SEC
-------------------------------	-----

Gel Filtration Chromatography	GFC
-------------------------------	-----

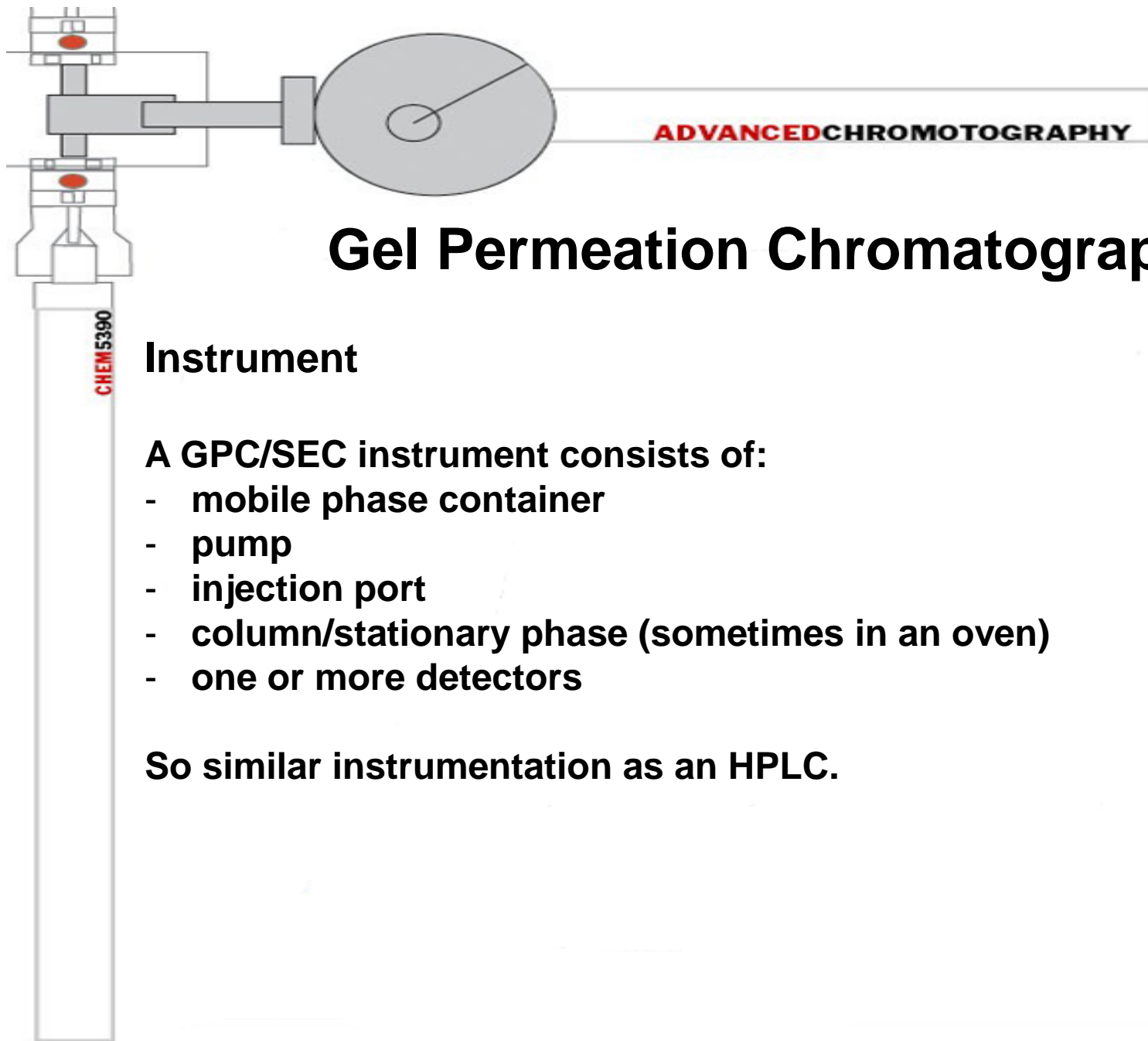


# Gel Permeation Chromatography

## History

In the 1950s, Porath and Flodin discovered that biomolecules could be separated based on their size, rather than on their charge or polarity, by passing, or filtering, them through a controlled-porosity, hydrophilic dextran polymer.

Introduced in 1963, the first commercial HPLC instruments were designed for GPC applications.



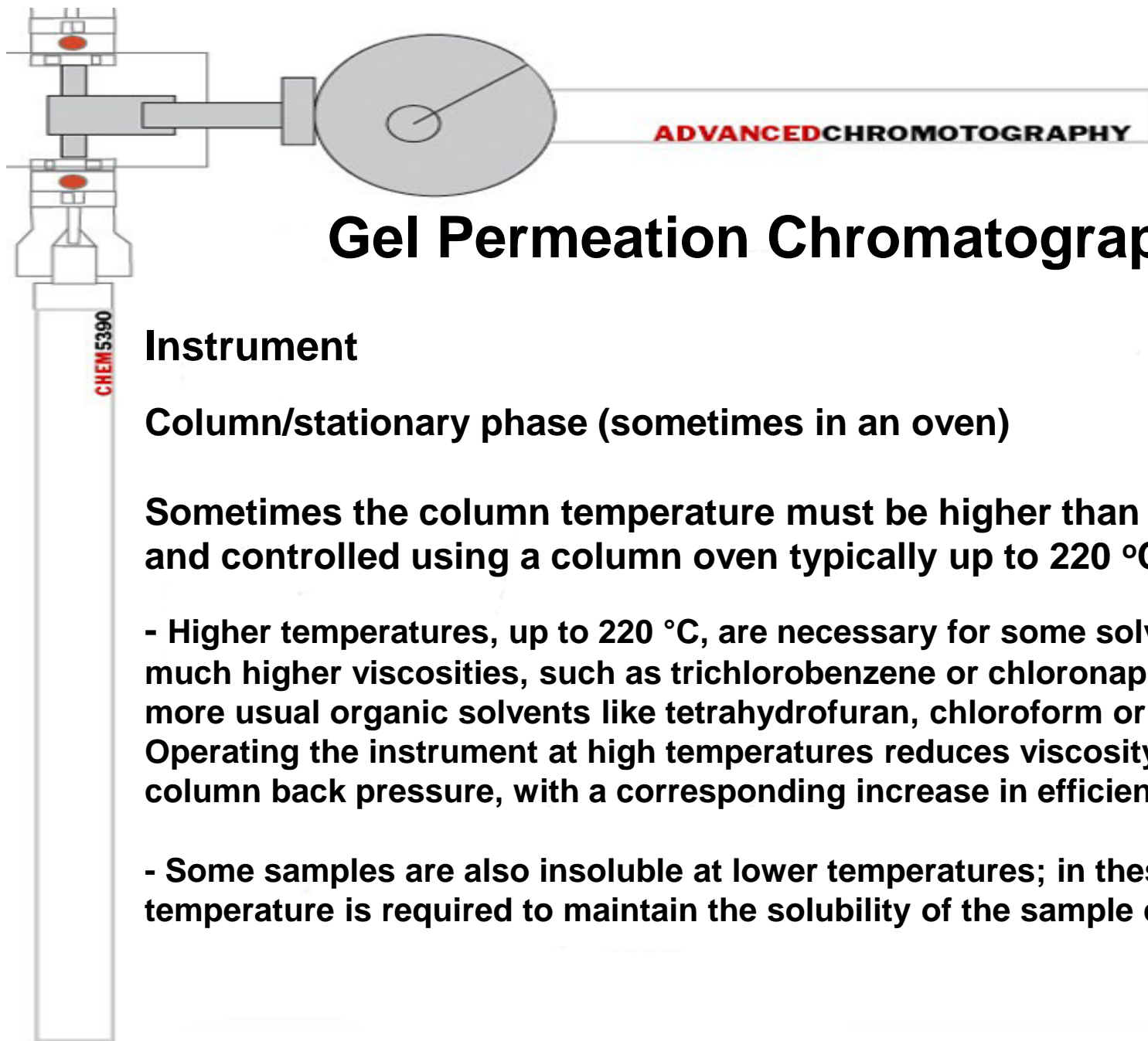
# Gel Permeation Chromatography

## Instrument

A GPC/SEC instrument consists of:

- mobile phase container
- pump
- injection port
- column/stationary phase (sometimes in an oven)
- one or more detectors

So similar instrumentation as an HPLC.



# Gel Permeation Chromatography

## Instrument

Column/stationary phase (sometimes in an oven)

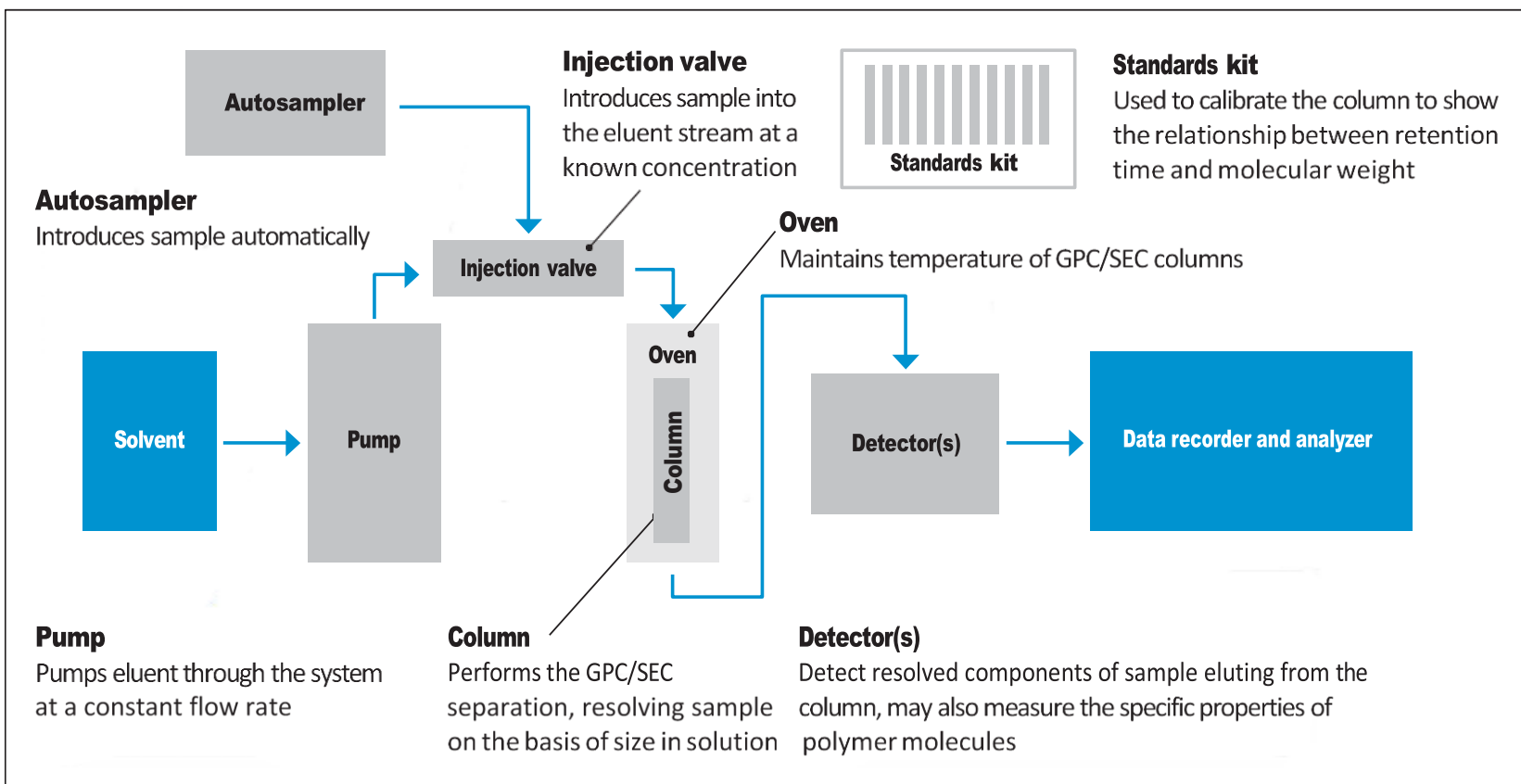
Sometimes the column temperature must be higher than room temp and controlled using a column oven typically up to 220 °C:

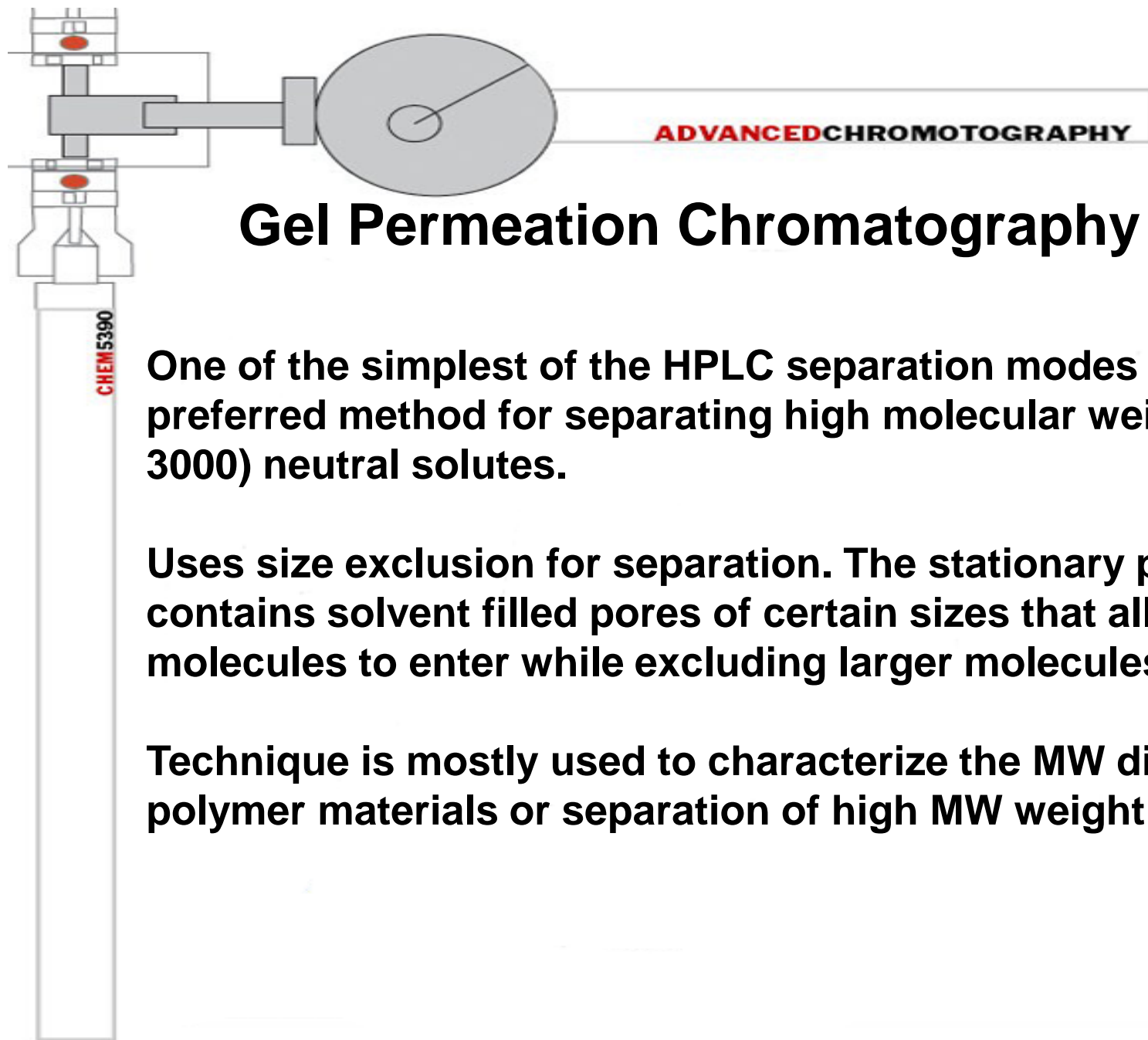
- Higher temperatures, up to 220 °C, are necessary for some solvents that have much higher viscosities, such as trichlorobenzene or chloronaphthalene, than more usual organic solvents like tetrahydrofuran, chloroform or toluene. Operating the instrument at high temperatures reduces viscosity and hence column back pressure, with a corresponding increase in efficiency.
- Some samples are also insoluble at lower temperatures; in these cases high temperature is required to maintain the solubility of the sample during analysis.



# Gel Permeation Chromatography

## Instrument



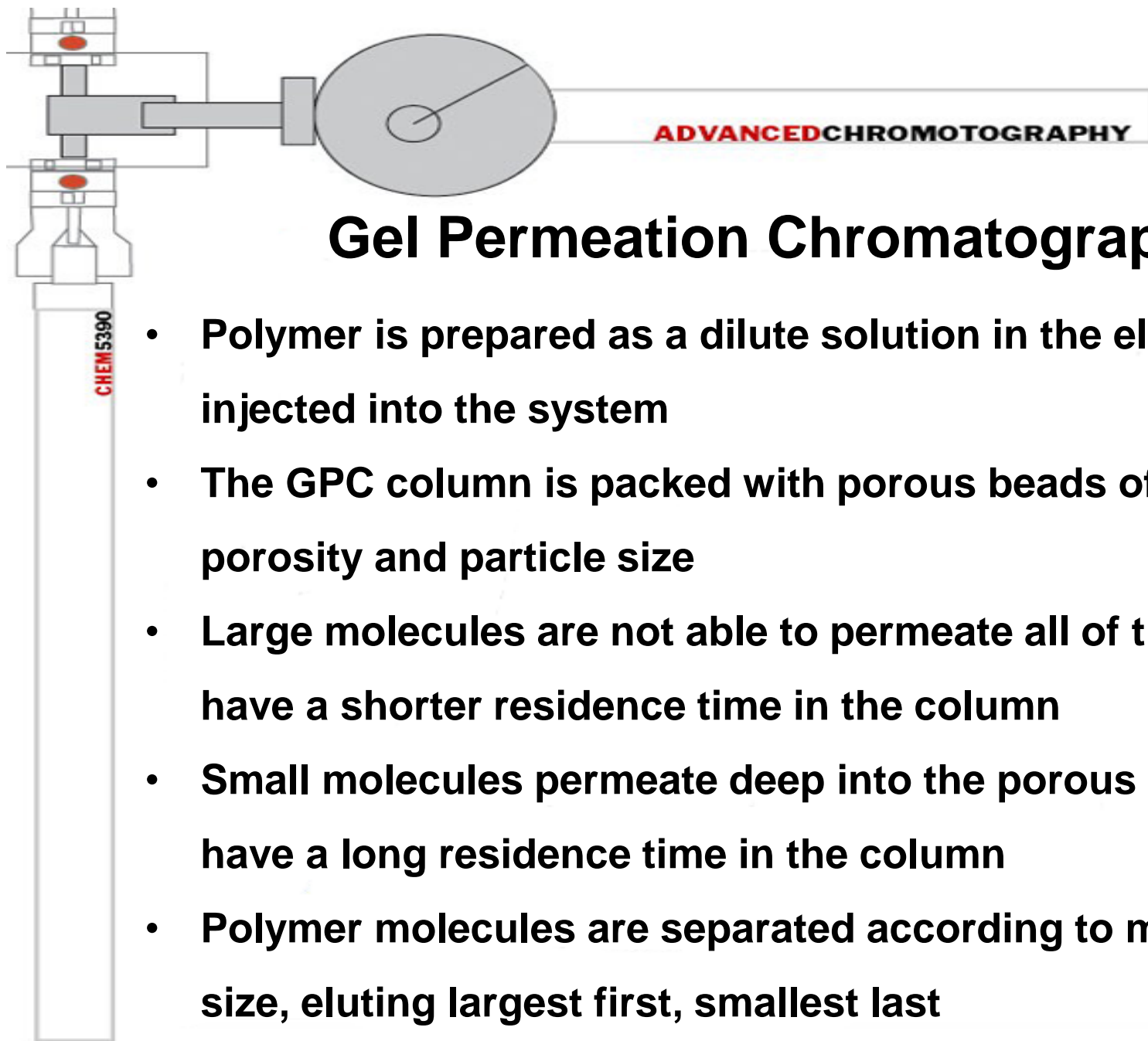


## **Gel Permeation Chromatography (GPC)**

**One of the simplest of the HPLC separation modes and is the preferred method for separating high molecular weight (>2000-3000) neutral solutes.**

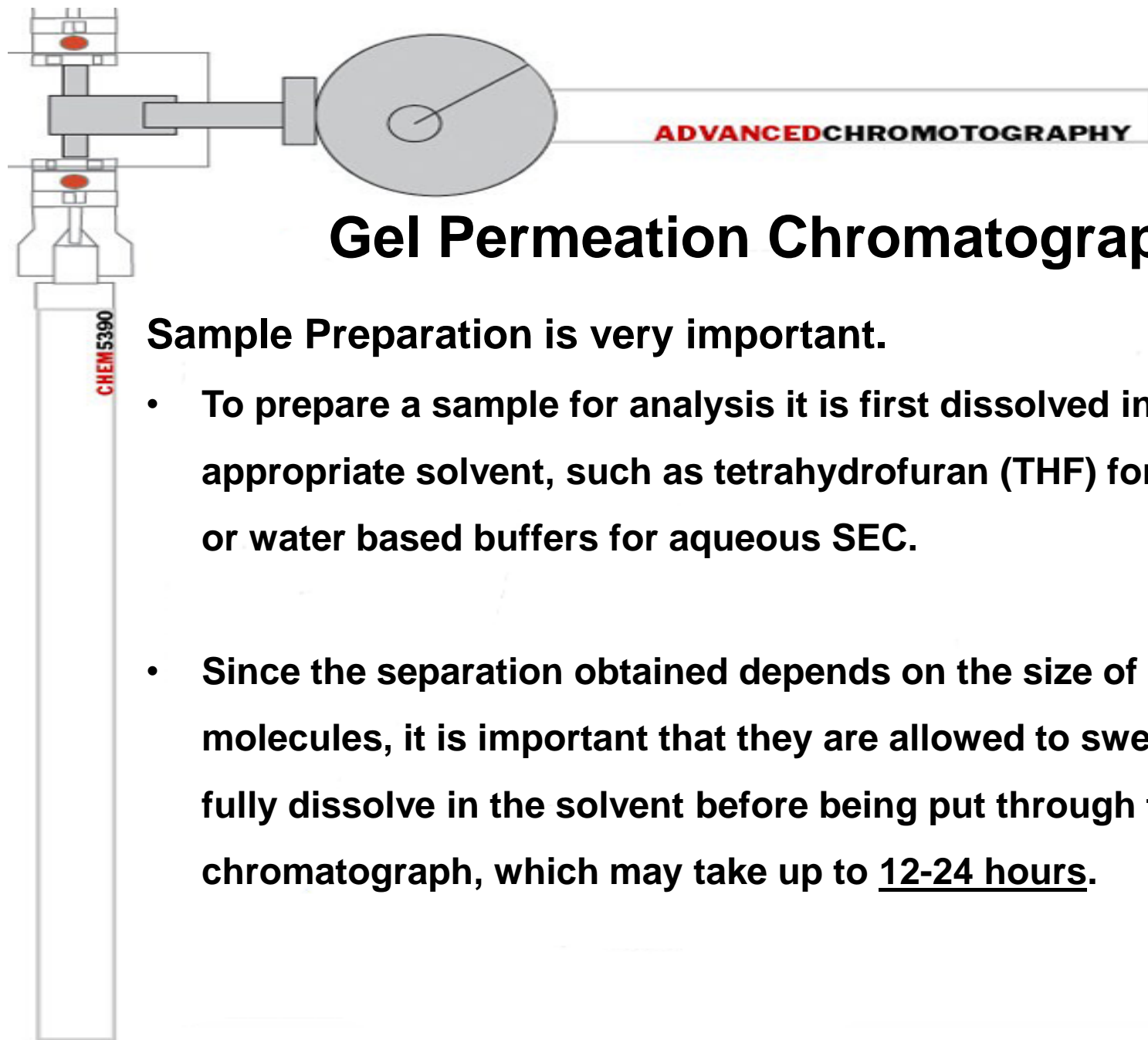
**Uses size exclusion for separation. The stationary phase contains solvent filled pores of certain sizes that allow smaller molecules to enter while excluding larger molecules.**

**Technique is mostly used to characterize the MW distribution of polymer materials or separation of high MW weight proteins.**



## Gel Permeation Chromatography

- Polymer is prepared as a dilute solution in the eluent and injected into the system
- The GPC column is packed with porous beads of controlled porosity and particle size
- Large molecules are not able to permeate all of the pores and have a shorter residence time in the column
- Small molecules permeate deep into the porous matrix and have a long residence time in the column
- Polymer molecules are separated according to molecular size, eluting largest first, smallest last

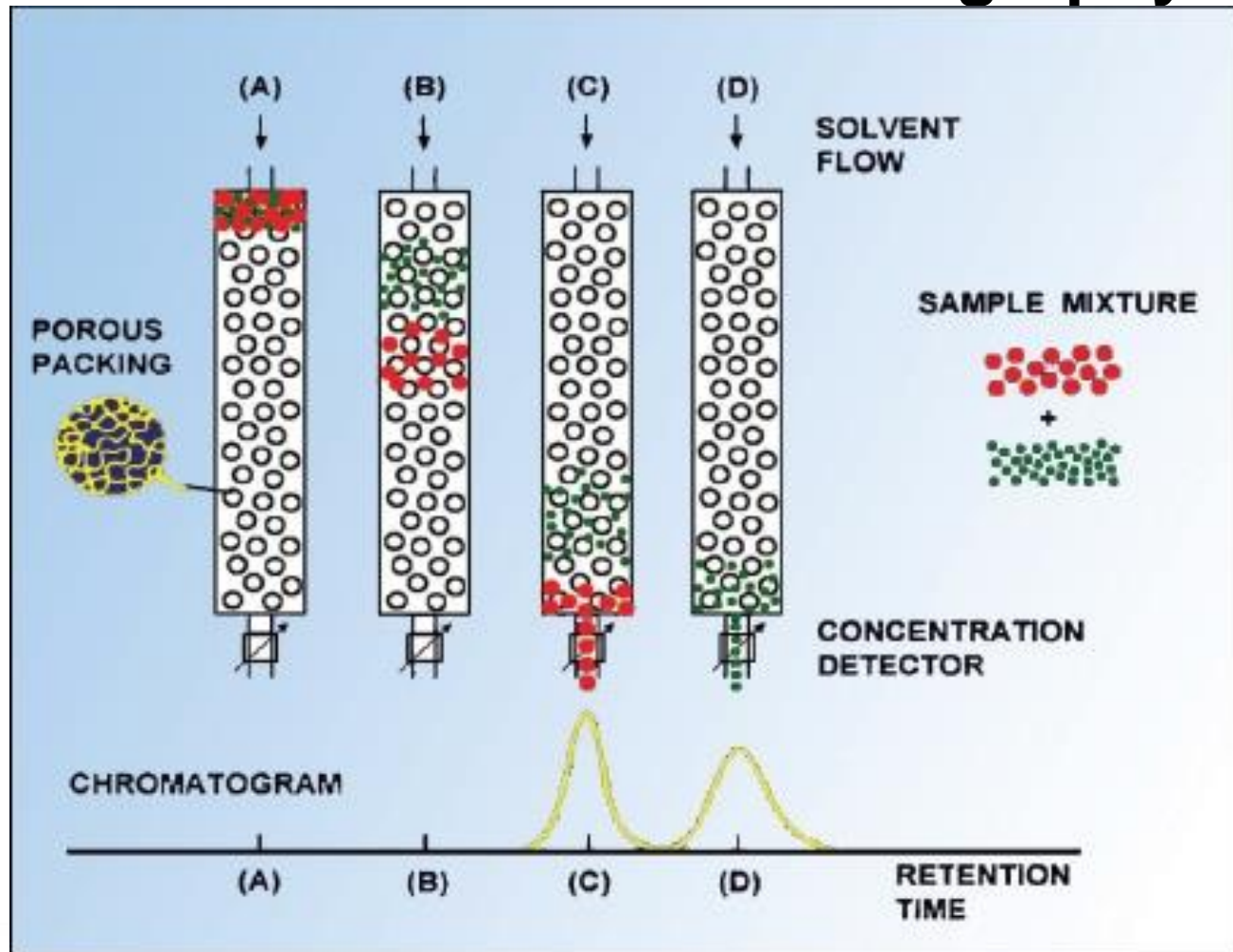


# Gel Permeation Chromatography

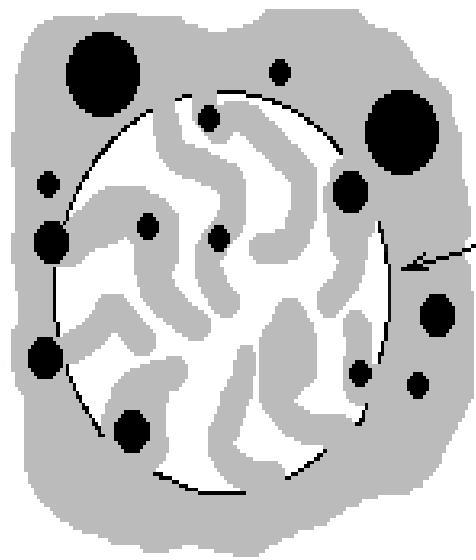
**Sample Preparation is very important.**

- To prepare a sample for analysis it is first dissolved in an appropriate solvent, such as tetrahydrofuran (THF) for organic GPC or water based buffers for aqueous SEC.
- Since the separation obtained depends on the size of the sample molecules, it is important that they are allowed to swell and then fully dissolve in the solvent before being put through the chromatograph, which may take up to 12-24 hours.

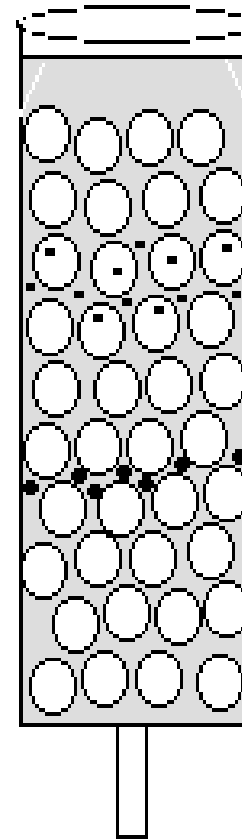
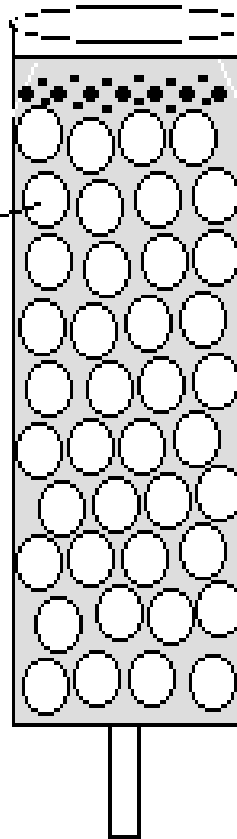
# Gel Permeation Chromatography



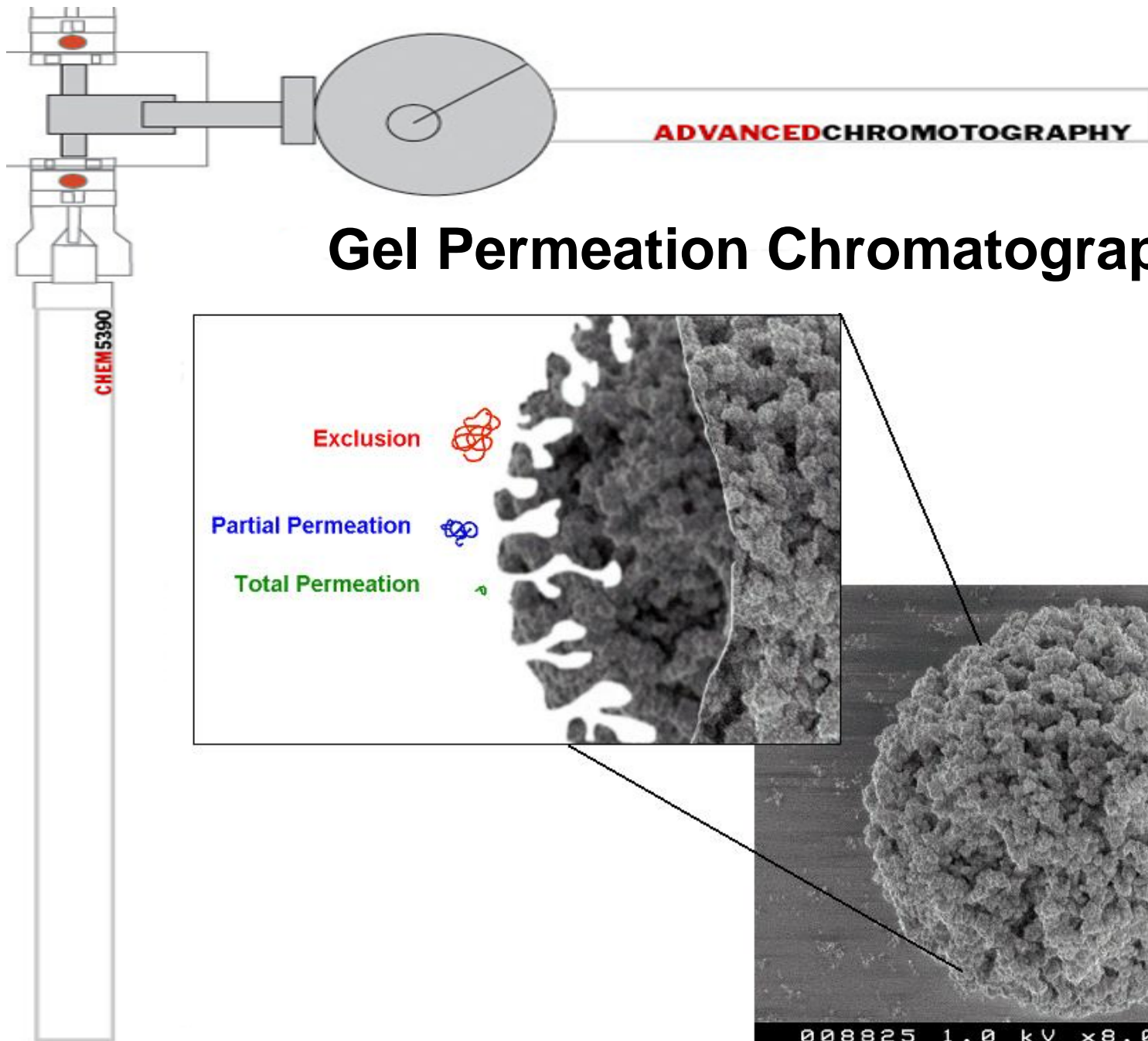
# Gel Permeation Chromatography

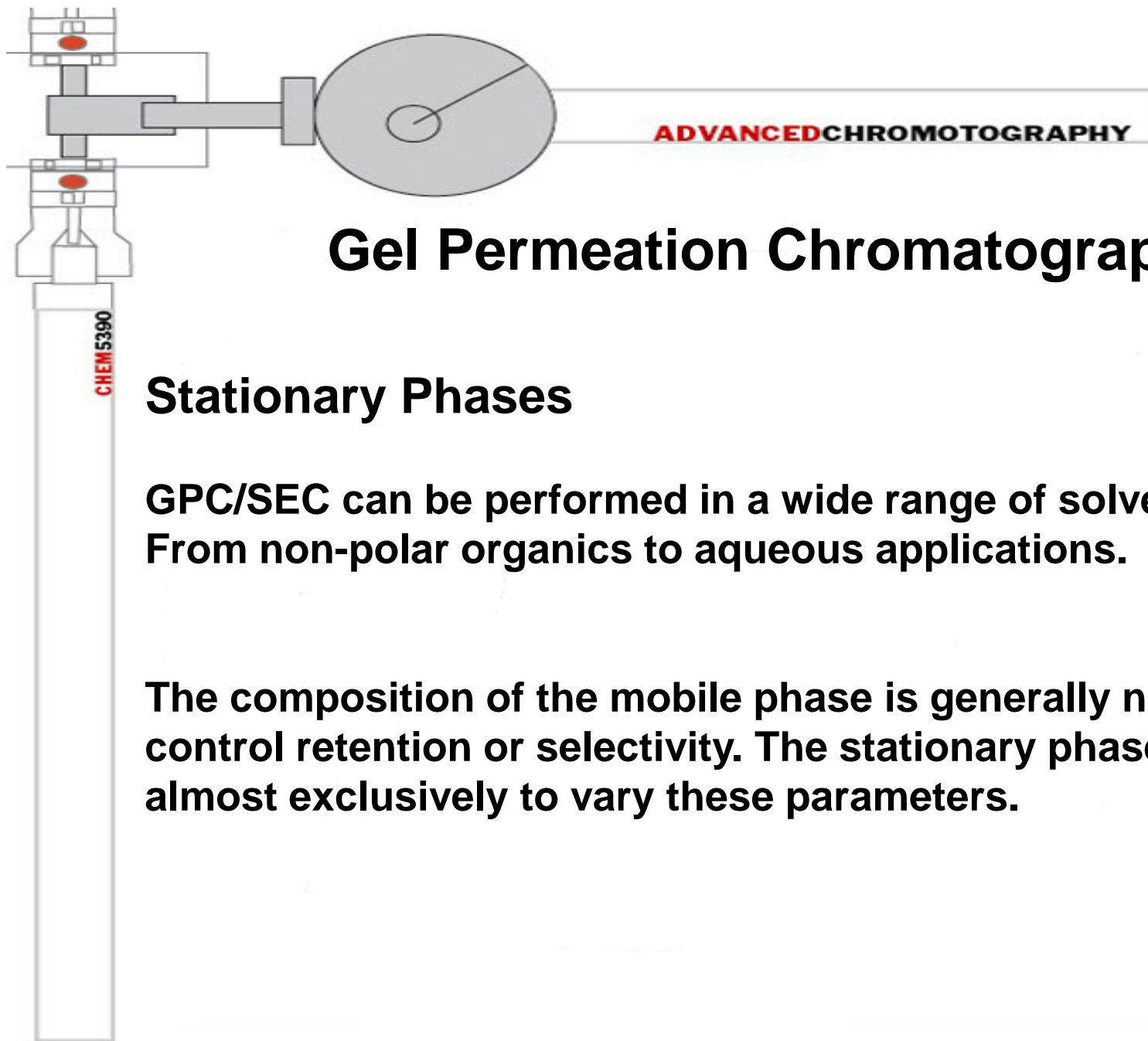


Gel beads have pores in them of a defined size range which allows smaller molecules to enter but excludes molecules larger than the pore diameters.



- ← Molecules smaller than gel bead pores
- ← Molecules larger than gel bead pores





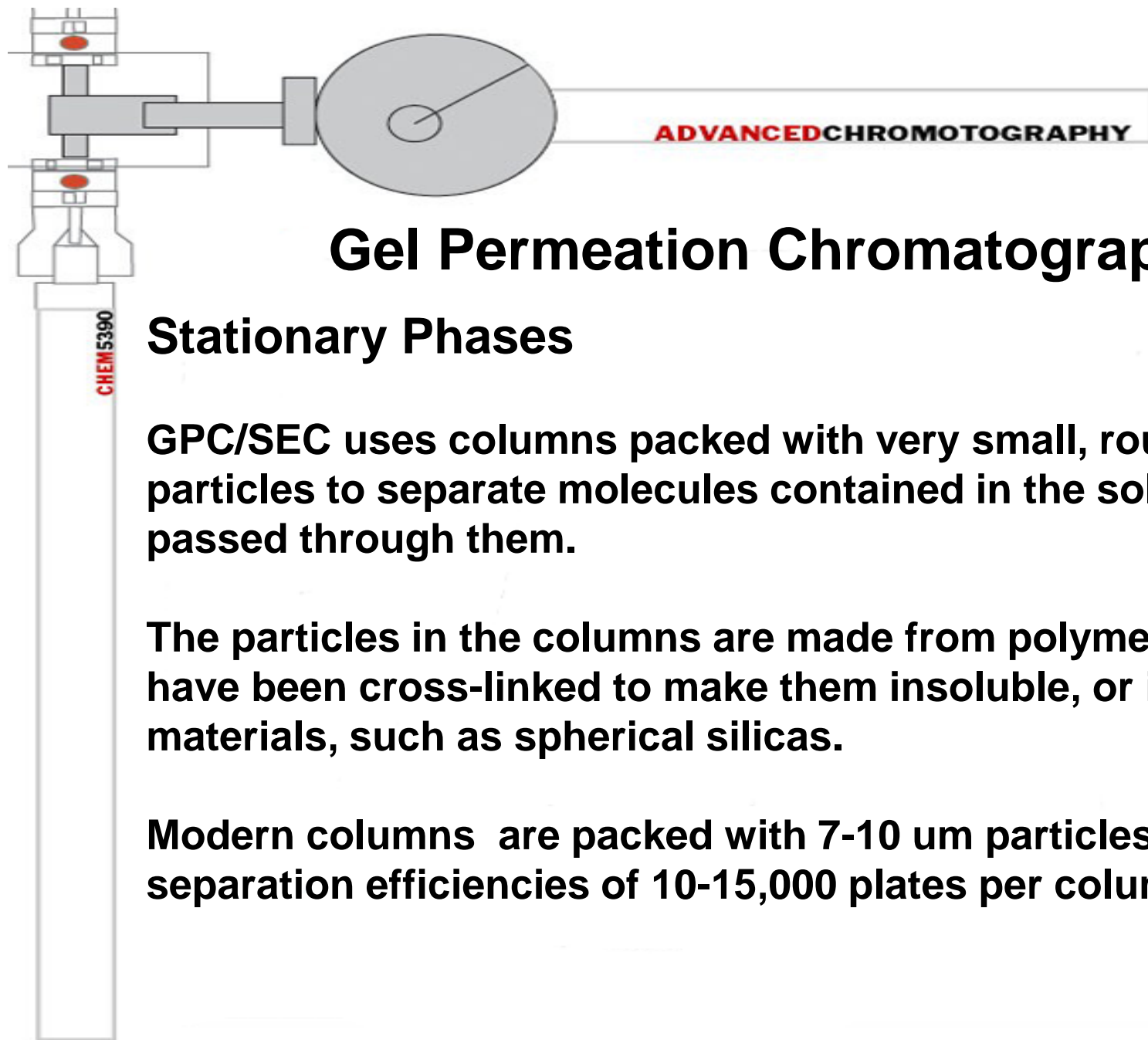
# Gel Permeation Chromatography

## Stationary Phases

GPC/SEC can be performed in a wide range of solvents. From non-polar organics to aqueous applications.

The composition of the mobile phase is generally not used to control retention or selectivity. The stationary phase is used almost exclusively to vary these parameters.





# Gel Permeation Chromatography

## Stationary Phases

GPC/SEC uses columns packed with very small, round, porous particles to separate molecules contained in the solvent that is passed through them.

The particles in the columns are made from polymers that have been cross-linked to make them insoluble, or inorganic materials, such as spherical silicas.

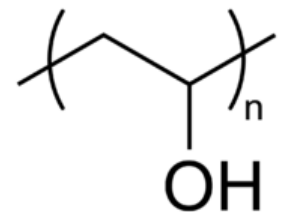
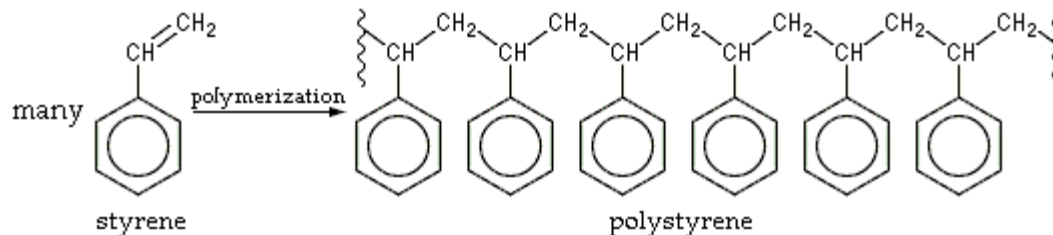
Modern columns are packed with 7-10  $\mu\text{m}$  particles and have separation efficiencies of 10-15,000 plates per column.

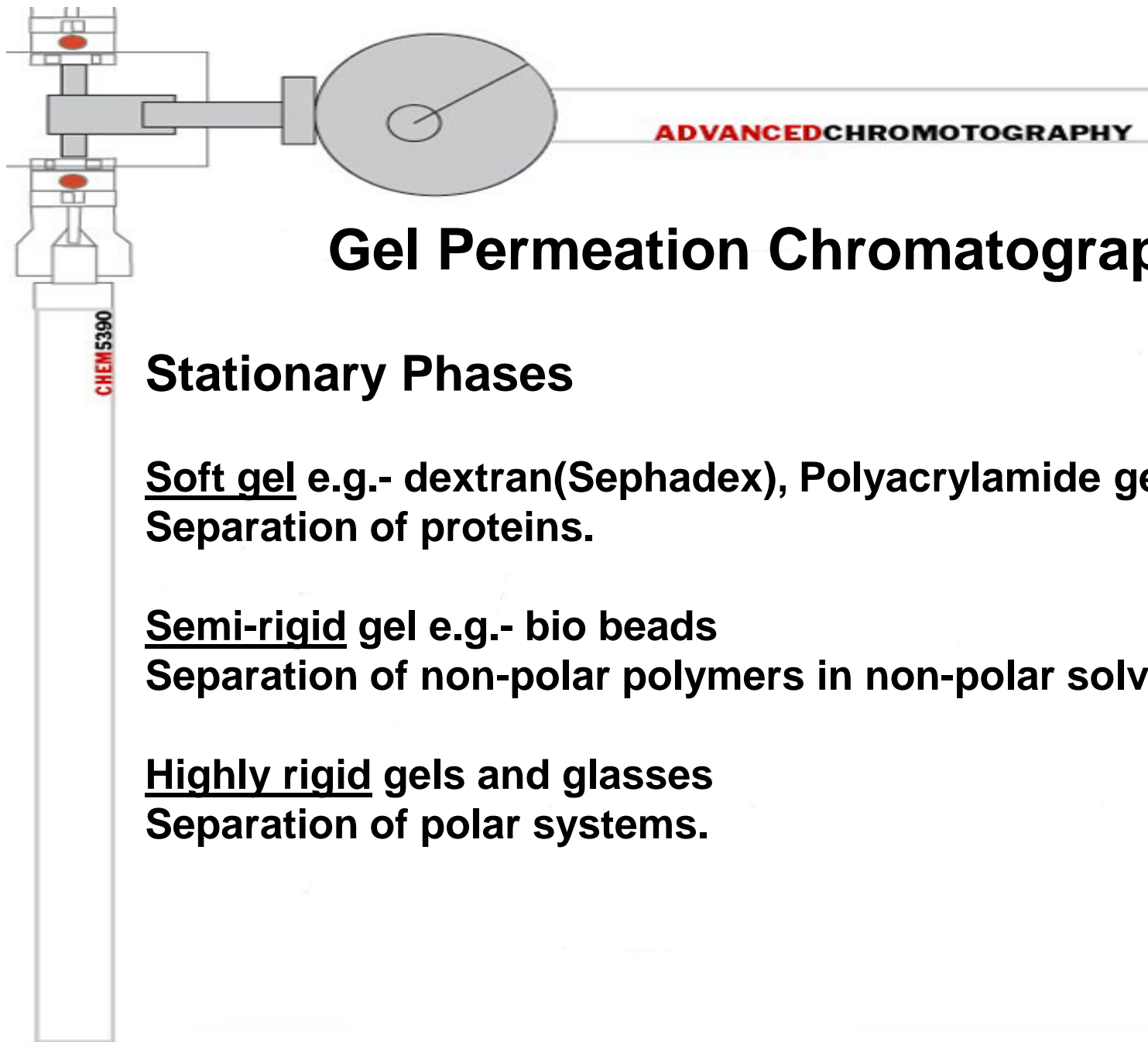
# Gel Permeation Chromatography

## Stationary Phases

Divided into two classes:

- cross-linked, rigid polystyrene divinylbenzene gels (Shodex)
- polydextrans, polyvinylalcohol, hydroxylated methacrylate, and silica gels





# Gel Permeation Chromatography

## Stationary Phases

Soft gel e.g.- dextran(Sephadex), Polyacrylamide gels  
Separation of proteins.

Semi-rigid gel e.g.- bio beads  
Separation of non-polar polymers in non-polar solvents.

Highly rigid gels and glasses  
Separation of polar systems.



# Gel Permeation Chromatography

## Stationary Phases

### Dextran

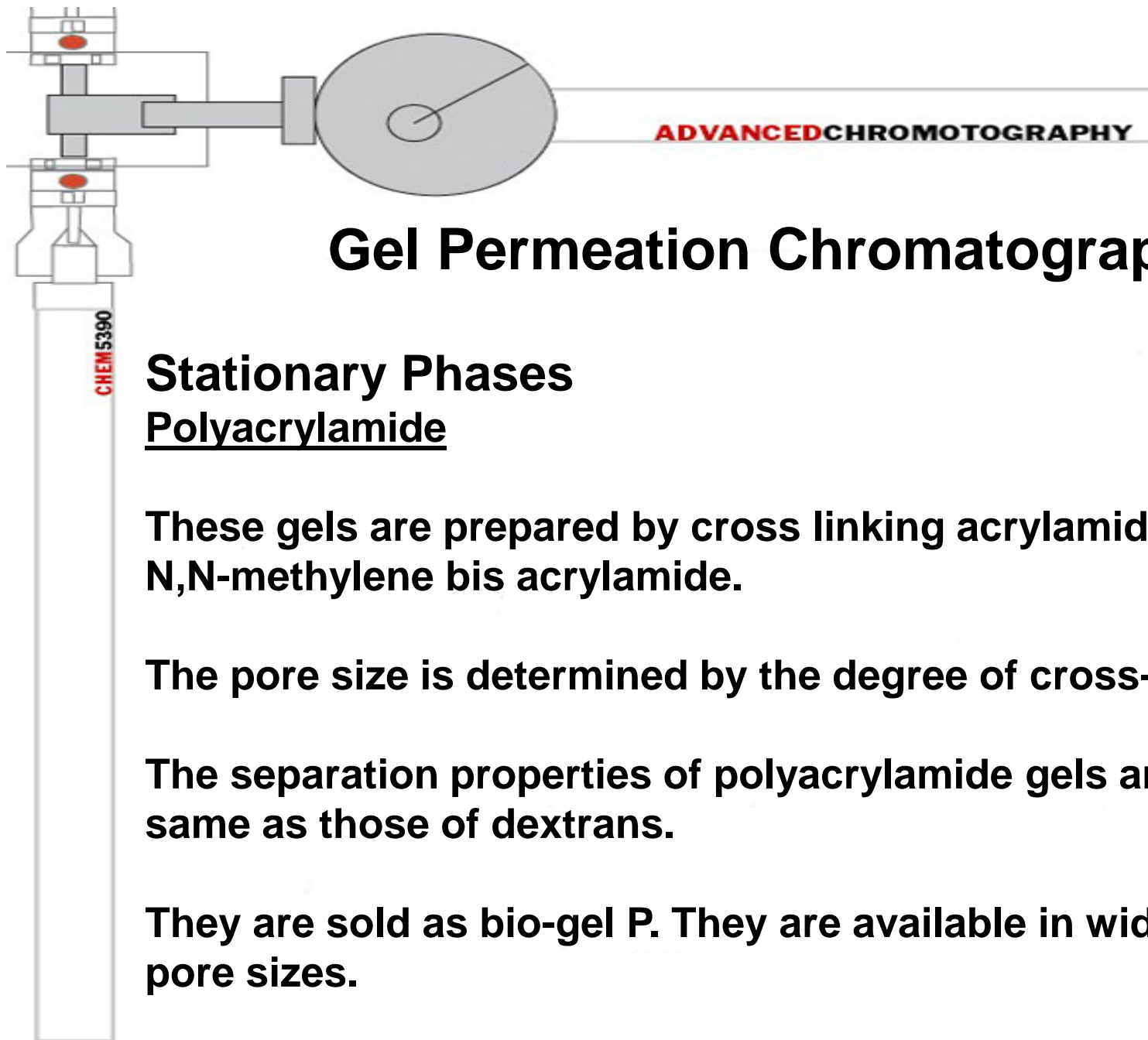
A homopolysaccharide of glucose residues.

Prepared with various degrees of cross-linking to control pore size.

Bought as dry beads, the beads swell when water is added.

The trade name is sephadex.

Mainly used for separation of small peptides and globular proteins with small to average molecular mass



# Gel Permeation Chromatography

## Stationary Phases

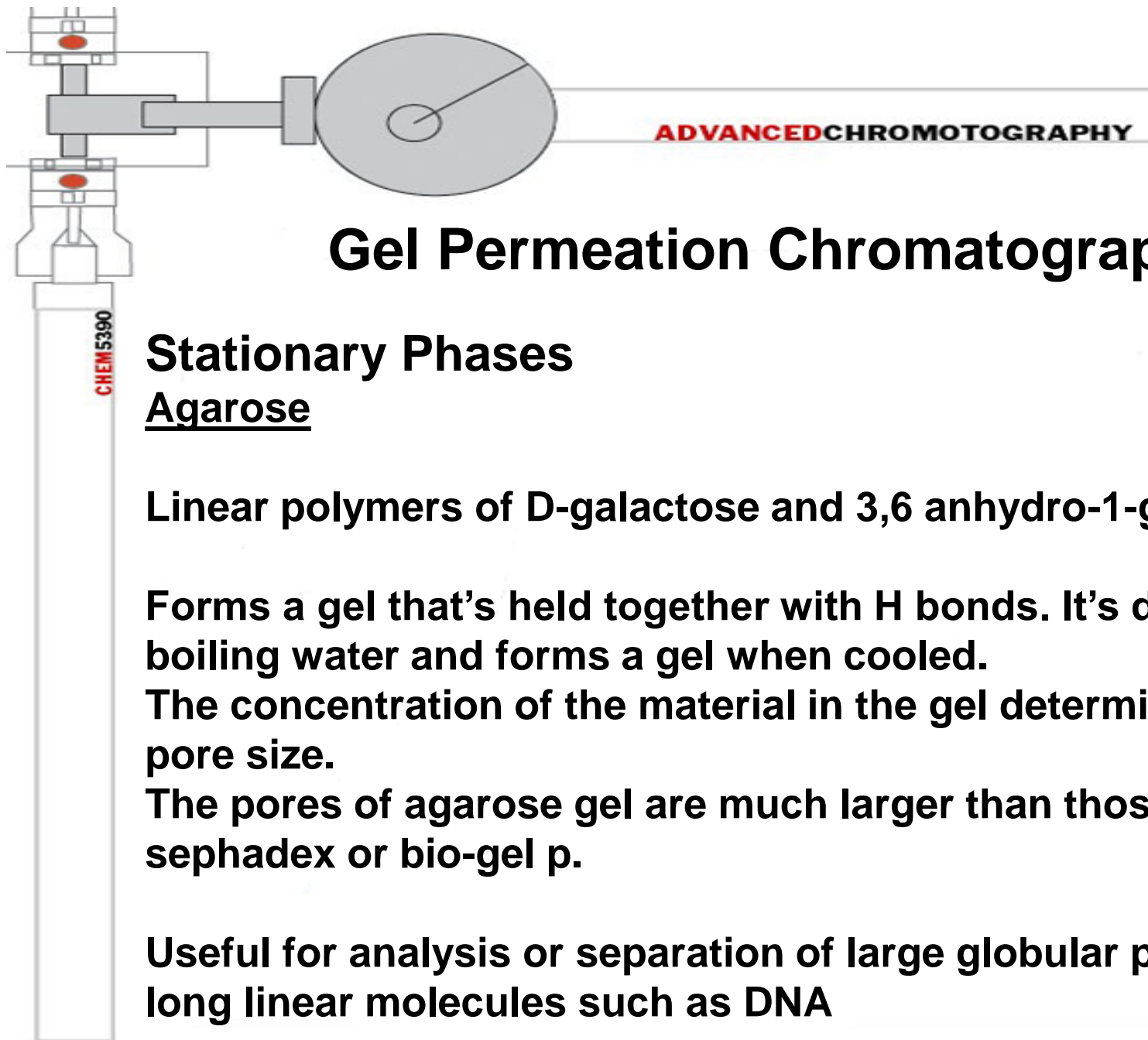
### Polyacrylamide

These gels are prepared by cross linking acrylamide with N,N-methylene bis acrylamide.

The pore size is determined by the degree of cross-linking.

The separation properties of polyacrylamide gels are mainly the same as those of dextrans.

They are sold as bio-gel P. They are available in wide range of pore sizes.



# Gel Permeation Chromatography

## Stationary Phases

### Agarose

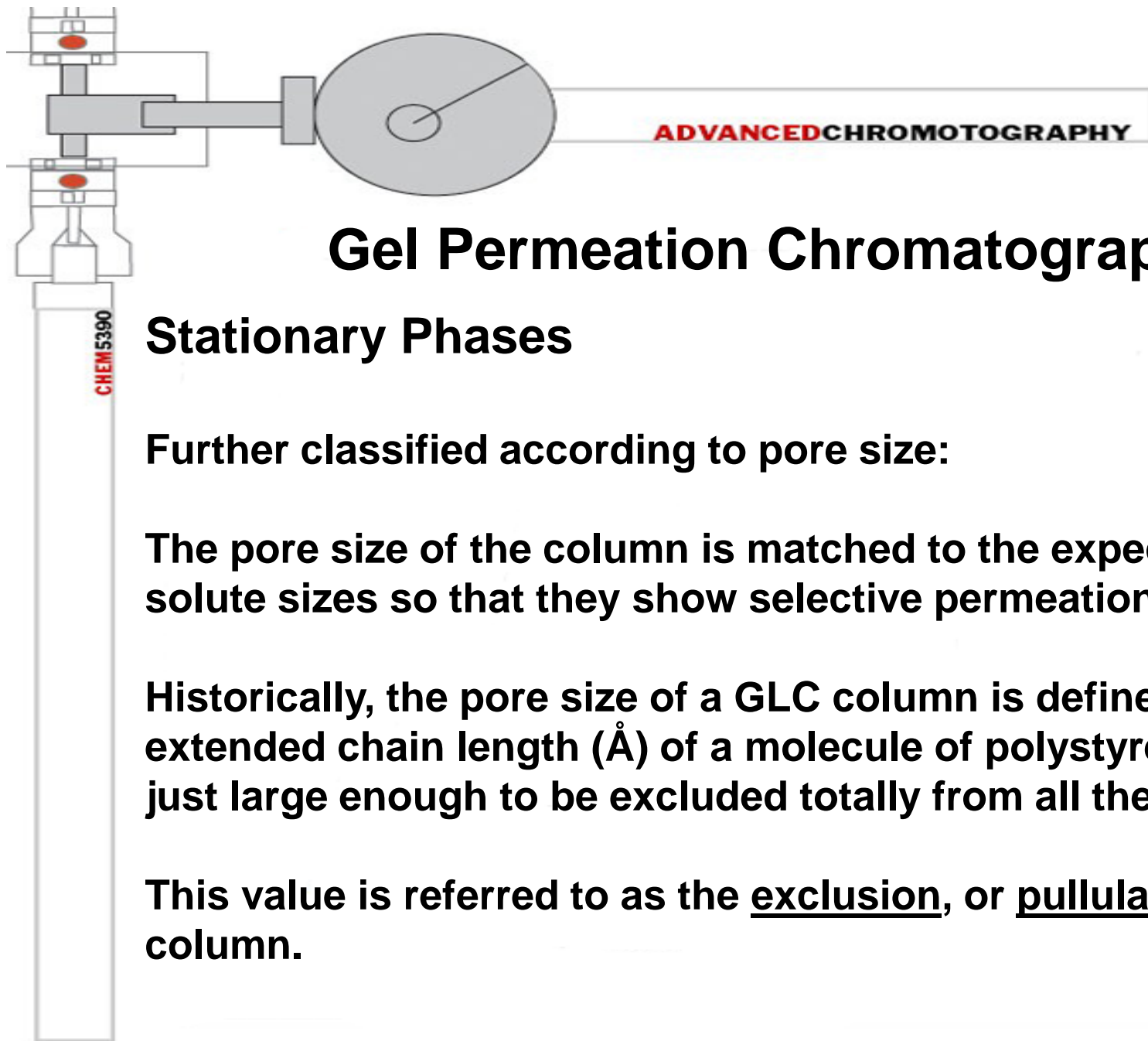
Linear polymers of D-galactose and 3,6 anhydro-1-galactose.

Forms a gel that's held together with H bonds. It's dissolved in boiling water and forms a gel when cooled.

The concentration of the material in the gel determines the pore size.

The pores of agarose gel are much larger than those of sephadex or bio-gel p.

Useful for analysis or separation of large globular proteins or long linear molecules such as DNA



# Gel Permeation Chromatography

## Stationary Phases

Further classified according to pore size:

The pore size of the column is matched to the expected range of solute sizes so that they show selective permeation on the gel.

Historically, the pore size of a GLC column is defined as the extended chain length ( $\text{\AA}$ ) of a molecule of polystyrene which is just large enough to be excluded totally from all the pores.

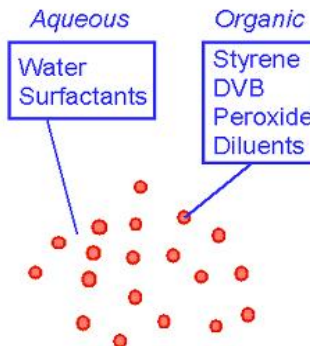
This value is referred to as the exclusion, or pullulan, limit of the column.

# Gel Permeation Chromatography

## Synthesis of Stationary Phase

High cross-link content gives a rigid, low swelling product with a well-defined pore structure

### 2 PHASE SYSTEM



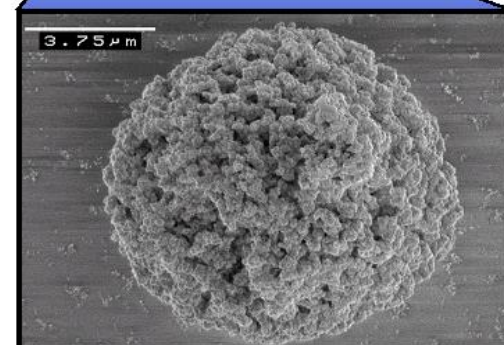
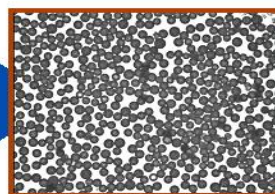
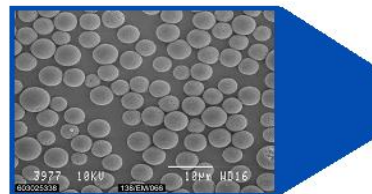
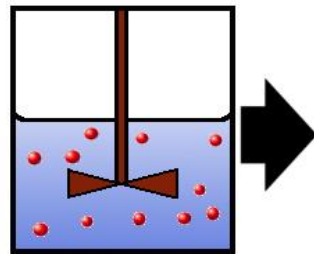
### MICROSPHERE FORMATION & FUSION

*Porous particle*

### PARTICLE SIZING

*Refine particle size distribution*

3 $\mu$ m  
5 $\mu$ m  
10 $\mu$ m  
20 $\mu$ m



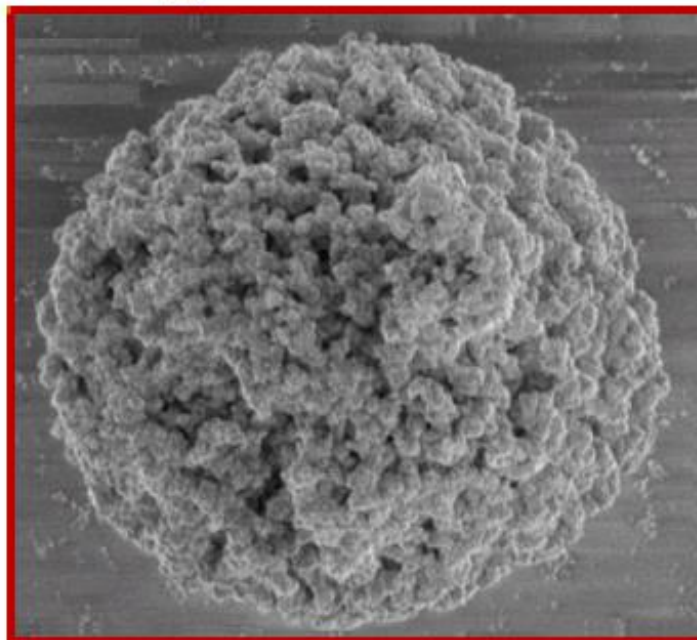




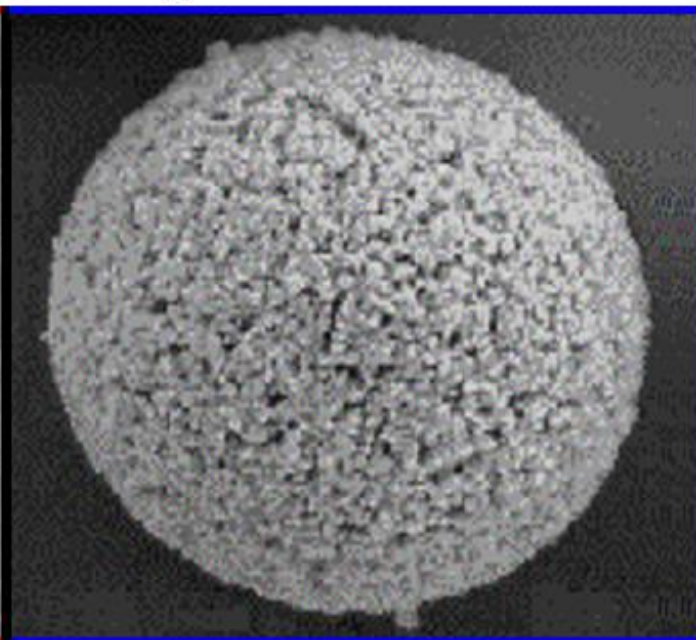
# Gel Permeation Chromatography

## Synthesis of Stationary Phase

PLgel 10  $\mu\text{m}$   $10^6\text{\AA}$



PLgel 10  $\mu\text{m}$   $10^3\text{\AA}$



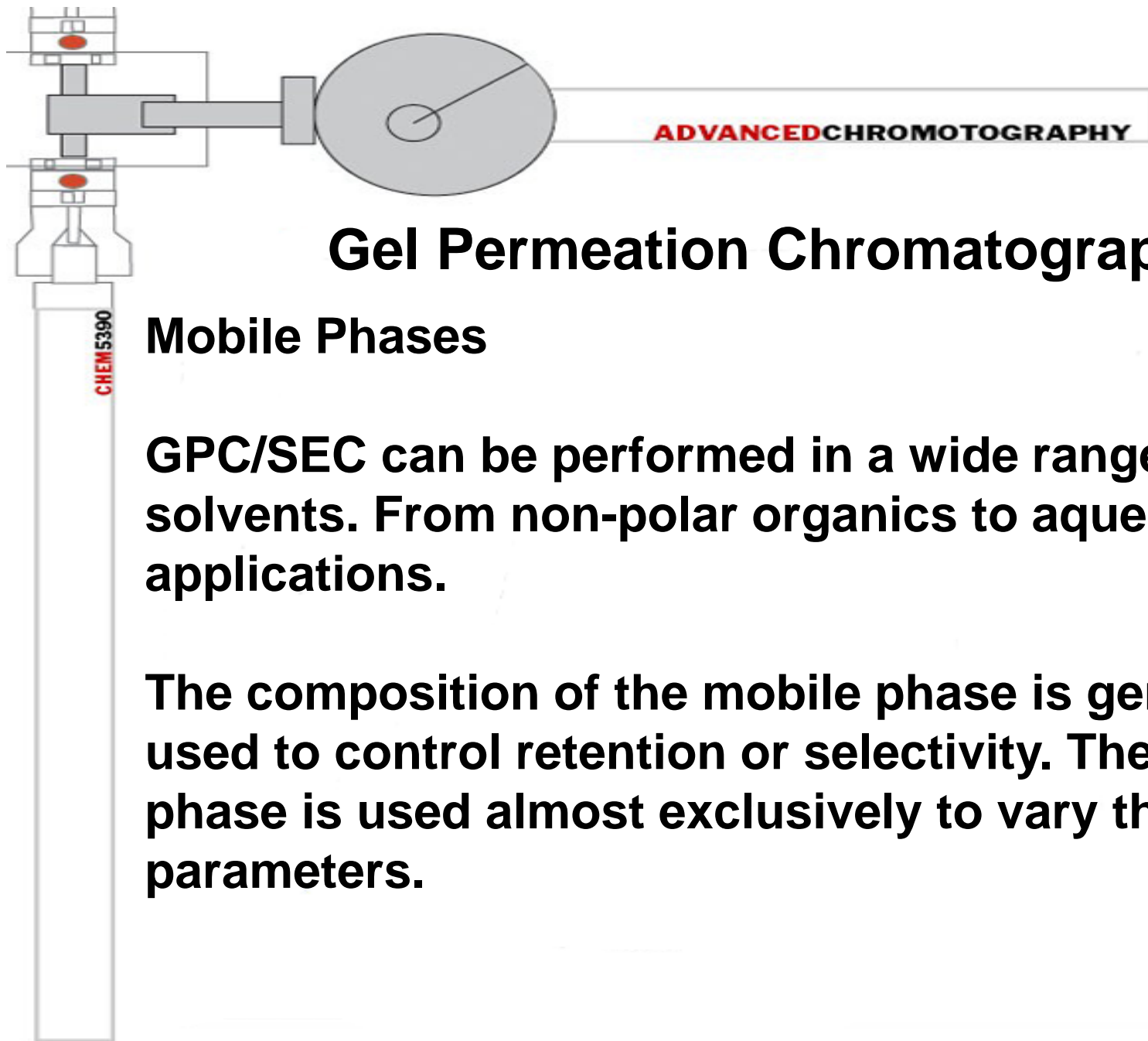


## ADVANCED CHROMATOGRAPHY

**Table 6.9** Characteristics of non-aqueous (upper) and aqueous (lower) GPC column ranges available commercially from Waters.

Column range and individual designations	Packing material	Column range exclusion limits (lowest to highest)	Solvent(s)
<b>Nonaqueous columns</b>			
Ultrastaygel 100, 500, $10^3$ , $10^4$ , $10^5$ , $10^6$ , linear	PS-DVB	500 up to $10^7$	THF, toluene
$\mu$ Styragel 100, 500, $10^3$ , $10^4$ , $10^5$ , $10^6$ , linear	PS-DVB	500 up to $10^7$	Toluene
$\mu$ Styragel HT $10^3$ , $10^4$ , $10^5$ , $10^6$ , linear	PS-DVB	30 000 up to $10^7$	MEK
Styragel 60, 100, 200, 500, $10^3$ , $10^4$ , $10^5$ , $10^6$ , $10^7$	PS-DVB	500 up to $2 \times 10^7$	Toluene
Shodex K series 801, 802, 802.5, 803, 804, 804L, 805, 806, 805, 806, 807, 80-linear	PS-DVB	$1.5 \times 10^3$ up to $2 \times 10^8$	THF, DMF, chloroform
Shodex HFIP series 803, 804, 805, 806, 807, 80-linear	PS-DVB	$7 \times 10^4$ up to $2 \times 10^8$	HFIP
Shodex AT series 803, 804, 805, 806, 807, 80-linear	PS-DVB	$7 \times 10^4$ up to $2 \times 10^8$	Toluene
<b>Aqueous columns</b>			
Shodex OHpak B series 803, 804, 805, 806	Methacrylate	$10^5$ up to $2 \times 10^7$	Water
Shodex OHpak Q series 803, 804, 805, 806	Polyvinylalcohol	$10^5$ up to $2 \times 10^7$	Water
Ultrahydrogel 120, 250, 500, 1000, 2000, linear, DP	Methacrylate	$5 \times 10^3$ up to $7 \times 10^6$	Water
Protein-Pak 60, 125, 300SW	Silica gel	$2 \times 10^4$ up to $3 \times 10^5$	Propanol
Shodex Protein KW 802.5, 803, 804	Diol-bonded silica gel	$5 \times 10^4$ up to $6 \times 10^5$	Water

PS-DVB, polystyrene divinylbenzene; THF, tetrahydrofuran; MEK, methylethyl ketone; DMF, dimethylformamide; HFIP, hexafluoroisopropanone.

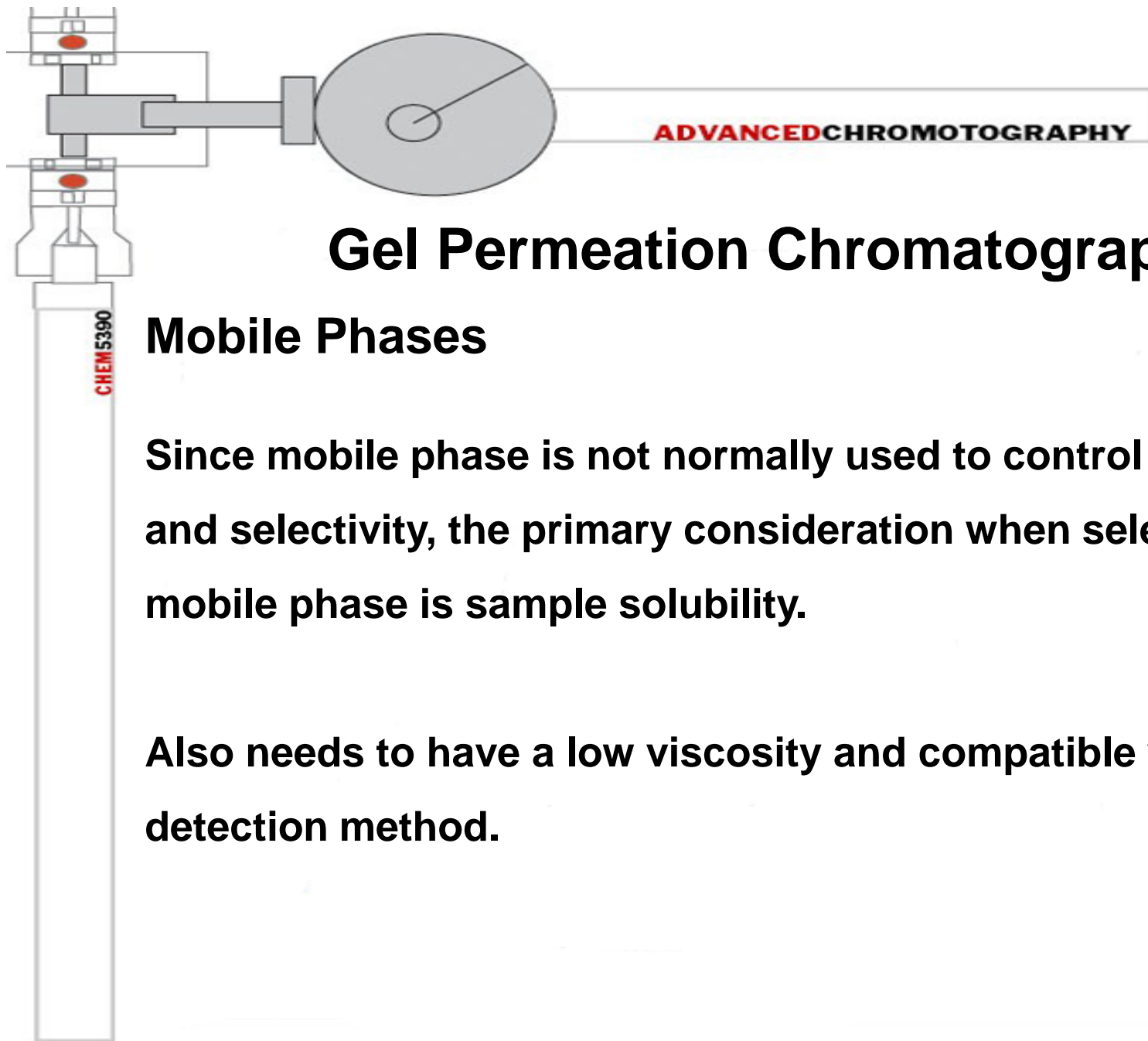


# **Gel Permeation Chromatography**

## **Mobile Phases**

**GPC/SEC can be performed in a wide range of solvents. From non-polar organics to aqueous applications.**

**The composition of the mobile phase is generally not used to control retention or selectivity. The stationary phase is used almost exclusively to vary these parameters.**

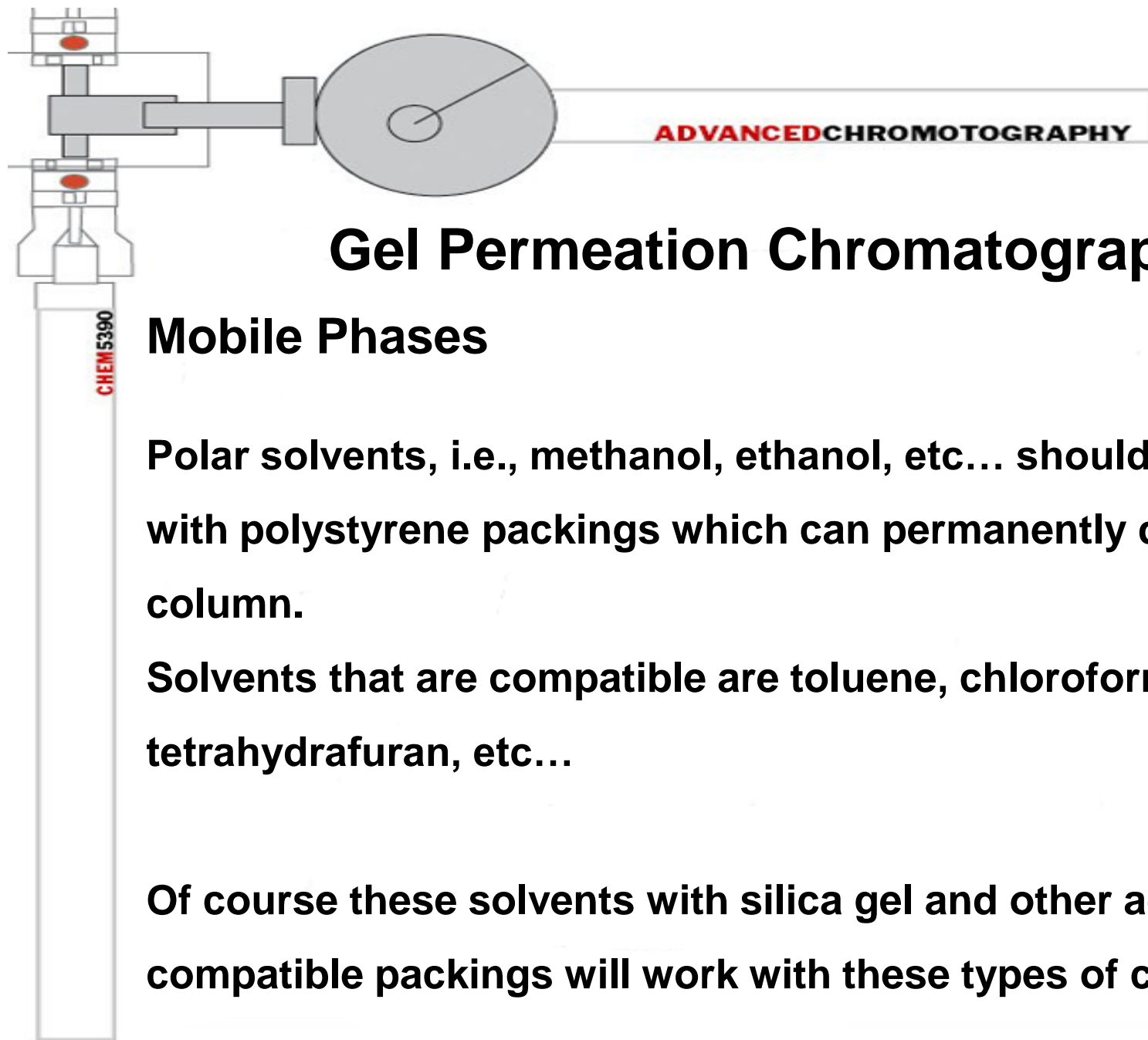


# **Gel Permeation Chromatography**

## **Mobile Phases**

**Since mobile phase is not normally used to control retention and selectivity, the primary consideration when selecting a mobile phase is sample solubility.**

**Also needs to have a low viscosity and compatible with the detection method.**



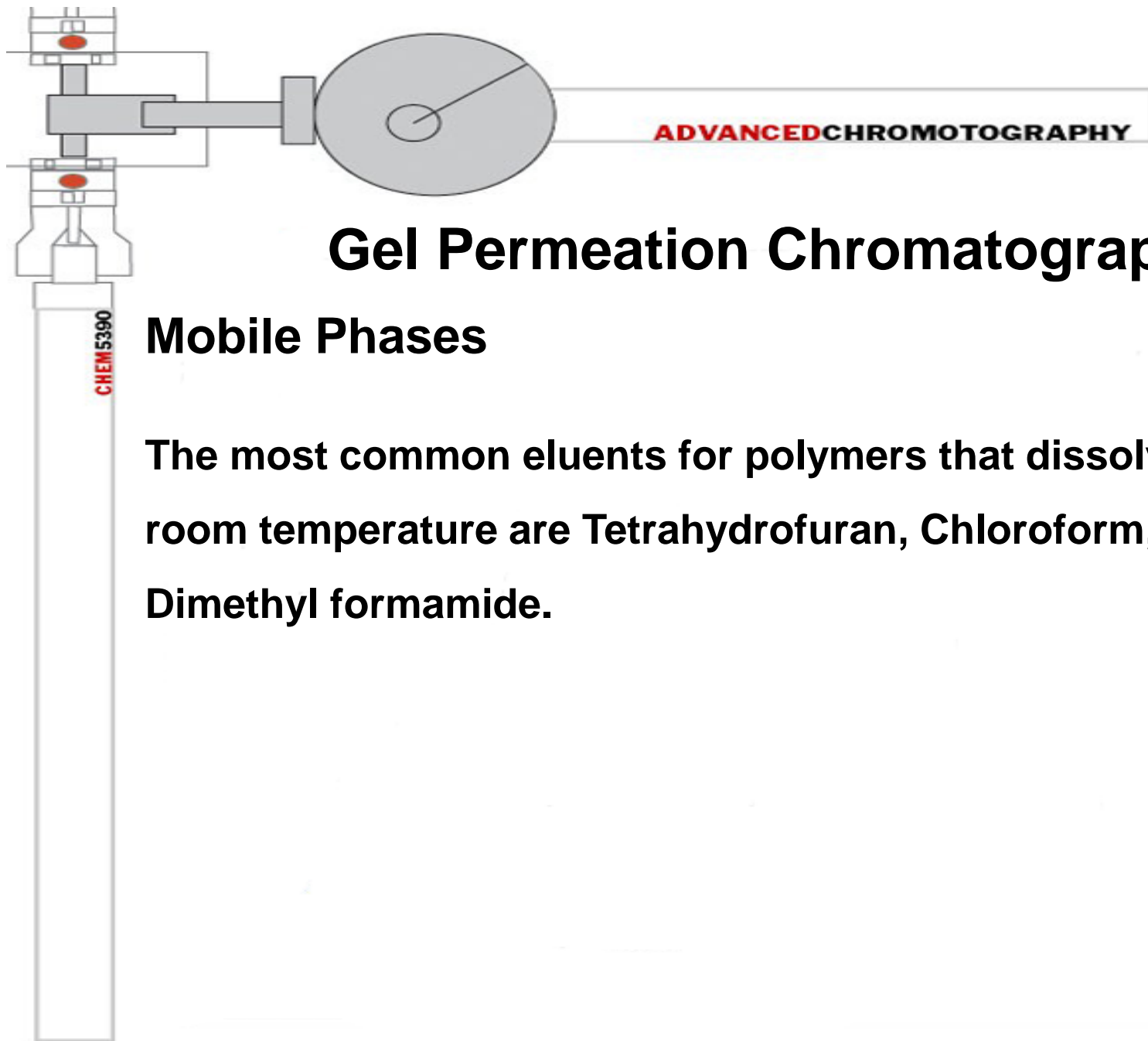
# **Gel Permeation Chromatography**

## **Mobile Phases**

**Polar solvents, i.e., methanol, ethanol, etc... should not be used with polystyrene packings which can permanently damage the column.**

**Solvents that are compatible are toluene, chloroform, tetrahydrofuran, etc...**

**Of course these solvents with silica gel and other aqueous compatible packings will work with these types of columns.**

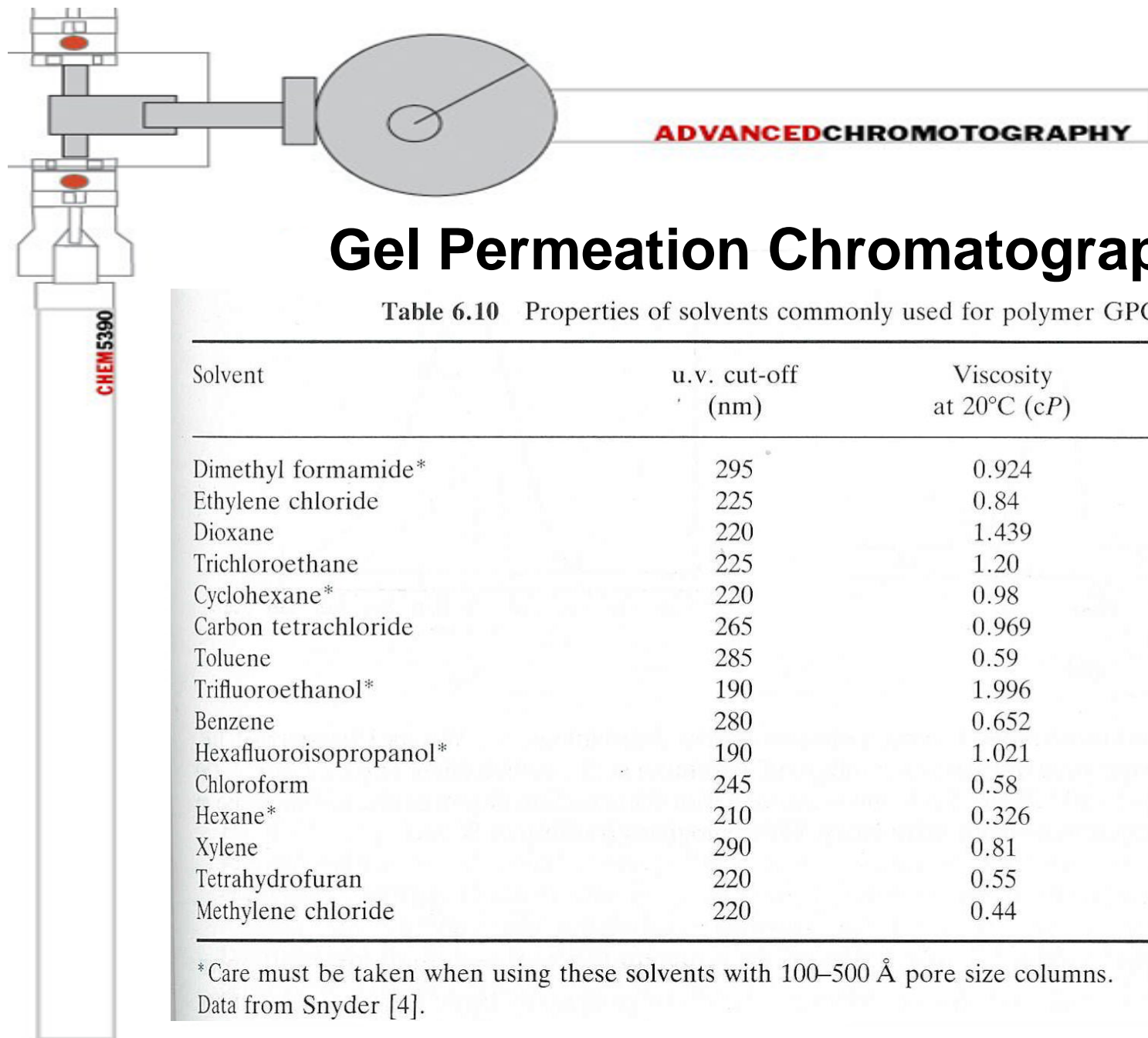


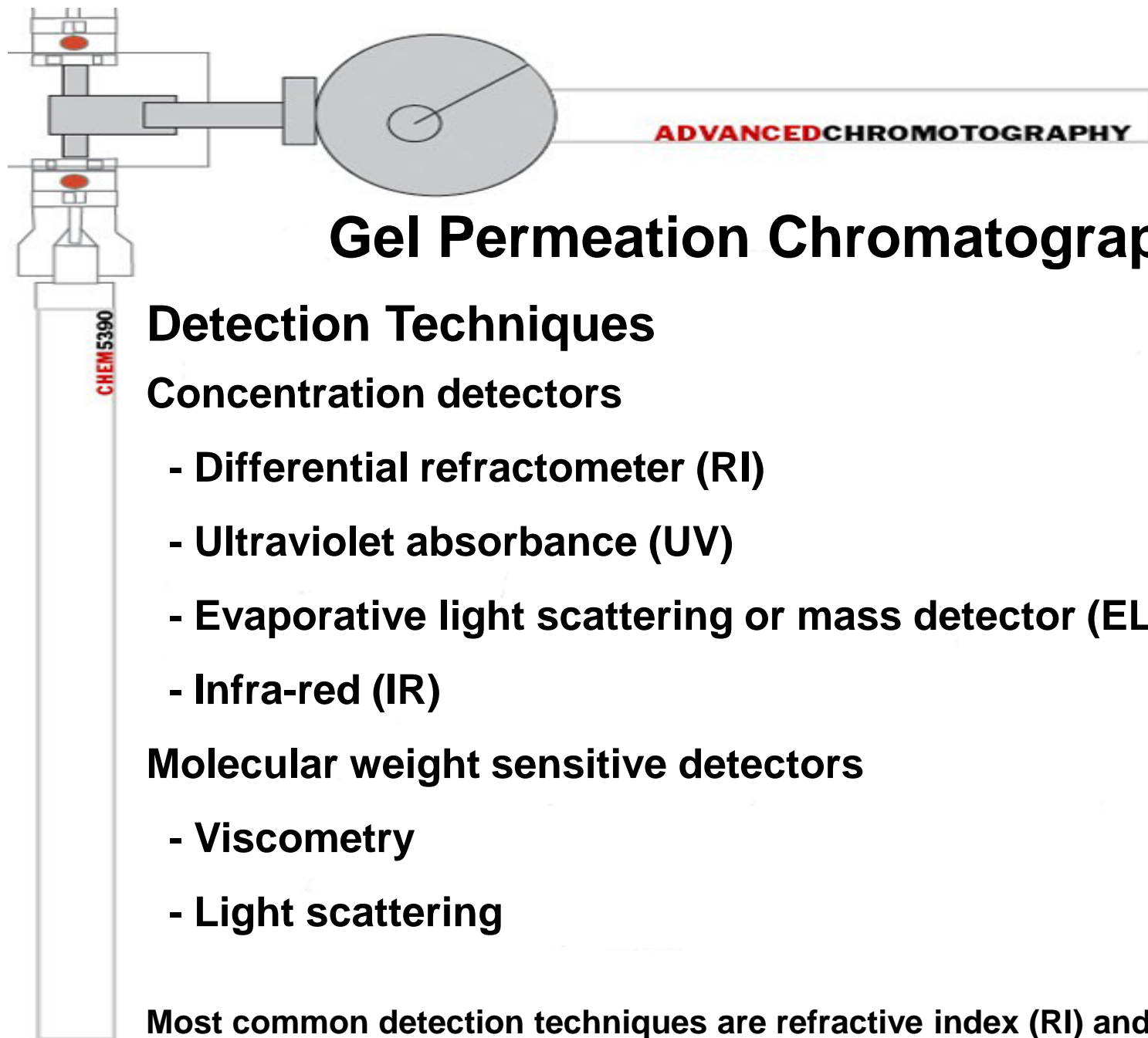
# **Gel Permeation Chromatography**

## **Mobile Phases**

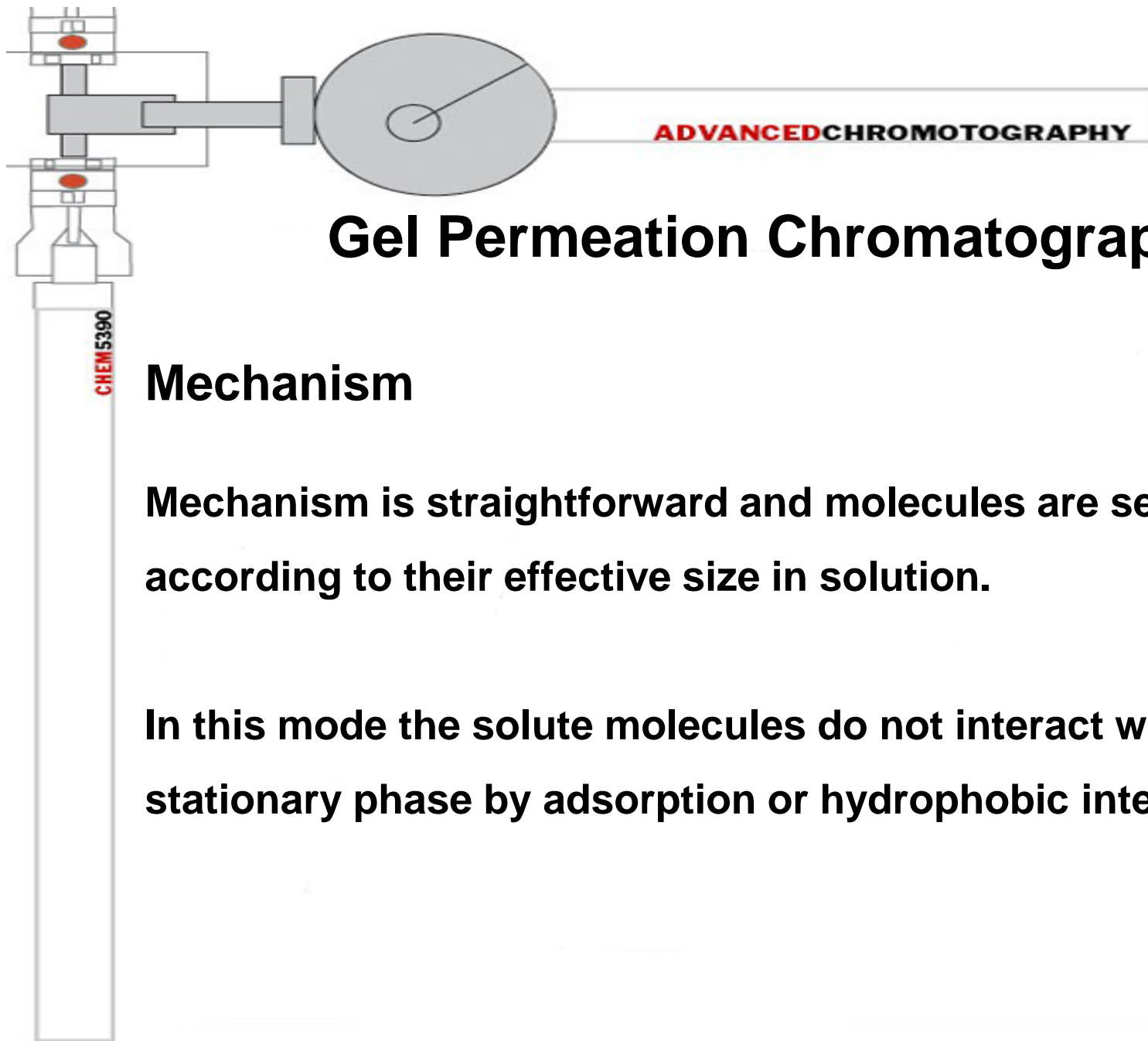
**The most common eluents for polymers that dissolve at room temperature are Tetrahydrofuran, Chloroform, and Dimethyl formamide.**









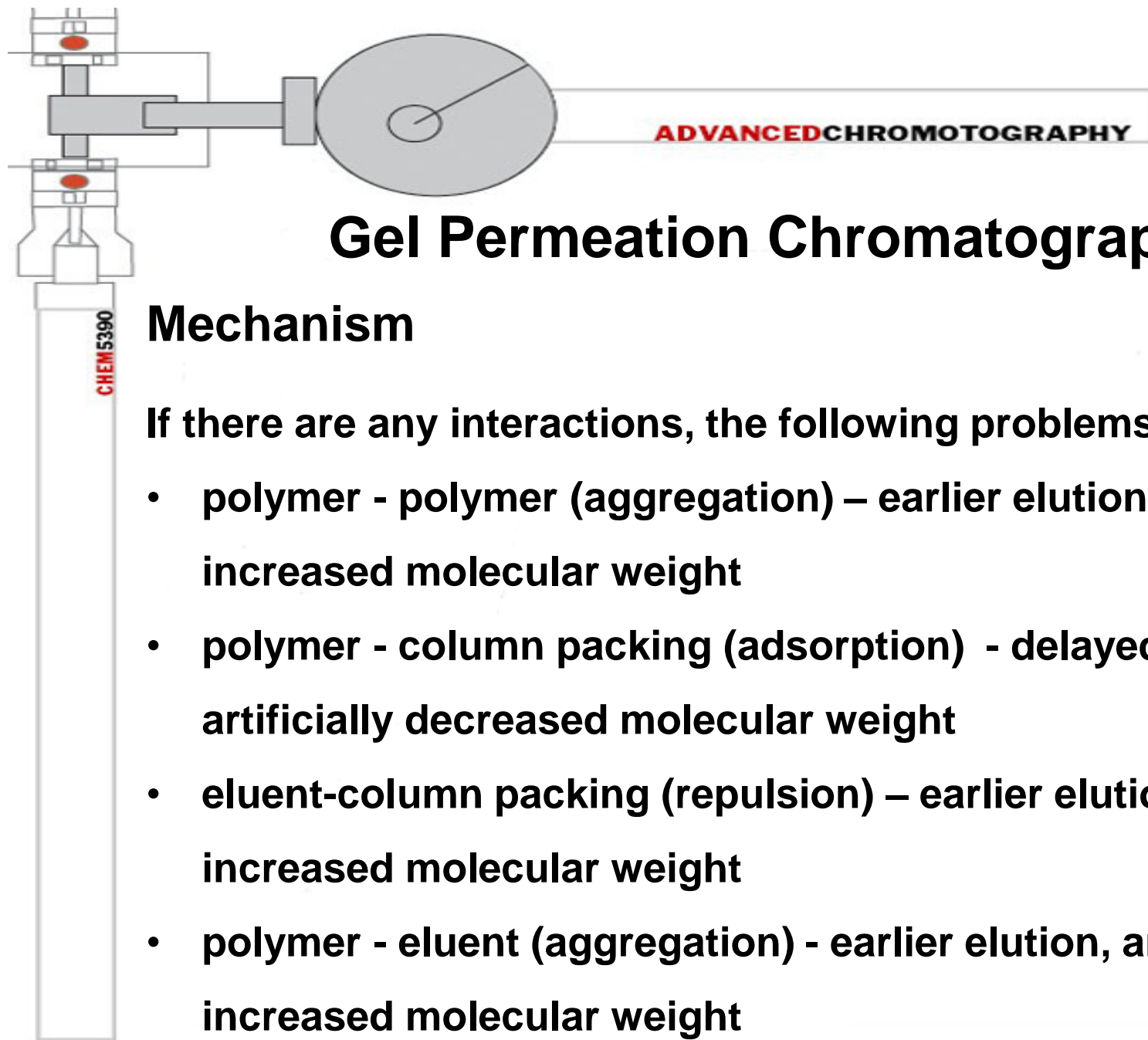


# Gel Permeation Chromatography

## Mechanism

Mechanism is straightforward and molecules are separated according to their effective size in solution.

In this mode the solute molecules do not interact with the stationary phase by adsorption or hydrophobic interaction.

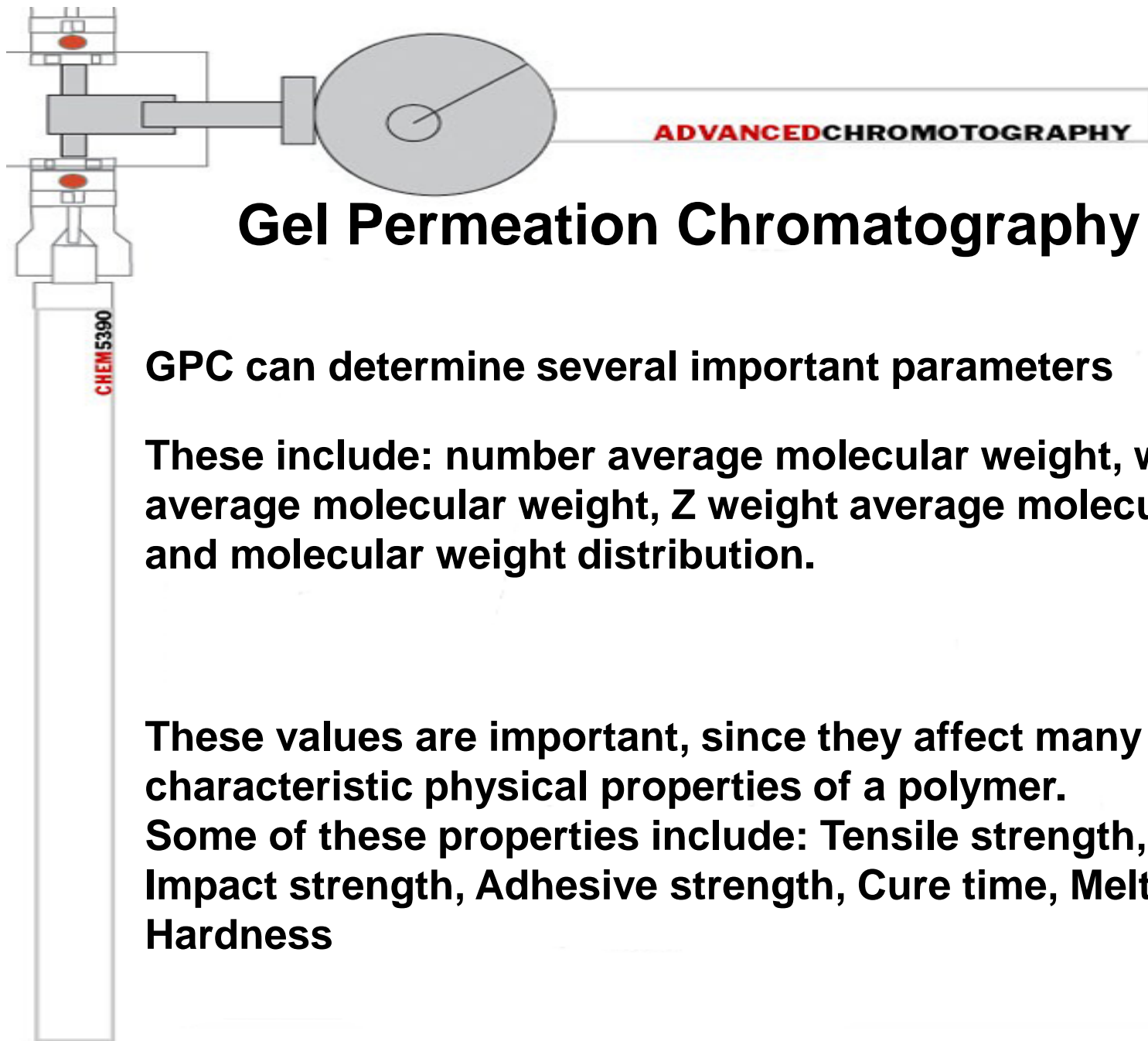


# Gel Permeation Chromatography

## Mechanism

If there are any interactions, the following problems can occur:

- polymer - polymer (aggregation) – earlier elution, artificially increased molecular weight
- polymer - column packing (adsorption) - delayed elution, artificially decreased molecular weight
- eluent-column packing (repulsion) – earlier elution, artificially increased molecular weight
- polymer - eluent (aggregation) - earlier elution, artificially increased molecular weight



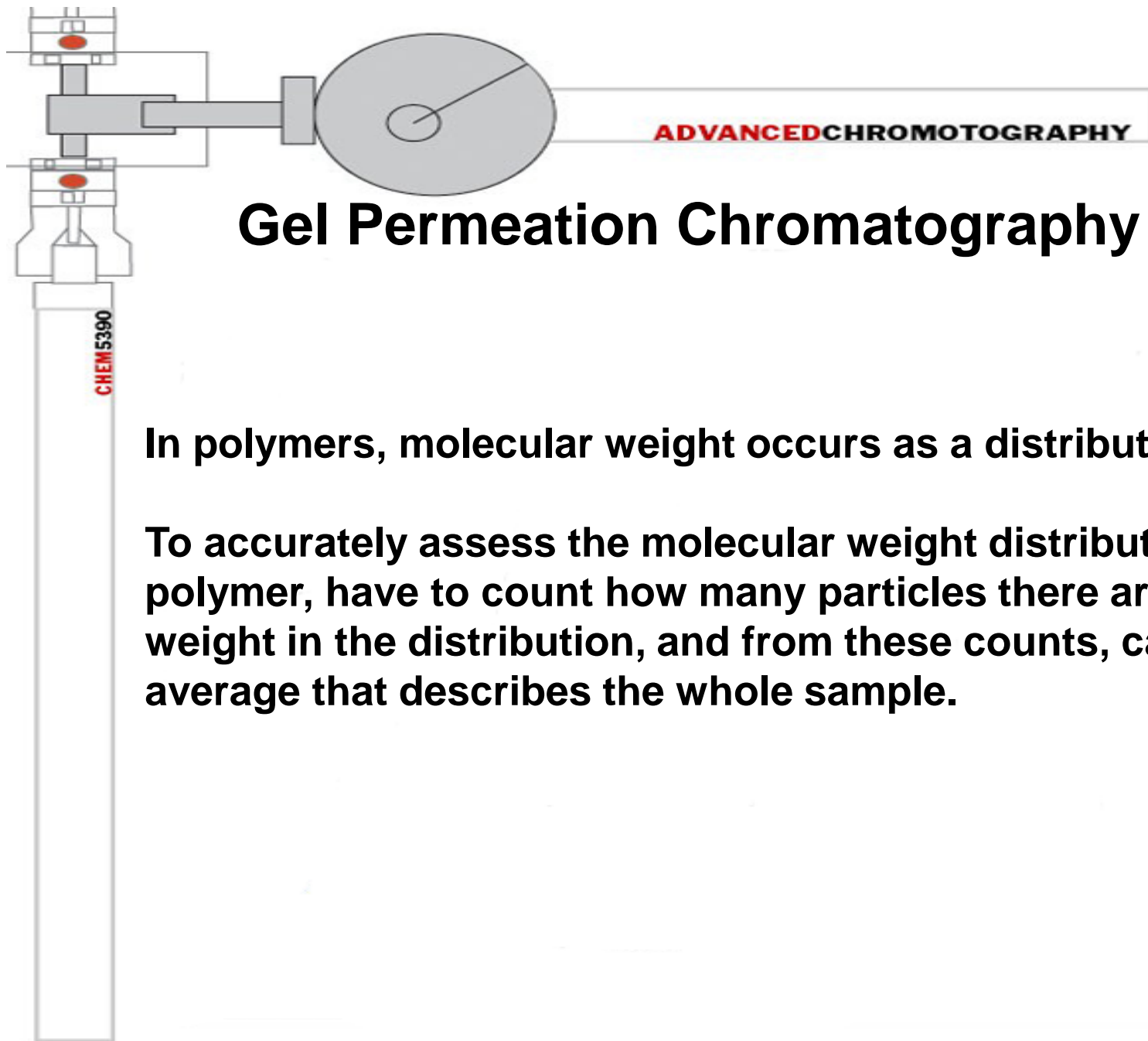
# **Gel Permeation Chromatography (GPC)**

**GPC can determine several important parameters**

**These include: number average molecular weight, weight average molecular weight, Z weight average molecular weight, and molecular weight distribution.**

**These values are important, since they affect many of the characteristic physical properties of a polymer.**

**Some of these properties include: Tensile strength, Brittleness, Impact strength, Adhesive strength, Cure time, Melt viscosity, Hardness**



# Gel Permeation Chromatography (GPC)

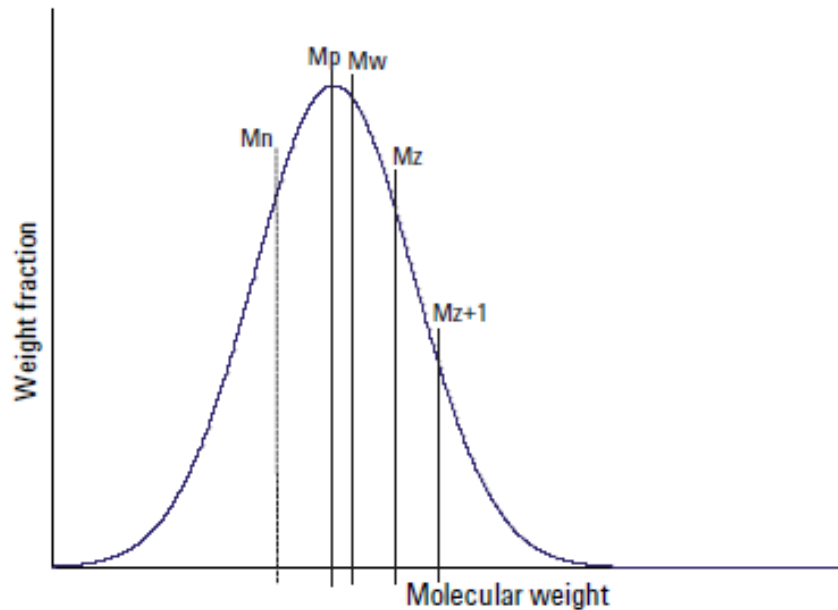
**In polymers, molecular weight occurs as a distribution.**

**To accurately assess the molecular weight distribution of a polymer, have to count how many particles there are at every weight in the distribution, and from these counts, calculate an average that describes the whole sample.**

# Gel Permeation Chromatography (GPC)

In polymers, molecular weight occurs as a distribution.

The typical position of molecular weight average.

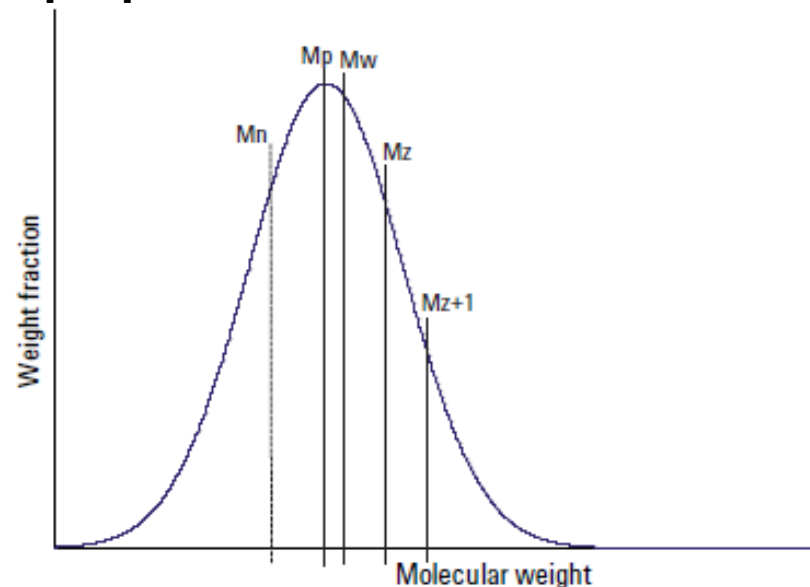


**Figure 3.** The average molecular weights of a mono-modal polymer – in this case the distribution is nearly symmetrical

# Gel Permeation Chromatography (GPC)

The commonly calculated average is called the number average molecular weight, abbreviated to  $M_n$ .

Looking at the distribution in the figure – the  $M_n$  value marks the value at which there are equal numbers of molecules on each side, at higher and lower molecular weight. The value of  $M_n$  influences the thermodynamic properties of the molecule.

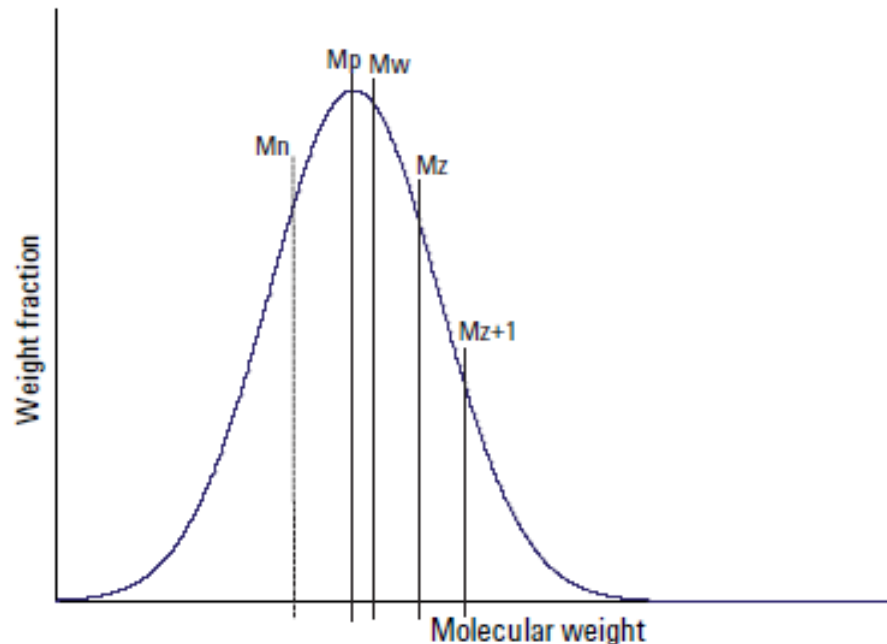


*Figure 3. The average molecular weights of a mono-modal polymer – in this case the distribution is nearly symmetrical*

# Gel Permeation Chromatography (GPC)

Other ways of describing molecular weight average include weight average molecular weight ( $M_w$ ).

$M_w$  is defined as the value at which there are equal masses of molecules on each side, at higher and lower molecular weight.

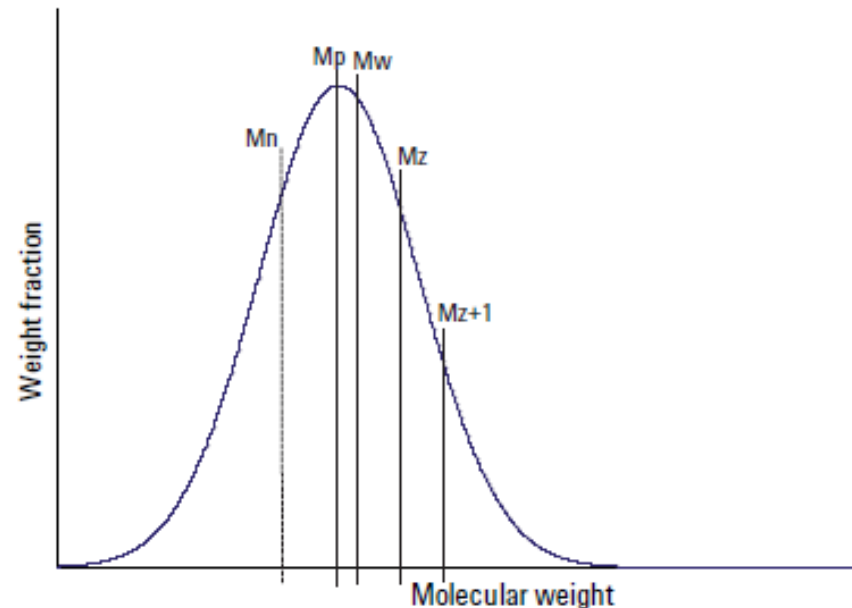


*Figure 3. The average molecular weights of a mono-modal polymer – in this case the distribution is nearly symmetrical*

# Gel Permeation Chromatography (GPC)

The  $M_w$  value is always greater than the  $M_n$  value unless the polymer is completely monodisperse.

$M_w$  is large-molecule sensitive and influences the bulk properties and toughness of the polymer, and is the most often quoted molecular weight average.



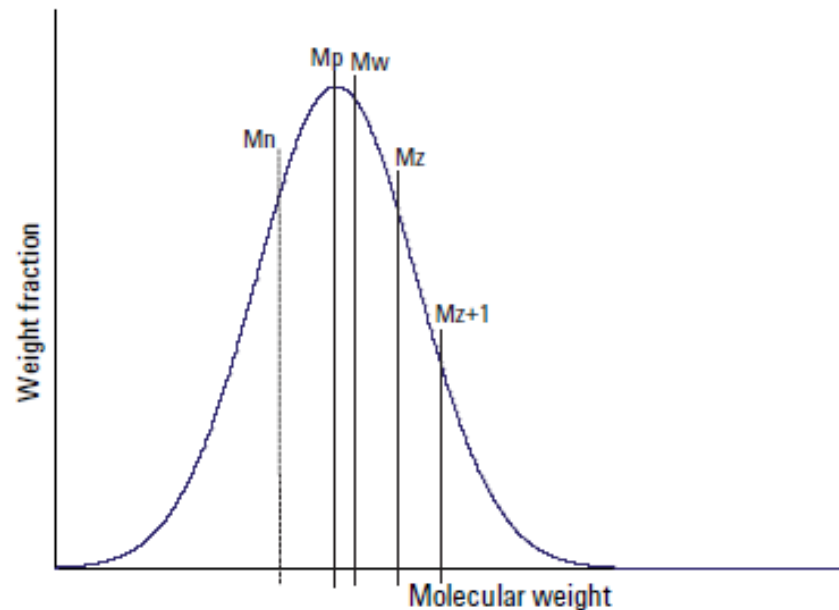
*Figure 3. The average molecular weights of a mono-modal polymer – in this case the distribution is nearly symmetrical*



# Gel Permeation Chromatography (GPC)

The ratio of  $M_w$  to  $M_n$  is used to calculate the polydispersity index (PDI) of a polymer, which provides an indication of the material's range of molecular mass.

The broader the molecular weight distribution, the larger the PDI.

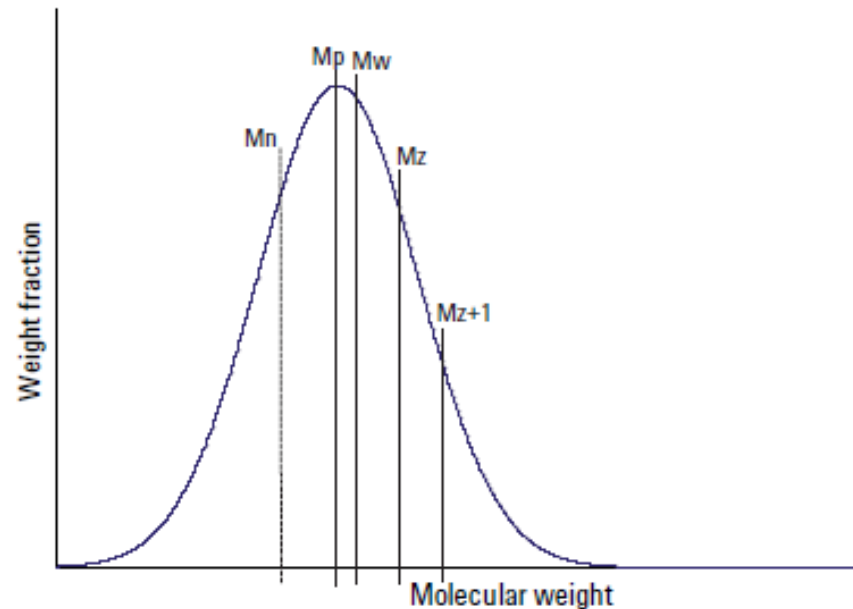


**Figure 3.** The average molecular weights of a mono-modal polymer – in this case the distribution is nearly symmetrical

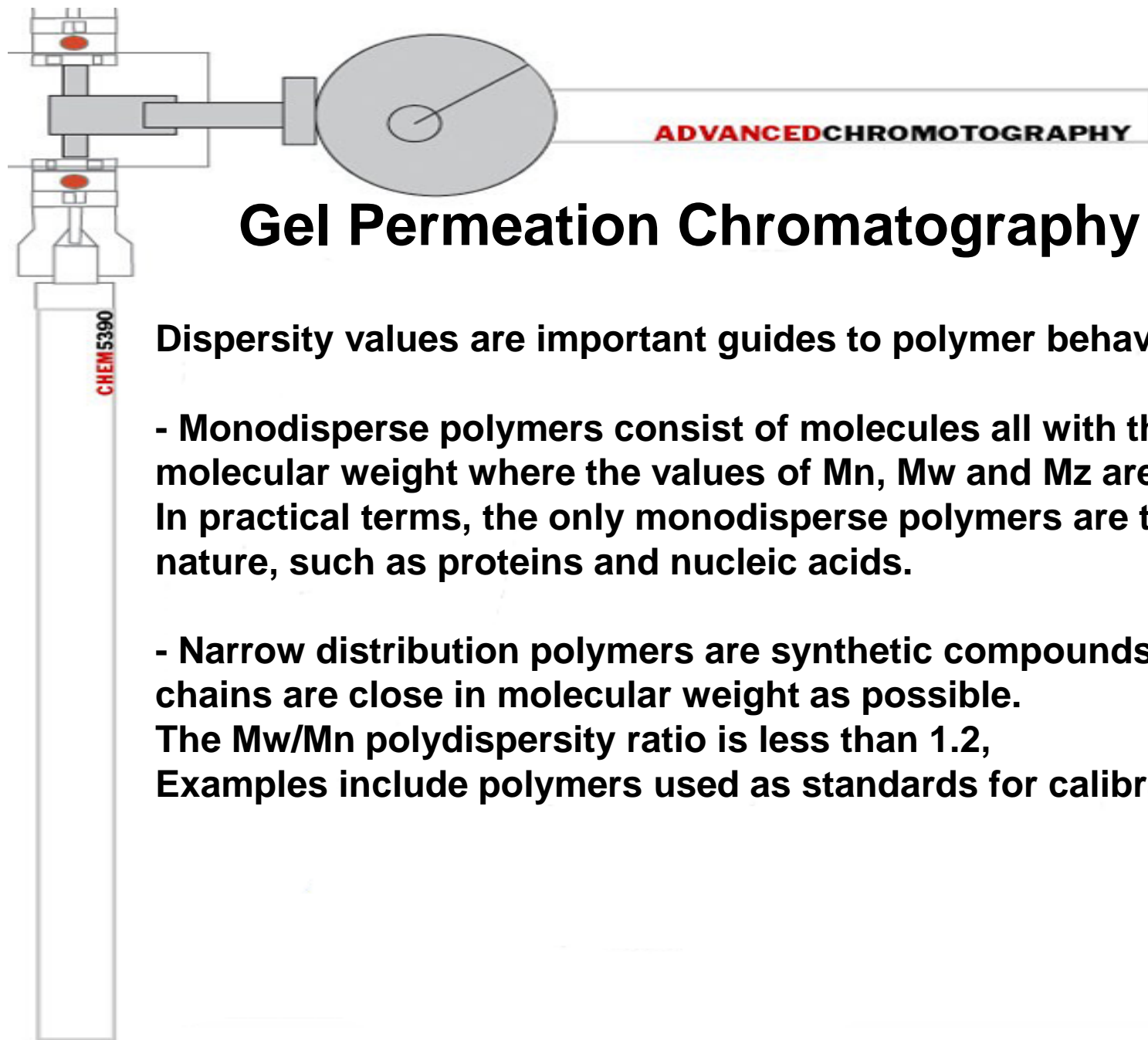
# Gel Permeation Chromatography (GPC)

Other molecular weight averages take into account the higher molecular weight components of the sample, such as z-average molecular weight ( $M_z$ ) and  $M_{z+1}$ .

$M_z$  is sensitive to even larger molecules and influences viscoelasticity and melt flow behavior.



*Figure 3. The average molecular weights of a mono-modal polymer – in this case the distribution is nearly symmetrical*



# Gel Permeation Chromatography (GPC)

Dispersity values are important guides to polymer behavior:

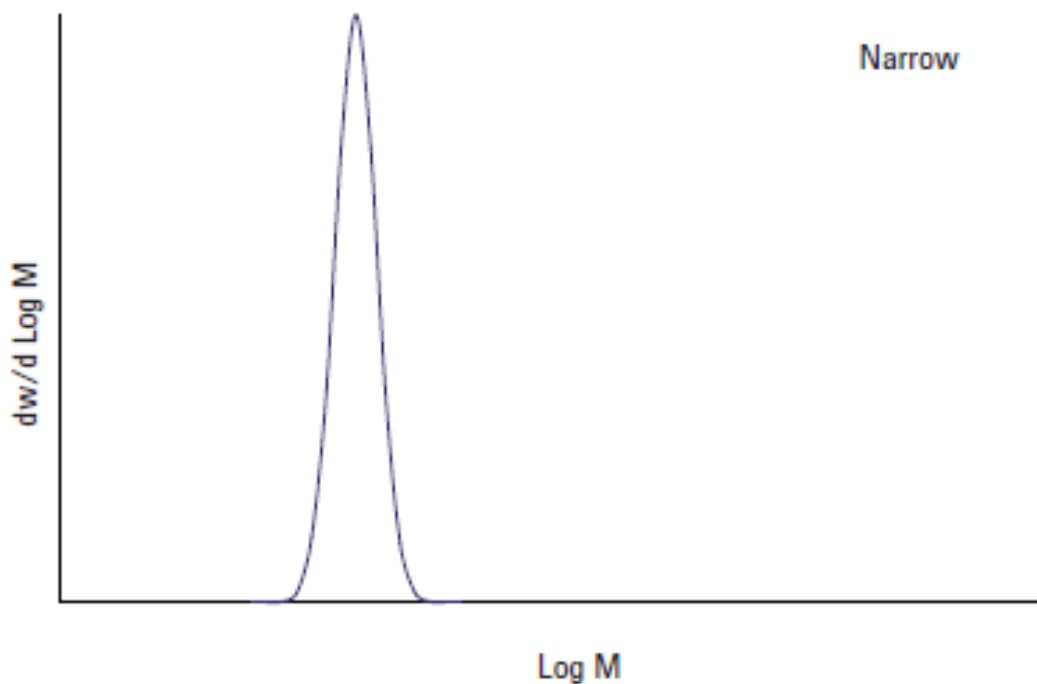
- Monodisperse polymers consist of molecules all with the same molecular weight where the values of  $M_n$ ,  $M_w$  and  $M_z$  are identical. In practical terms, the only monodisperse polymers are those found in nature, such as proteins and nucleic acids.
- Narrow distribution polymers are synthetic compounds where all the chains are close in molecular weight as possible. The  $M_w/M_n$  polydispersity ratio is less than 1.2, Examples include polymers used as standards for calibrations.

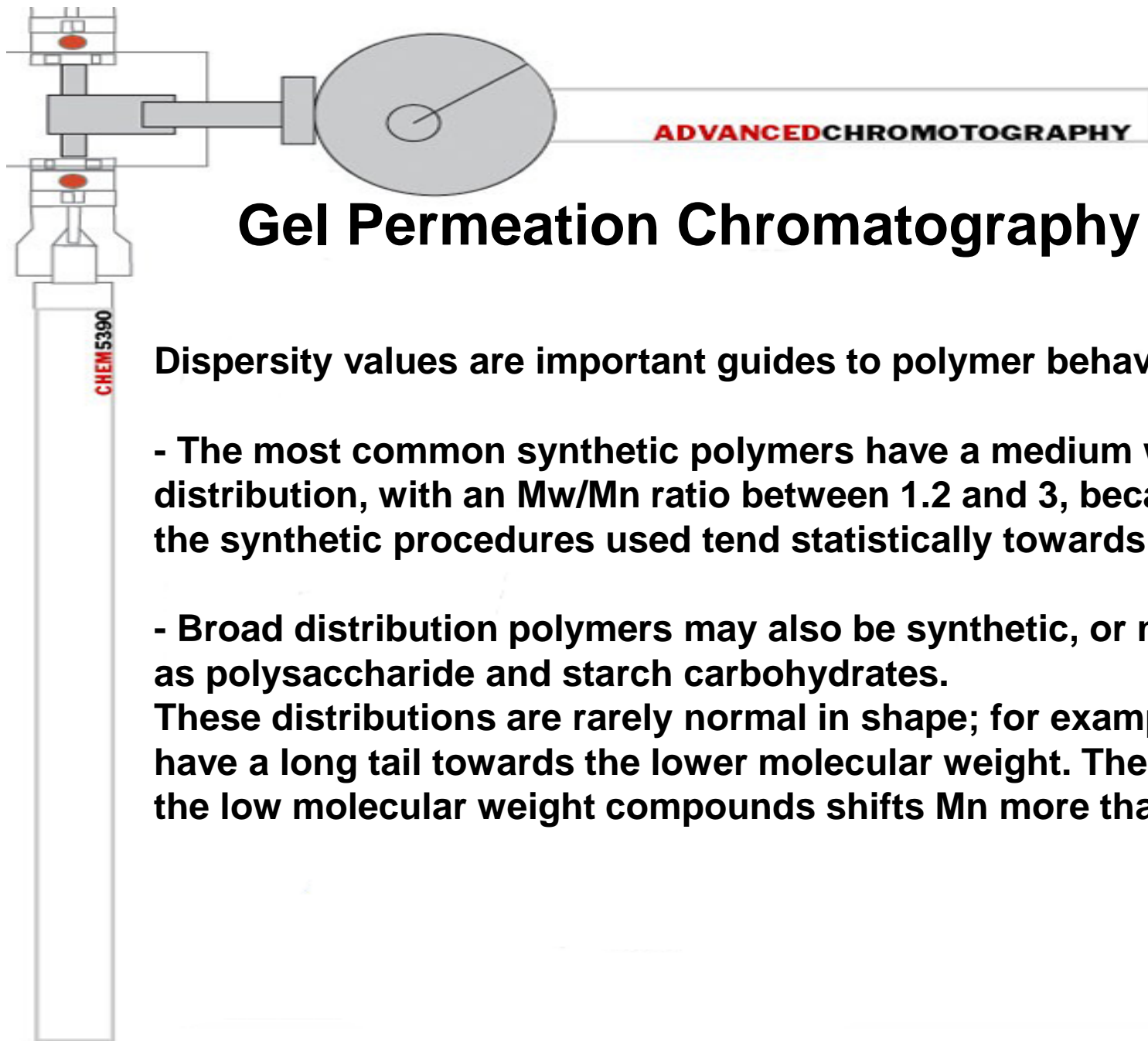


# Gel Permeation Chromatography (GPC)

Dispersity values are important guides to polymer behavior:

- Narrow distribution polymers





# Gel Permeation Chromatography (GPC)

Dispersity values are important guides to polymer behavior:

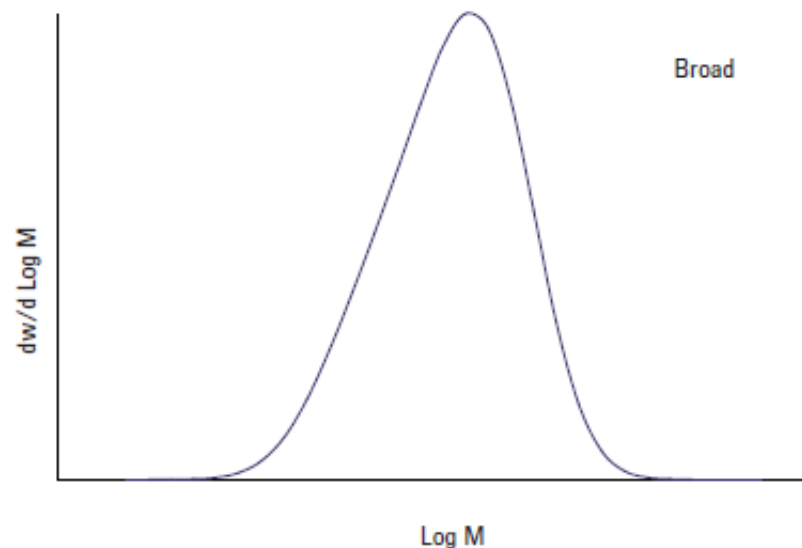
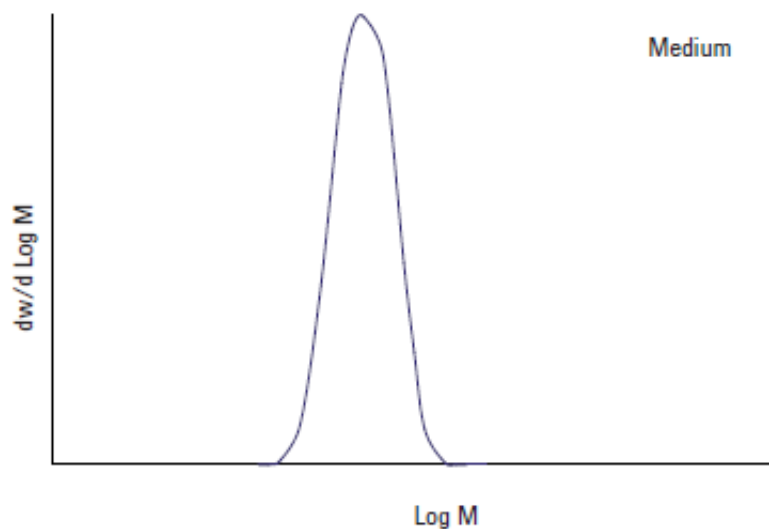
- The most common synthetic polymers have a medium width distribution, with an  $M_w/M_n$  ratio between 1.2 and 3, because many of the synthetic procedures used tend statistically towards these values.
  - Broad distribution polymers may also be synthetic, or natural such as polysaccharide and starch carbohydrates.
- These distributions are rarely normal in shape; for example, they may have a long tail towards the lower molecular weight. The presence of the low molecular weight compounds shifts  $M_n$  more than  $M_w$  and  $M_z$ .



# Gel Permeation Chromatography (GPC)

Dispersity values are important guides to polymer behavior:

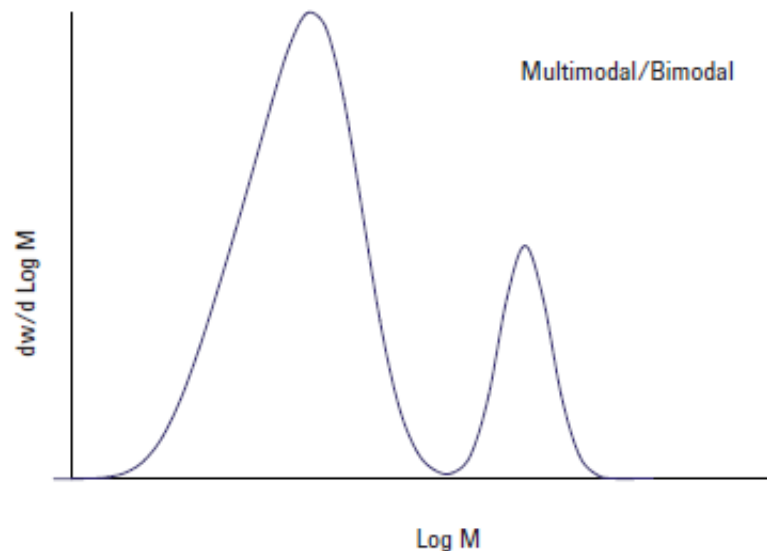
- Medium width distribution polymers
- Broad distribution polymers



# Gel Permeation Chromatography (GPC)

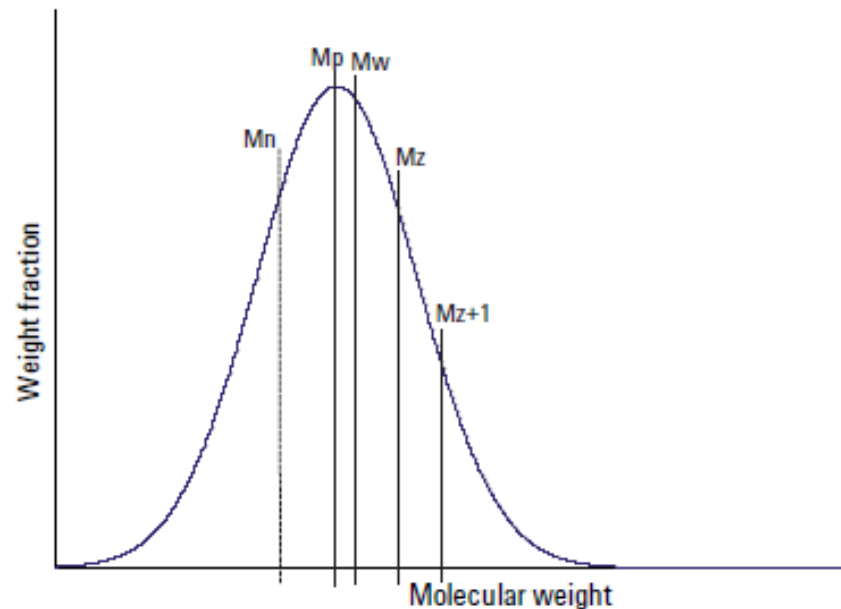
Dispersity values are important guides to polymer behavior:

Sometimes synthetic polymers have multimodal distributions. If two or more polymers of differing molecular weight are present, then the two peaks can overlap, resulting in a bimodal distribution



# Gel Permeation Chromatography (GPC)

For standards - need to be of very high quality and with extremely narrow molecular weight distributions so that the position of the top of the peak, the peak molecular weight ( $M_p$ ) can be assigned with confidence. It is the  $M_p$  value used to set up the calibration.



*Figure 3. The average molecular weights of a mono-modal polymer – in this case the distribution is nearly symmetrical*





# Gel Permeation Chromatography

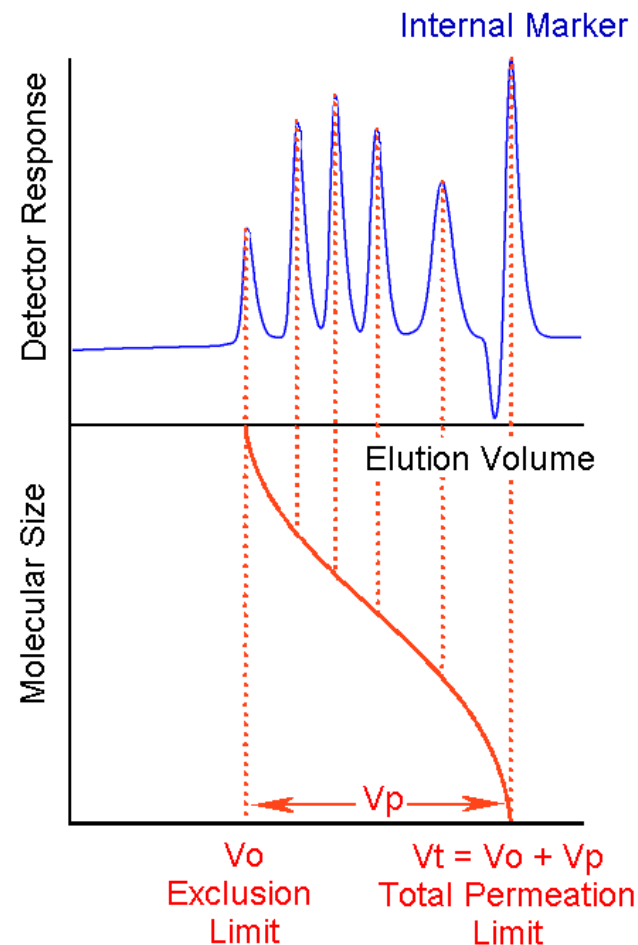
## Elution Profile

**Exclusion volume ( $V_o$ ) –**  
Upper MW limit  
(also known as void volume)

**Total permeation volume ( $V_t$ ) –**  
Lower MW limit

**Pore volume ( $V_p$ ) –**  
Working resolving range of MW

$$V_p = V_t - V_o$$



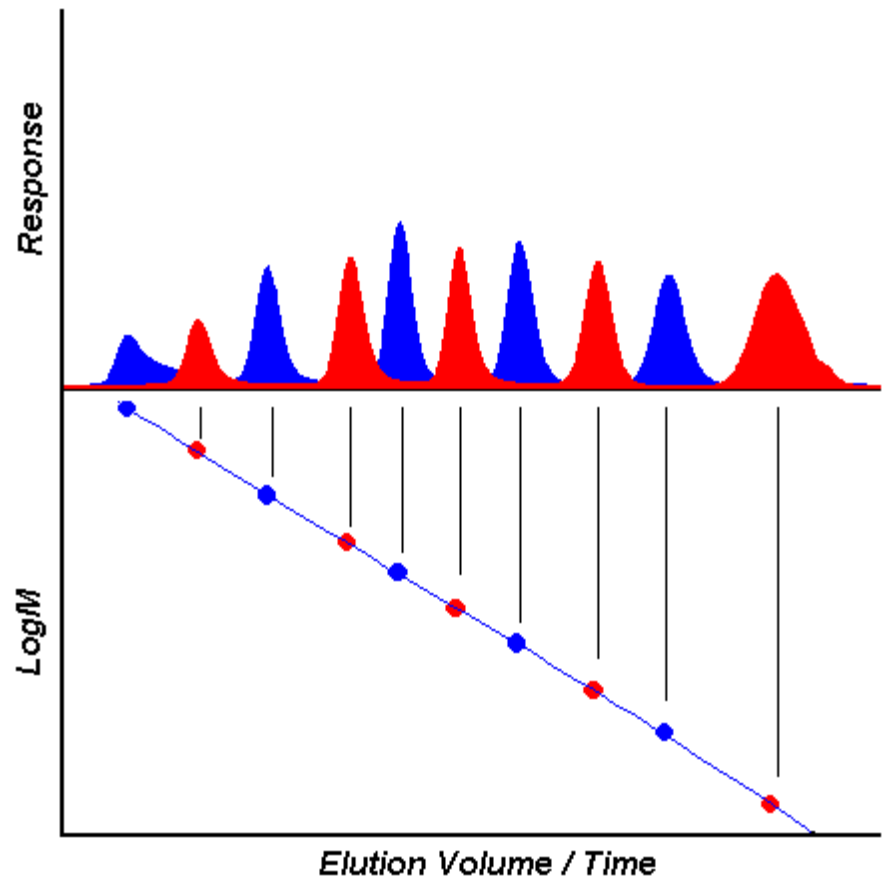


ADVANCED CHROMATOGRAPHY

# Gel Permeation Chromatography

## Calibration

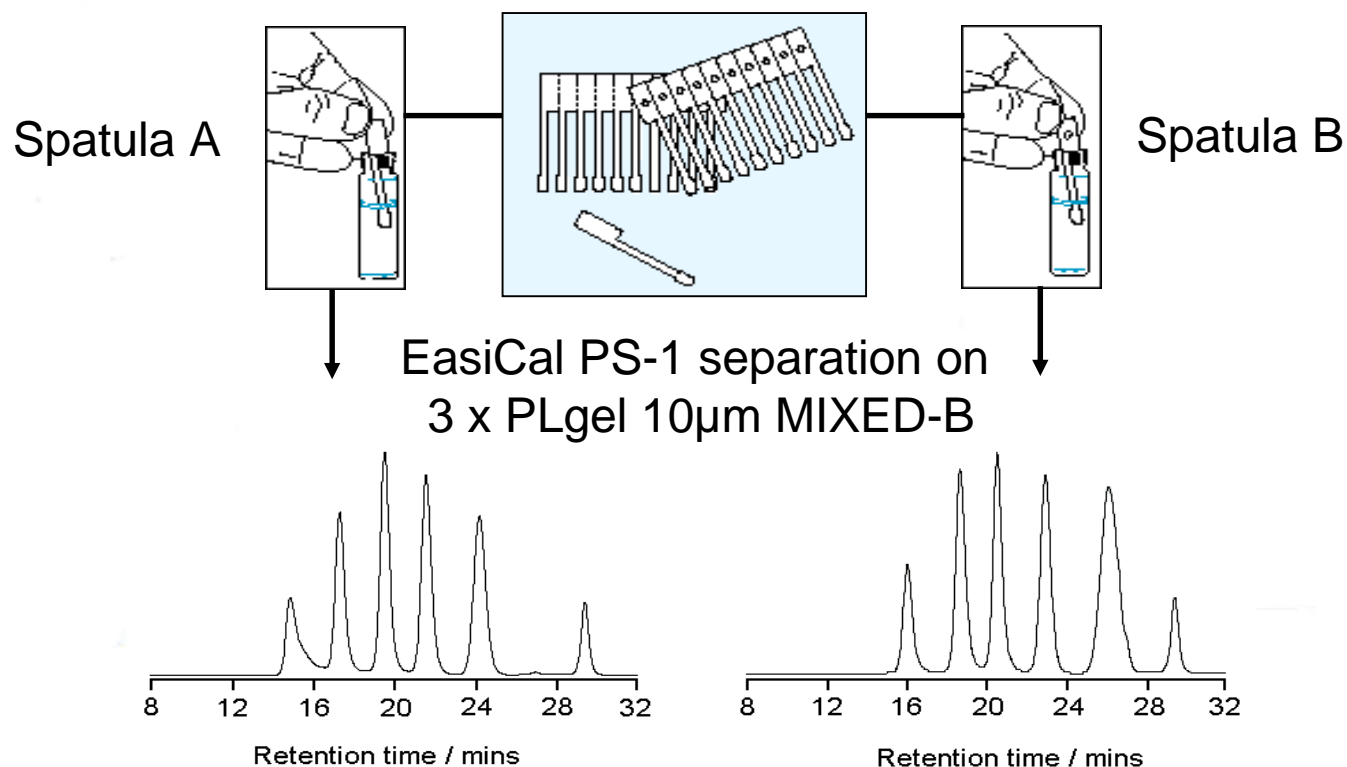
Injections of multiple narrow standards reduces the time taken to calibrate the system.





# Gel Permeation Chromatography

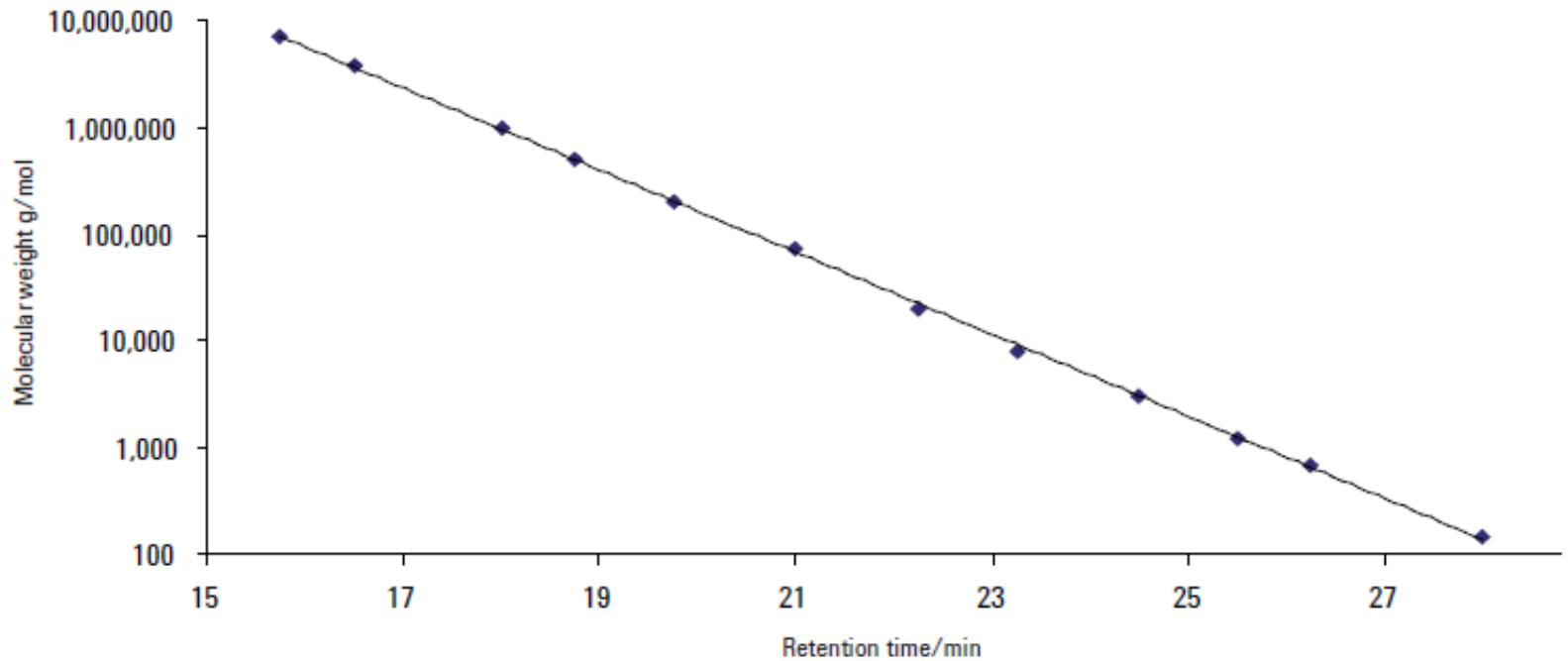
## Calibration - EasiCal Pre-prepared stds (example: Varian)



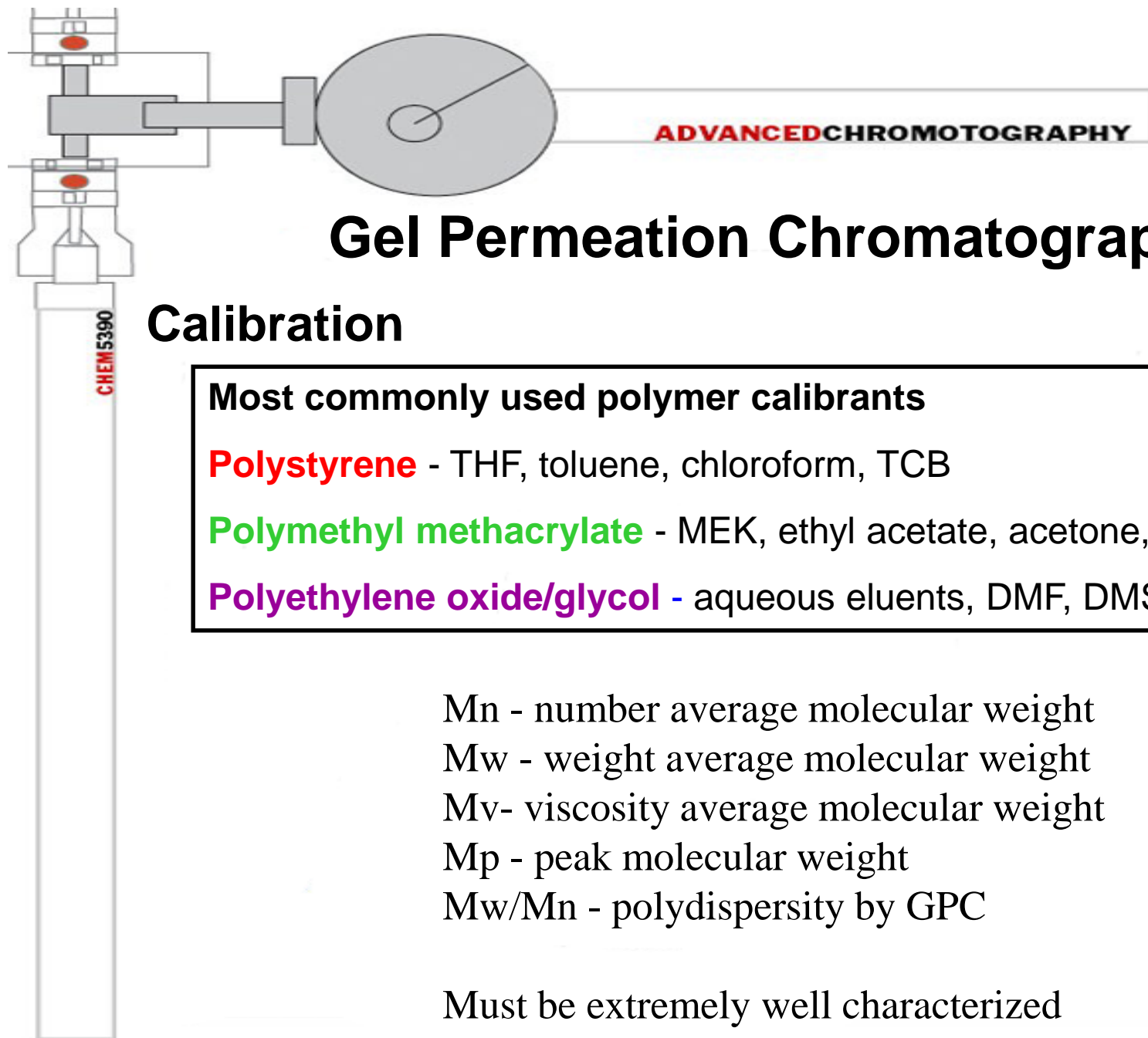
ADVANCED CHROMATOGRAPHY

# Gel Permeation Chromatography

## Calibration - EasiCal Pre-pared example



*Figure 2. A calibration graph used to determine the molecular weight of a polymer from its retention time*



## Calibration

**Most commonly used polymer calibrants**

**Polystyrene** - THF, toluene, chloroform, TCB

**Polymethyl methacrylate** - MEK, ethyl acetate, acetone, DMF

**Polyethylene oxide/glycol** - aqueous eluents, DMF, DMSO

M<sub>n</sub> - number average molecular weight

M<sub>w</sub> - weight average molecular weight

M<sub>v</sub> - viscosity average molecular weight

M<sub>p</sub> - peak molecular weight

M<sub>w</sub>/M<sub>n</sub> - polydispersity by GPC

Must be extremely well characterized

# Gel Permeation Chromatography

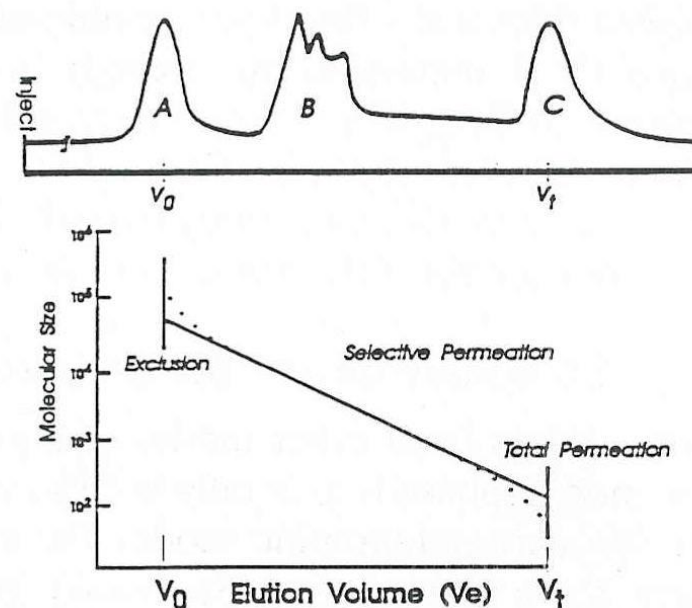
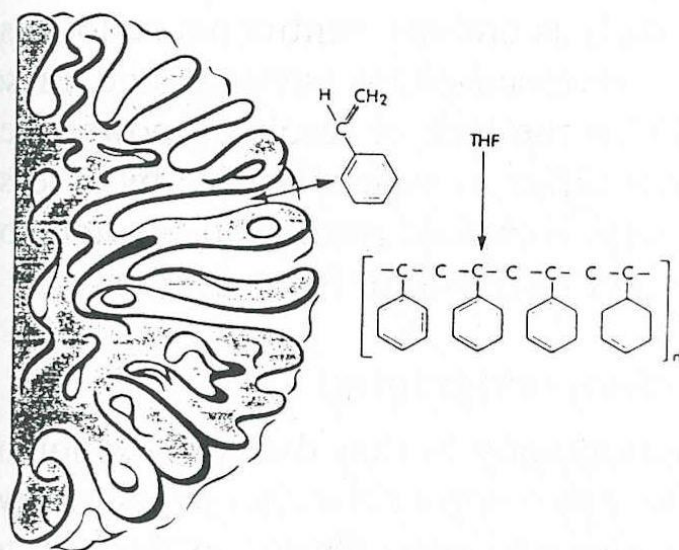
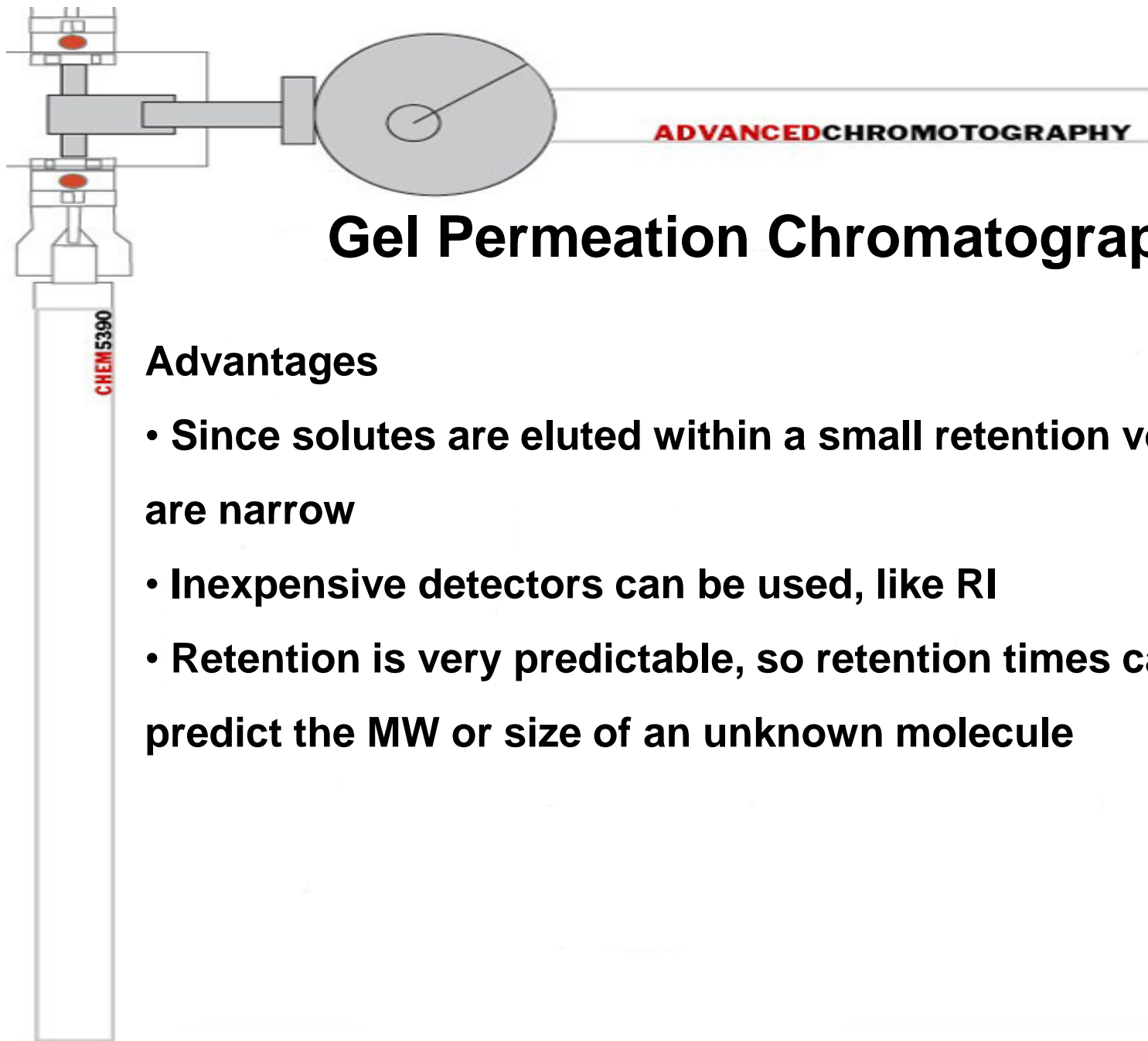


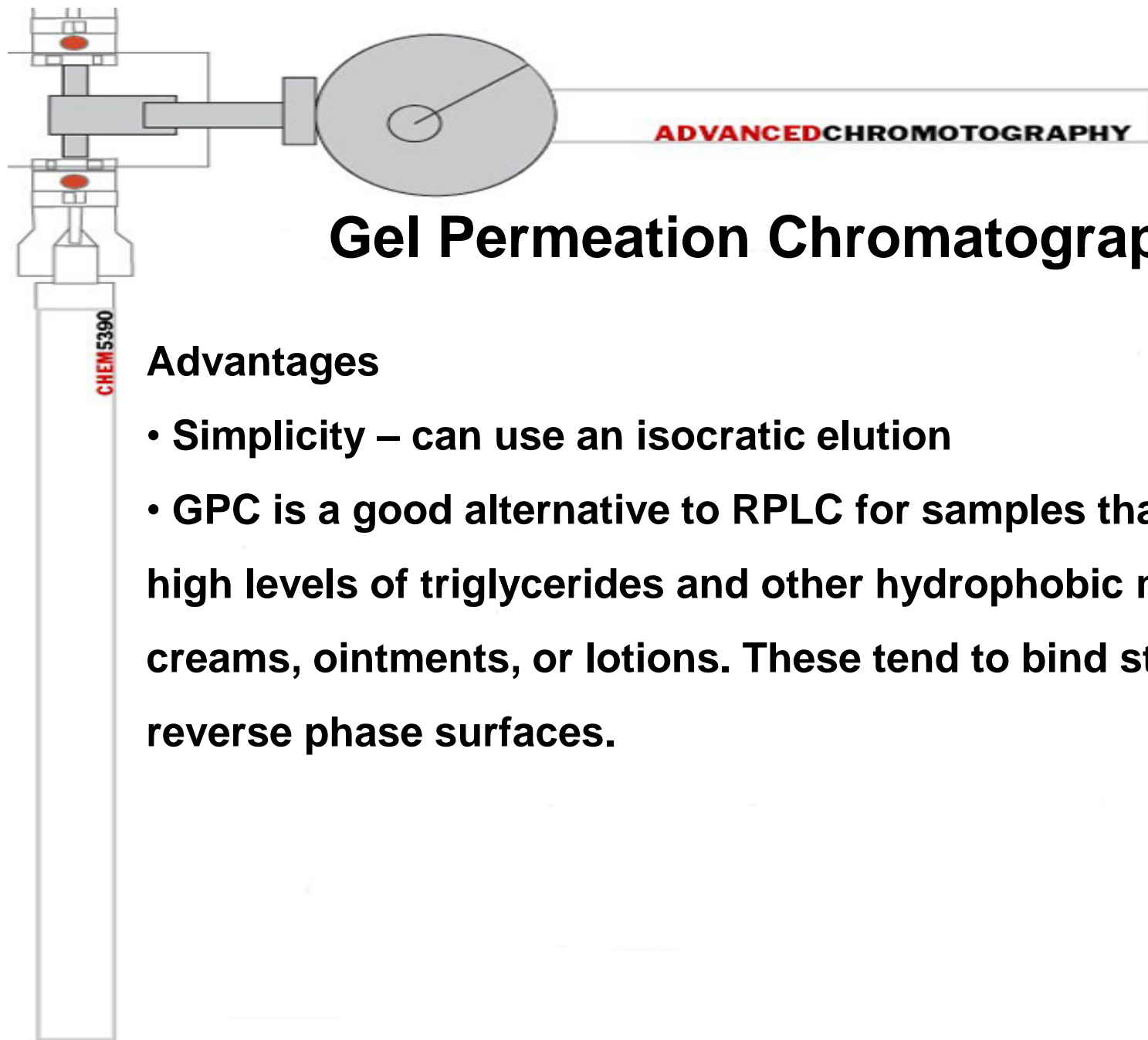
Fig. 6.22. Schematic illustration of the permeation process. In the diagram on the left, the smaller molecule is able to diffuse into solvent within the pores, while the larger solute is excluded. In the diagram on the right,  $V_0$  and  $V_t$  represent the totally excluded and included volumes, respectively. Reprinted from *Developing HPLC Separations, Book One* (1991), Millipore, Milford, with permission.



# Gel Permeation Chromatography

## Advantages

- Since solutes are eluted within a small retention volume, peaks are narrow
- Inexpensive detectors can be used, like RI
- Retention is very predictable, so retention times can be used to predict the MW or size of an unknown molecule

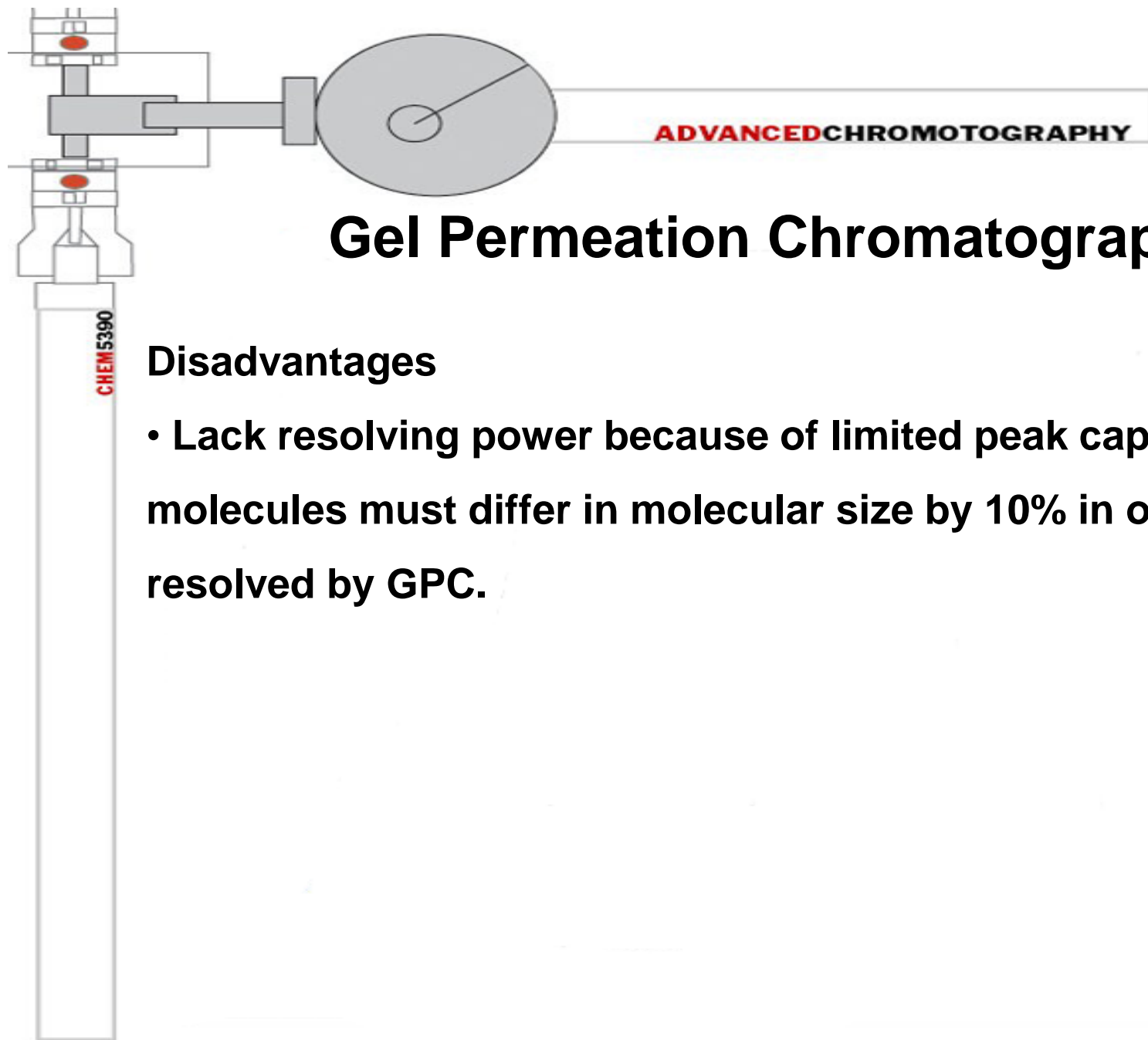


# Gel Permeation Chromatography

## Advantages

- Simplicity – can use an isocratic elution
- GPC is a good alternative to RPLC for samples that contain high levels of triglycerides and other hydrophobic materials, i.e. creams, ointments, or lotions. These tend to bind strongly on reverse phase surfaces.





# Gel Permeation Chromatography

## Disadvantages

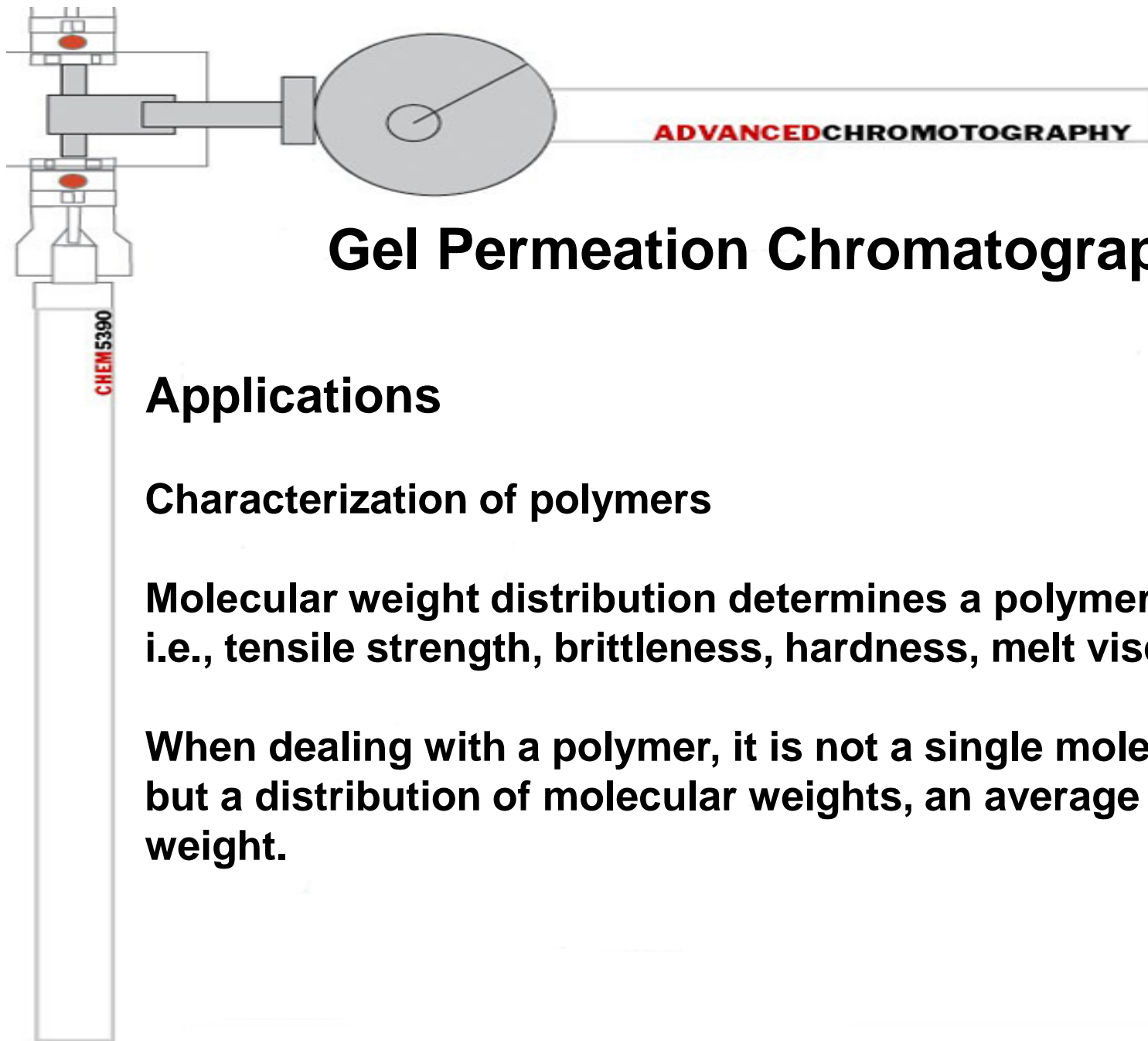
- Lack resolving power because of limited peak capacity, molecules must differ in molecular size by 10% in order to be resolved by GPC.



# Gel Permeation Chromatography

## Applications

- to characterize polymers and mixtures into discrete fractions, such as polymer, oligomer, monomer and any non-polymeric additives.
- to separate biomolecules (i.e. proteins fractionation)
- has been used to purify certain RNA and viruses



# Gel Permeation Chromatography

## Applications

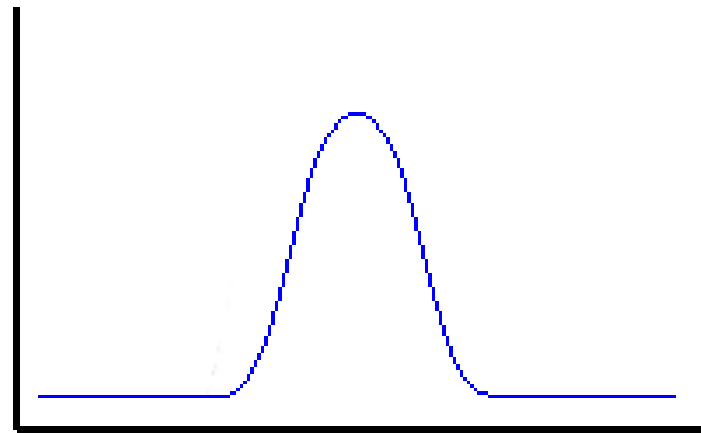
### Characterization of polymers

**Molecular weight distribution determines a polymer's properties, i.e., tensile strength, brittleness, hardness, melt viscosity.**

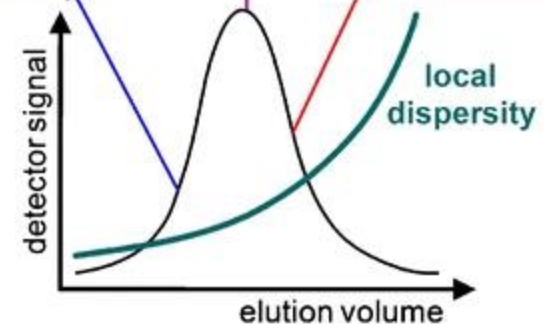
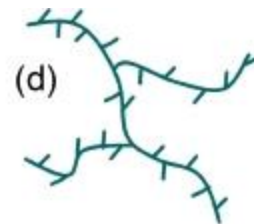
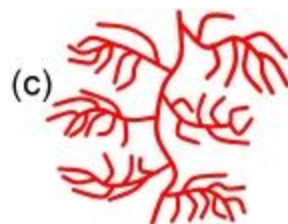
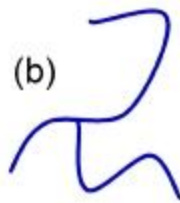
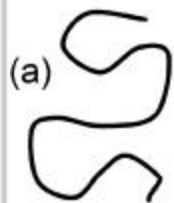
**When dealing with a polymer, it is not a single molecular weight, but a distribution of molecular weights, an average molecular weight.**

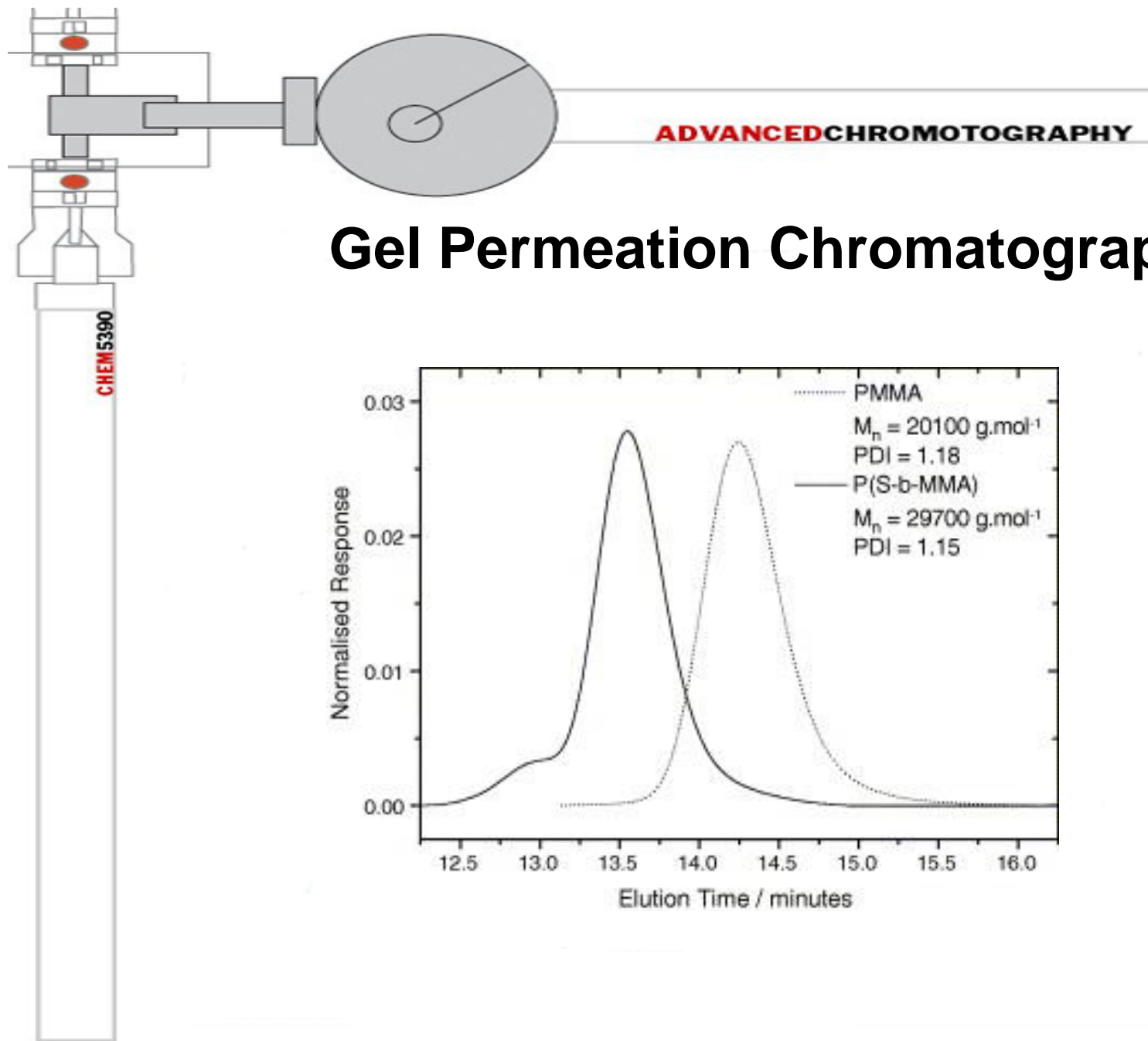
# Gel Permeation Chromatography

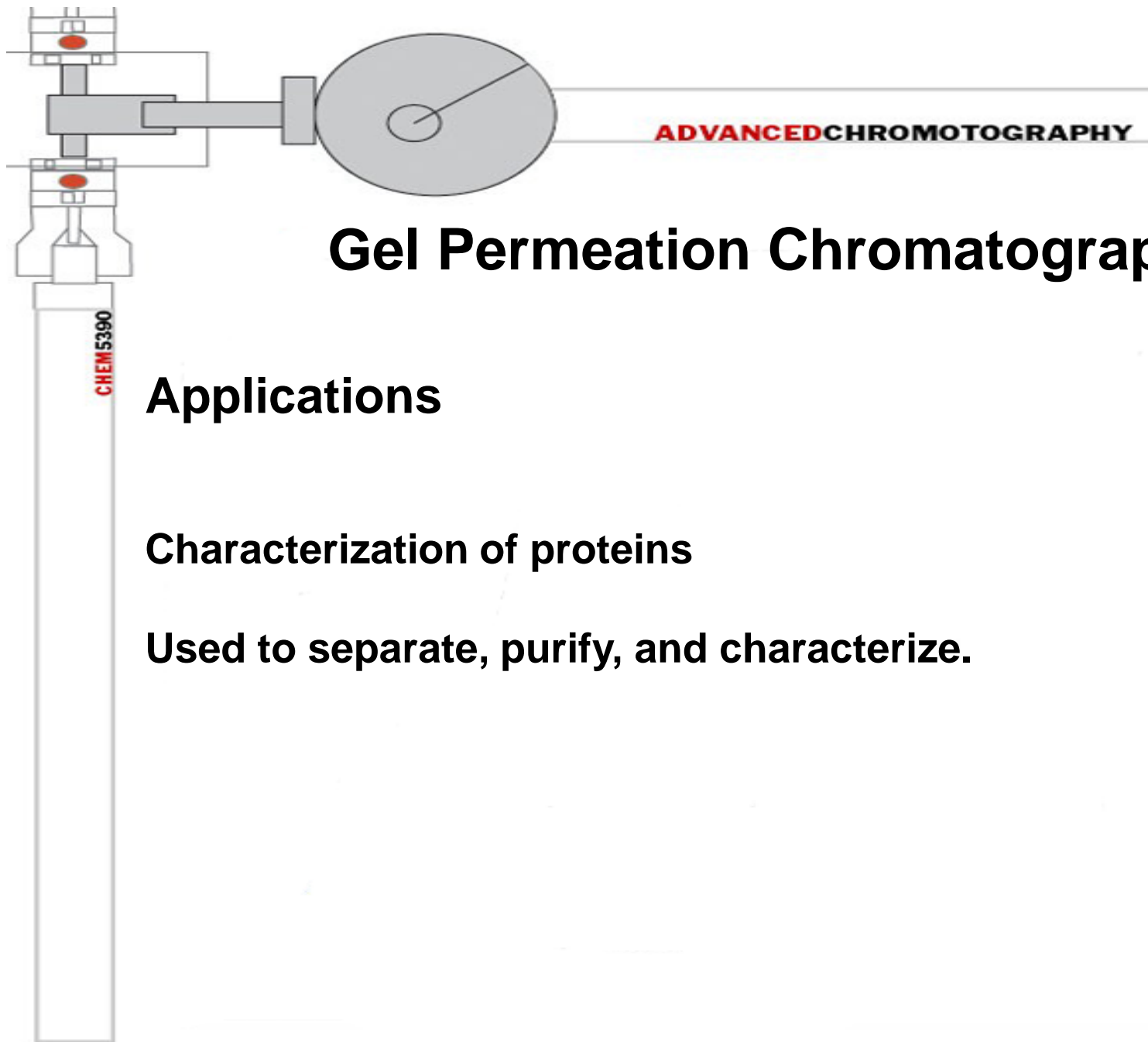
number  
of molecules



molecular weight







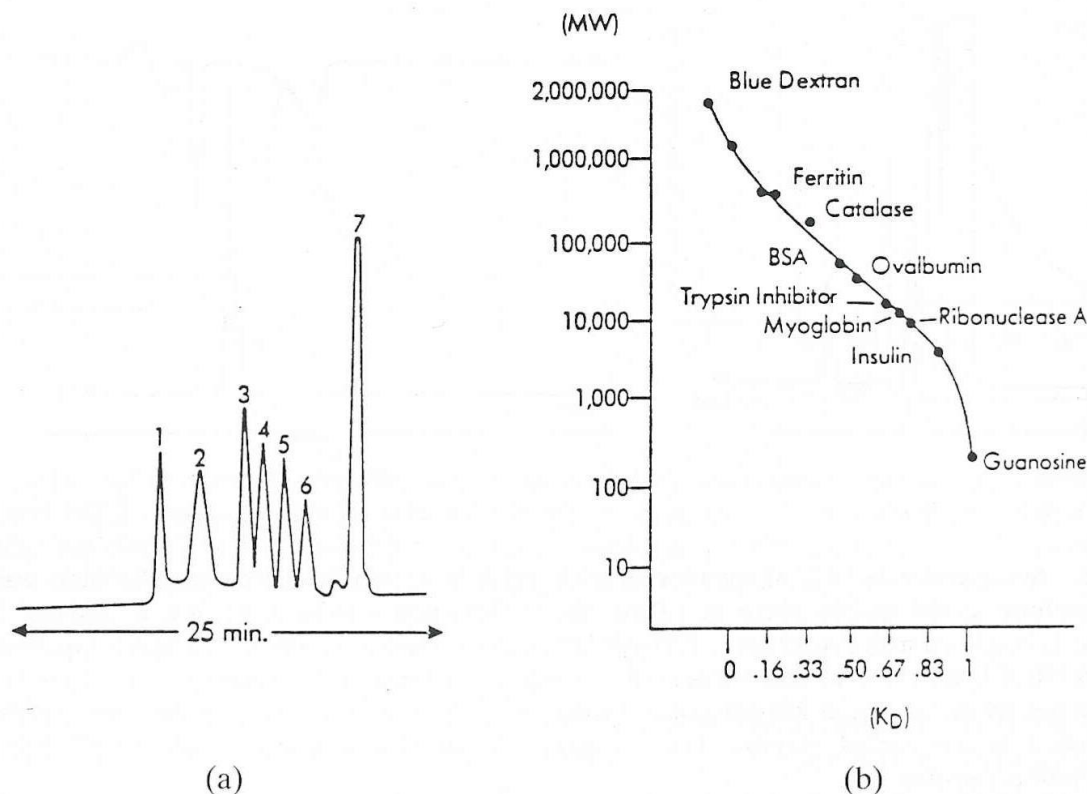
# Gel Permeation Chromatography

## Applications

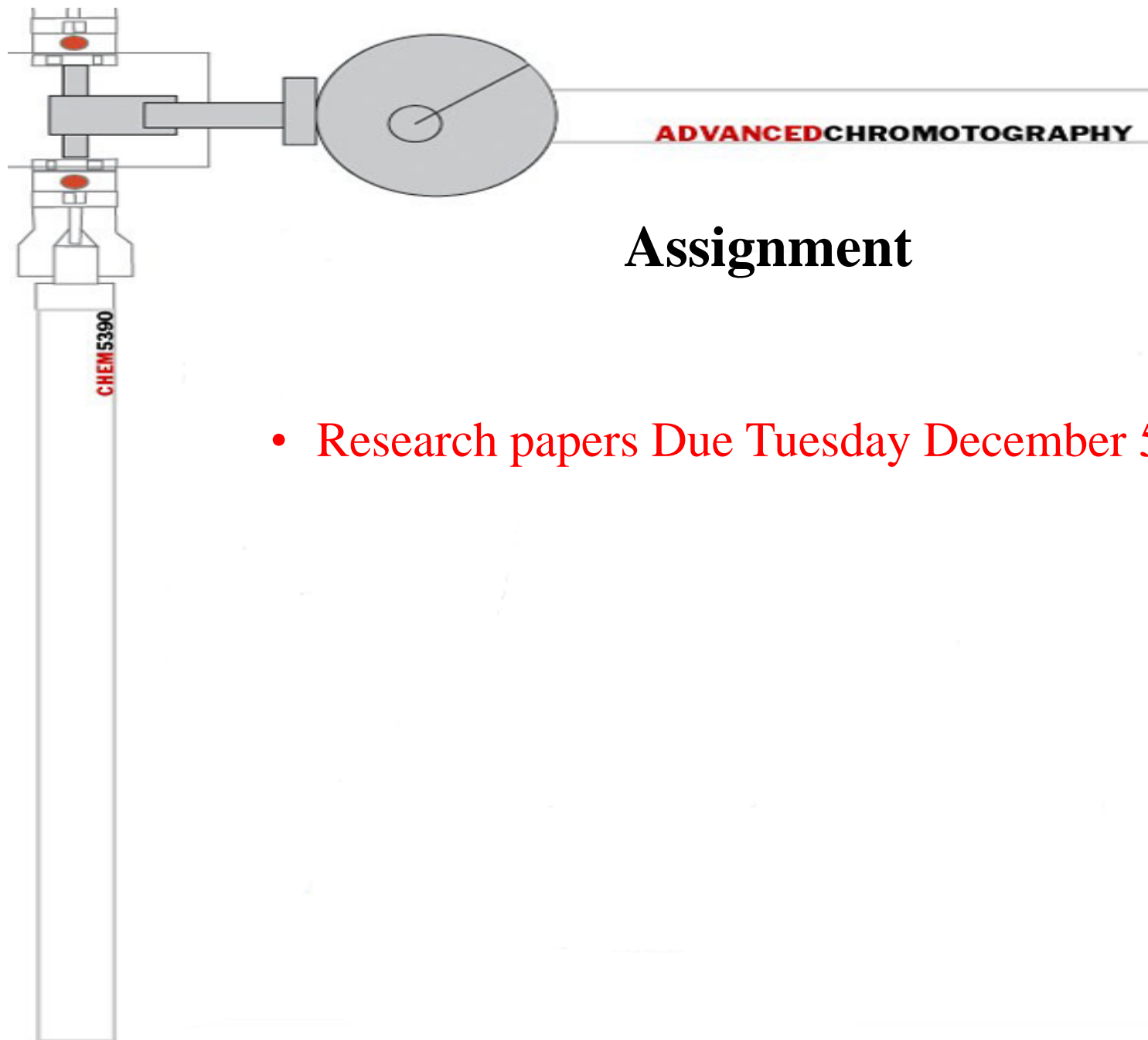
Characterization of proteins

Used to separate, purify, and characterize.

# Gel Permeation Chromatography



**Fig. 6.26.** Gel filtration chromatography of proteins. Waters Protein-Pak 300 SW columns ( $\times 2$ ) were used with 0.1 M potassium phosphate monobasic as the mobile phase at  $1.0 \text{ ml min}^{-1}$ . Detection was by direct u.v. at 280 nm. Solute identities: 1, blue dextran; 2, ferritin; 3, bovine serum albumin; 4, ovalbumin; 5, trypsin inhibitor; 6, ribonuclease A; 7, guanosine. Reprinted from *Waters Sourcebook of Chromatography* (1992), Millipore, Milford, with permission.



## Assignment

- Research papers Due Tuesday December 5<sup>th</sup>